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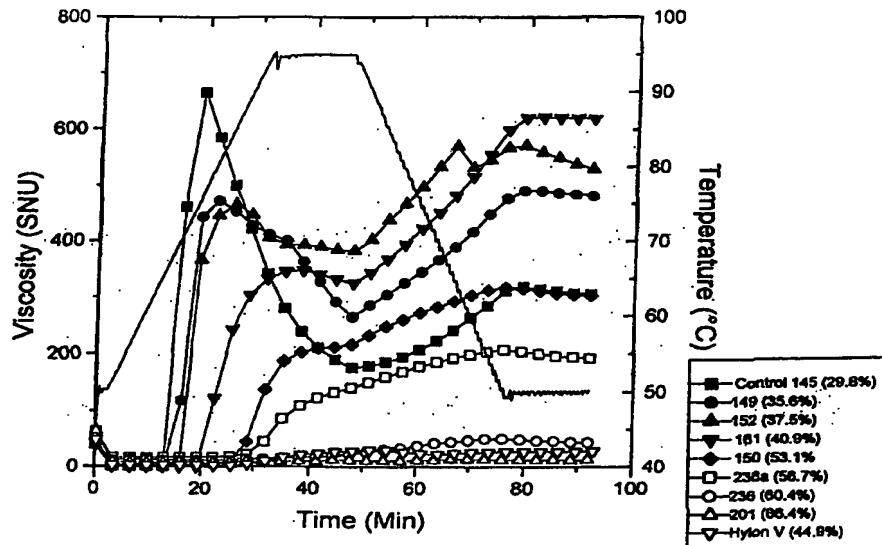
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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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(30) Priority Data:	9509229.2 9607409.1	5 May 1995 (05.05.95) 10 April 1996 (10.04.96)	GB GB	[GB/GB]; 46 Castle Street, Wellingborough, Northamptonshire NN8 1LW (GB).
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(72) Inventors; and				(81) Designated States: AU, BR, CA, JP, KR, US, European patent (AT, BE, CH, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE).
(75) Inventors/Applicants (for US only):	COOKE, David [GB/GB]; 2 Hunts Path, Oakley, Bedfordshire MK43 7SR (GB). DEBET, Martine [FR/GB]; 12 Bentley Close, Rectory Farm, Northampton NN3 5JS (GB). GIDLEY, Michael, John [GB/GB]; 45 Holmes Avenue, Raunds, Northamptonshire NN9 6SZ (GB). JOBLING, Stephen, Alan [GB/GB]; 19 Burwell Road, Eaton Socon, Huntingdon PE19 3QQ (GB). SAFFORD, Richard [GB/GB]; 10 Furness Close, Bedford, Bedfordshire MK41 8RN (GB). SIDEBOTTOM, Christopher, Michael [GB/GB]; 35 Waterloo Road, Bedford, Bedfordshire MK40 3PQ (GB). WESTCOTT, Roger, John			Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
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(54) Title: IMPROVEMENTS IN OR RELATING TO PLANT STARCH COMPOSITION



(57) Abstract

Disclosed is a nucleotide sequence encoding an effective portion of a class A starch branching enzyme (SBE) obtainable from potato plants, or a functional equivalent thereof, together with, inter alia, a corresponding polypeptide, a method of altering the characteristics of a plant, a plant having altered characteristics; and starch, particularly starch obtained from a potato plant, having novel properties.

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INTERNATIONAL SEARCH REPORT

International Application No
PCT/GB 96/01075

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12N15/82 C12N15/54 C08B30/04 A01H5/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 C12N C08B A01H

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	THE PLANT JOURNAL, vol. 7, no. 1, January 1995, pages 3-15, XP002014042 BURTON, R.A., ET AL.: "Starch branching enzymes belonging to distinct enzyme families are differentially expressed during pea embryo development" see the whole document ---	28-32, 34,39-41
X	DATABASE WPI Section Ch, Week 9442 Derwent Publications Ltd., London, GB; Class C06, AN 94-337418 XP002014047 & JP,A,06 261 767 (MITSUI GYOSAI SHOKUBUTSU BIO KENKYUSHO), 20 September 1994 see abstract ---	28-37, 39-42, 48-50

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

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International Application No
PCT/GB 96/01075

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X	WO,A,95 07355 (INST GENBIOLOGISCHE FORSCHUNG ;KOSSMANN JENS (DE); VIRGIN IVAR (DE) 16 March 1995 see the whole document ---	64,65
X	WO,A,92 11375 (AMYLOGENE HB) 9 July 1992 see the whole document ---	64,65
P,X	WO,A,95 26407 (NAT STARCH CHEM INVEST ;COOKE DAVID (GB); GIDLEY MICHAEL JOHN (GB)) 5 October 1995 see the whole document ---	1-26, 53-68
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A	PLANT POLYMERIC CARBOHYDRATES. INTERNATIONAL SYMPOSIUM HELD BERLIN, JULY 1-3, 1993, pages 33-39, XP002014045 WILLMITZER, L., ET AL.: "Starch synthesis in transgenic plants" see page 38 ---	27-41
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INTERNATIONAL SEARCH REPORT

International Application No
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International Application No
PCT/GB 96/01075

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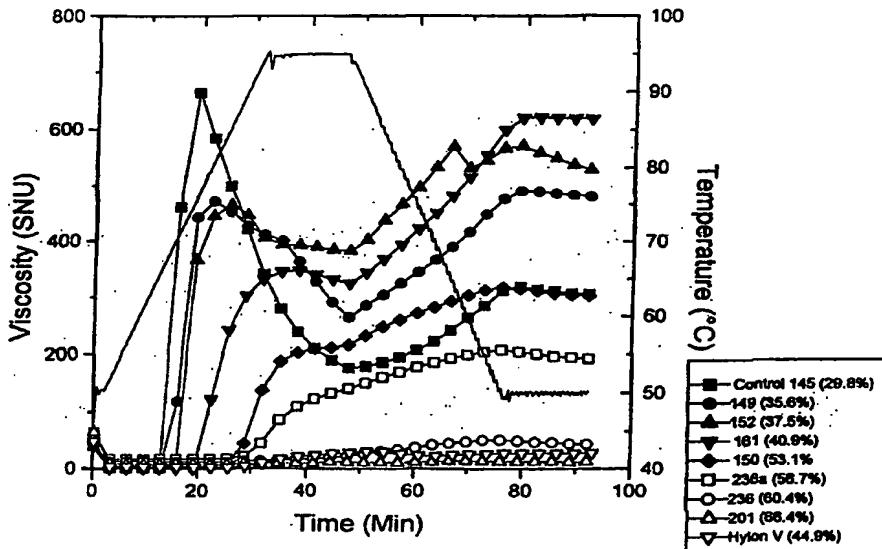
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(72) Inventors; and (75) Inventors/Applicants (for US only): COOKE, David [GB/GB]; 2 Hunts Path, Oakley, Bedfordshire MK43 7SR (GB). DEBET, Martine [FR/GB]; 12 Bentley Close, Rectory Farm, Northampton NN3 5JS (GB). GIDLEY, Michael, John [GB/GB]; 45 Holmes Avenue, Raunds, Northamptonshire NN9 6SZ (GB). JOBLING, Stephen, Alan [GB/GB]; 19 Burwell Road, Eaton Socon, Huntingdon PE19 3QQ (GB). SAFFORD, Richard [GB/GB]; 10 Furness Close, Bedford, Bedfordshire MK41 8RN (GB). SIDEBOTTOM, Christopher, Michael [GB/GB]; 35 Waterloo Road, Bedford, Bedfordshire MK40 3PQ (GB). WESTCOTT, Roger, John		

(54) Title: IMPROVEMENTS IN OR RELATING TO PLANT STARCH COMPOSITION



(57) Abstract

Disclosed is a nucleotide sequence encoding an effective portion of a class A starch branching enzyme (SBE) obtainable from potato plants, or a functional equivalent thereof, together with, inter alia, a corresponding polypeptide, a method of altering the characteristics of a plant, a plant having altered characteristics; and starch, particularly starch obtained from a potato plant, having novel properties.

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Title: Improvements in or Relating to Plant Starch Composition

Field of the Invention

This invention relates to novel nucleotide sequences, polypeptides encoded thereby, vectors and host cells and host organisms comprising one or more of the novel sequences, and to a method of altering one or more characteristics of an organism. The invention also relates to starch having novel properties and to uses thereof.

Background of the Invention

Starch is the major form of carbon reserve in plants, constituting 50% or more of the dry weight of many storage organs - e.g. tubers, seeds of cereals. Starch is used in numerous food and industrial applications. In many cases, however, it is necessary to modify the native starches, via chemical or physical means, in order to produce distinct properties to suit particular applications. It would be highly desirable to be able to produce starches with the required properties directly in the plant, thereby removing the need for additional modification. To achieve this via genetic engineering requires knowledge of the metabolic pathway of starch biosynthesis. This includes characterisation of genes and encoded gene products which catalyse the synthesis of starch. Knowledge about the regulation of starch biosynthesis raises the possibility of "re-programming" biosynthetic pathways to create starches with novel properties that could have new commercial applications.

The commercially useful properties of starch derive from the ability of the native granular form to swell and absorb water upon suitable treatment. Usually heat is required to cause granules to swell in a process known as gelatinisation, which has been defined (W A Atwell *et al*, Cereal Foods World 33, 306-311, 1988) as "... *the collapse (disruption) of molecular orders within the starch granule manifested in irreversible changes in properties such as granular swelling, native crystallite melting, loss of birefringence, and starch solubilisation*. *The point of initial gelatinisation and the range over which it occurs is governed by starch concentration, method of observation, granule type, and heterogeneities within the granule population under observation*". A number of techniques are available

for the determination of gelatinisation as induced by heating, a convenient and accurate method being differential scanning calorimetry, which detects the temperature range and enthalpy associated with the collapse of molecular orders within the granule. To obtain accurate and meaningful results, the peak and/or onset temperature of the endotherm observed by differential scanning calorimetry is usually determined.

The consequence of the collapse of molecular orders within starch granules is that the granules are capable of taking up water in a process known as pasting, which has been defined (W A Atwell *et al*, Cereal Foods World 33, 306-311, 1988) as "... *the phenomenon following gelatinisation in the dissolution of starch. It involves granular swelling, exudation of molecular components from the granule, and eventually, total disruption of the granules*". The best method of evaluating pasting properties is considered to be the viscoamylograph (Atwell *et al*, 1988 cited above) in which the viscosity of a stirred starch suspension is monitored under a defined time/temperature regime. A typical viscoamylograph profile for potato starch shows an initial rise in viscosity, which is considered to be due to granule swelling. In addition to the overall shape of the viscosity response in a viscoamylograph, a convenient quantitative measure is the temperature of initial viscosity development (onset). Figure 1 shows such a typical viscosity profile for potato starch, during and after cooking, and includes stages A-D which correspond to viscosity onset (A), maximum viscosity (B), complete dispersion (C) and reassociation of molecules (or retrogradation, D). In the figure, the dotted line represents viscosity (in stirring number units) of a 10% w/w starch suspension and the unbroken line shows the temperature in degrees centigrade. At a certain point, defined by the viscosity peak, granule swelling is so extensive that the resulting highly expanded structures are susceptible to mechanically-induced fragmentation under the stirring conditions used. With increased heating and holding at 95°C, further reduction in viscosity is observed due to increased fragmentation of swollen granules. This general profile has previously always been found for native potato starch.

After heating starches in water to 95°C and holding at that temperature (for typically 15 minutes), subsequent cooling to 50°C results in an increase in viscosity due to the process of retrogradation or set-back. Retrogradation (or set-back) is defined (Atwell *et al.*, 1988

cited above) as "...*a process which occurs when the molecules comprising gelatinised starch begin to reassociate in an ordered structure...*". At 50°C, it is primarily the amylose component which reassociates, as indicated by the increase in viscoamylograph viscosity for starch from normal maize (21.6% amylose) compared with starch from waxy maize (1.1% amylose) as shown in Figure 2. Figure 2 is a viscoamylograph of 10%w/w starch suspensions from waxy maize (solid line), conventional maize (dots and dashes), high amylose variety (hylon 5, dotted line) and a very high amylose variety (hylon 7, crosses). The temperature profile is also shown by a solid line, as in Figure 1. The extent of viscosity increase in the viscoamylograph on cooling and holding at 50°C depends on the amount of amylose which is able to reassociate due to its exudation from starch granules during the gelatinisation and pasting processes. A characteristic of amylose-rich starches from maize plants is that very little amylose is exuded from granules by gelatinisation and pasting up to 95°C, probably due to the restricted swelling of the granules. This is illustrated in Figure 2 which shows low viscosities for a high amylose (44.9%) starch (Hylon 5) from maize during gelatinisation and pasting at 95°C and little increase in viscosity on cooling and holding at 50°C. This effect is more extreme for a higher amylose content (58%, as in Hylon 7), which shows even lower viscosities in the viscoamylograph test (Figure 2). For commercially-available high amylose starches (currently available from maize plants, such as those described above), processing at greater than 100°C is usually necessary in order to generate the benefits of high amylose contents with respect to increased rates and strengths of reassociation, but use of such high temperatures is energetically unfavourable and costly. Accordingly, there is an unmet need for starches of high amylose content which can be processed below 100°C and still show enhanced levels of reassociation, as indicated for example by viscoamylograph measurements.

The properties of potato starch are useful in a variety of both food and non-food (paper, textiles, adhesives etc.) applications. However, for many applications, properties are not optimum and various chemical and physical modifications well known in the art are undertaken in order to improve useful properties. Two types of property manipulation which would be of use are: the controlled alteration of gelatinisation and pasting temperatures; and starches which suffer less granular fragmentation during pasting than

conventional starches.

Currently the only ways of manipulating the gelatinisation and pasting temperatures of potato starch are by the inclusion of additives such as sugars, polyhydroxy compounds or salts (Evans & Haisman, Starke 34, 224-231, 1982) or by extensive physical or chemical pre-treatments (e.g. Stute, Starke 44, 205-214, 1992). The reduction of granule fragmentation during pasting can be achieved either by extensive physical pretreatments (Stute, Starke 44, 205-214, 1992) or by chemical cross-linking. Such processes are inconvenient and inefficient. It is therefore desirable to obtain plants which produce starch which intrinsically possesses such advantageous properties.

Starch consists of two main polysaccharides, amylose and amylopectin. Amylose is a generally linear polymer containing α -1,4 linked glucose units, while amylopectin is a highly branched polymer consisting of a α -1,4 linked glucan backbone with α -1,6 linked glucan branches. In most plant storage reserves amylopectin constitutes about 75% of the starch content. Amylopectin is synthesized by the concerted action of soluble starch synthase and starch branching enzyme [α -1,4 glucan: α -1,4 glucan 6-glycosyltransferase, EC 2.4.1.18]. Starch branching enzyme (SBE) hydrolyses α -1,4 linkages and rejoins the cleaved glucan, via an α -1,6 linkage, to an acceptor chain to produce a branched structure. The physical properties of starch are strongly affected by the relative abundance of amylose and amylopectin, and SBE is therefore a crucial enzyme in determining both the quantity and quality of starches produced in plant systems.

In most plants studied to date e.g. maize (Boyer & Preiss, 1978 Biochem. Biophys. Res. Comm. 80, 169-175), rice (Smyth, 1988 Plant Sci. 57, 1-8) and pea (Smith, Planta 175, 270-279), two forms of SBE have been identified, each encoded by a separate gene. A recent review by Burton *et al.*, (1995 The Plant Journal 7, 3-15) has demonstrated that the two forms of SBE constitute distinct classes of the enzyme such that, in general, enzymes of the same class from different plants may exhibit greater similarity than enzymes of different classes from the same plant. In their review, Burton *et al.* termed the two respective enzyme families class "A" and class "B", and the reader is referred thereto (and to the references cited therein) for a detailed discussion of the distinctions

between the two classes. One general distinction of note would appear to be the presence, in class A SBE molecules, of a flexible N-terminal domain, which is not found in class B molecules. The distinctions noted by Burton *et al.* are relied on herein to define class A and class B SBE molecules, which terms are to be interpreted accordingly.

However in potato, only one isoform of the SBE molecule (belonging to class B) has thus far been reported and only one gene cloned (Blennow & Johansson, 1991 *Phytochem.* **30**, 437-444, and Koßmann *et al.*, 1991 *Mol. Gen. Genet.* **230**, 39-44). Further, published attempts to modify the properties of starch in potato plants (by preventing expression of the single known SBE) have generally not succeeded (e.g. Müller-Röber & Koßmann 1994 *Plant Cell and Environment* **17**, 601-613).

Summary of the Invention

In a first aspect the invention provides a nucleotide sequence encoding an effective portion of a class A starch branching enzyme (SBE) obtainable from potato plants.

Preferably the nucleotide sequence encodes a polypeptide comprising an effective portion of the amino acid sequence shown in Figure 5 (excluding the sequence MNKRIDL, which does not represent part of the SBE molecule), or a functional equivalent thereof (which term is discussed below). The amino acid sequence shown in Figure 5 (Seq ID No. 15) includes a leader sequence which directs the polypeptide, when synthesised in potato cells, to the amyloplast. Those skilled in the art will recognise that the leader sequence is removed to produce a mature enzyme and that the leader sequence is therefore not essential for enzyme activity. Accordingly, an "effective portion" of the polypeptide is one which possesses sufficient SBE activity to complement the branching enzyme mutation in *E. coli* KV 832 cells (described below) and which is active when expressed in *E. coli* in the phosphorylation stimulation assay. An example of an incomplete polypeptide which nevertheless constitutes an "effective portion" is the mature enzyme lacking the leader sequence. By analogy with the pea class A SBE sequence, the potato class A sequence shown in Figure 5 probably possesses a leader sequence of about 48 amino acid residues, such that the N terminal amino acid sequence is thought to commence around the glutamic acid residue (E) at position 49 (EKSSYN... etc.). Those skilled in the art will appreciate

that an effective portion of the enzyme may well omit other parts of the sequence shown in the figure without substantial detrimental effect. For example, the C-terminal glutamic acid-rich region could be reduced in length, or possibly deleted entirely, without abolishing class A SBE activity. A comparison with other known SBE sequences, especially other class A SBE sequences (see for example, Burton *et al.*, 1995 cited above), should indicate those portions which are highly conserved (and thus likely to be essential for activity) and those portions which are less well conserved (and thus are more likely to tolerate sequence changes without substantial loss of enzyme activity).

Conveniently the nucleotide sequence will comprise substantially nucleotides 289 to 2790 of the DNA sequence (Seq ID No. 14) shown in Figure 5 (which nucleotides encode the mature enzyme) or a functional equivalent thereof, and may also include further nucleotides at the 5' or 3' end. For example, for ease of expression, the sequence will desirably also comprise an in-frame ATG start codon, and may also encode a leader sequence. Thus, in one embodiment, the sequence further comprises nucleotides 145 to 288 of the sequence shown in Figure 5. Other embodiments are nucleotides 228 to 2855 of the sequence labelled "psbe2con.seq" in Figure 8, and nucleotides 57 to 2564 of the sequence shown in Figure 12 (preferably comprising an in-frame ATG start codon, such as the sequence of nucleotides 24 to 56 in the same Figure), or functional equivalents of the aforesaid sequences.

The term "functional equivalent" as applied herein to nucleotide sequences is intended to encompass those sequences which differ in their nucleotide composition to that shown in Figure 5 but which, by virtue of the degeneracy of the genetic code, encode polypeptides having identical or substantially identical amino acid sequences. It is intended that the term should also apply to sequences which are sufficiently homologous to the sequence of the invention that they can hybridise to the complement thereof under stringent hybridisation conditions - such equivalents will preferably possess at least 85%, more preferably at least 90%, and most preferably at least 95% sequence homology with the sequence of the invention as exemplified by nucleotides 289 to 2790 of the DNA sequence shown in Figure 5. It will be apparent to those skilled in the art that the nucleotide sequence of the invention may also find useful application when present as an "antisense"

sequence. Accordingly, functionally equivalent sequences will also include those sequences which can hybridise, under stringent hybridisation conditions, to the sequence of the invention (rather than the complement thereof). Such "antisense" equivalents will preferably possess at least 85%, more preferably at least 90%, and most preferably 95% sequence homology with the complement of the sequence of the invention as exemplified by nucleotides 289 to 2790 of the DNA sequence shown in Figure 5. Particular functional equivalents are shown, for example, in Figures 8 and 10 (if one disregards the various frameshift mutations noted therein).

The invention also provides vectors, particularly expression vectors, comprising the nucleotide sequence of the invention. The vector will typically comprise a promoter and one or more regulatory signals of the type well known to those skilled in the art. The invention also includes provision of cells transformed (which term encompasses transduction and transfection) with a vector comprising the nucleotide sequence of the invention.

The invention further provides a class A SBE polypeptide, obtainable from potato plants. In particular the invention provides the polypeptide in substantially pure form, especially in a form free from other plant-derived (especially potato plant-derived) components, which can be readily accomplished by expression of the relevant nucleotide sequence in a suitable non-plant host (such as any one of the yeast strains routinely used for expression purposes, e.g. *Pichia spp.* or *Saccharomyces spp.*). Typically the enzyme will substantially comprise the sequence of amino acid residues 49 to 882 shown in Figure 5 (disregarding the sequence MNKRIDL, which is not part of the enzyme), or a functional equivalent thereof. The polypeptide of the invention may be used in a method of modifying starch *in vitro*, comprising treating starch under suitable conditions (e.g. appropriate temperature, pH, etc) with an effective amount of the polypeptide according to the invention.

The term "functional equivalent", as applied herein to amino acid sequences, is intended to encompass amino acid sequences substantially similar to that shown in Figure 5, such that the polypeptide possesses sufficient activity to complement the branching enzyme mutation in *E. coli* KV 832 cells (described below) and which is active in *E. coli* in the

phosphorylation stimulation assay. Typically such functionally equivalent amino acid sequences will preferably possess at least 85%, more preferably at least 90%, and most preferably at least 95% sequence identity with the amino acid sequence of the mature enzyme (i.e. minus leader sequence) shown in Figure 5. Those skilled in the art will appreciate that conservative substitutions may be made generally throughout the molecule without substantially affecting the activity of the enzyme. Moreover, some non-conservative substitutions may be tolerated, especially in the less highly conserved regions of the molecule. Such substitutions may be made, for example, to modify slightly the activity of the enzyme. The polypeptide may, if desired, include a leader sequence, such as that exemplified by residues 1 to 48 of the amino acid sequence shown in Figure 5, although other leader sequences and signal peptides and the like are known and may be included.

A portion of the nucleotide sequence of the invention has been introduced into a plant and found to affect the characteristics of the plant. In particular, introduction of the sequence of the invention, operably linked in the antisense orientation to a suitable promoter, was found to reduce the amount of branched starch molecules in the plant. Additionally, it has recently been demonstrated in other experimental systems that "sense suppression" can also occur (i.e. expression of an introduced sequence operably linked in the sense orientation can interfere, by some unknown mechanism, with the expression of the native gene), as described by Matzke & Matzke (1995 *Plant Physiol.* **107**, 679-685). Any one of the methods mentioned by Matzke & Matzke could, in theory, be used to affect the expression in a host of a homologous SBE gene.

It is believed that antisense methods are mainly operable by the production of antisense mRNA which hybridises to the sense mRNA, preventing its translation into functional polypeptide, possibly by causing the hybrid RNA to be degraded (e.g. Sheehy *et al.*, 1988 *PNAS* **85**, 8805-8809; Van der Krol *et al.*, *Mol. Gen. Genet.* **220**, 204-212). Sense suppression also requires homology between the introduced sequence and the target gene, but the exact mechanism is unclear. It is apparent however that, in relation to both antisense and sense suppression, neither a full length nucleotide sequence, nor a "native" sequence is essential. Preferably the "effective portion" used in the method will comprise

at least one third of the full length sequence, but by simple trial and error other fragments (smaller or larger) may be found which are functional in altering the characteristics of the plant.

Thus, in a further aspect the invention provides a method of altering the characteristics of a plant, comprising introducing into the plant an effective portion of the sequence of the invention operably linked to a suitable promoter active in the plant. Conveniently the sequence will be linked in the anti-sense orientation to the promoter. Preferably the plant is a potato plant. Conveniently, the characteristic altered relates to the starch content and/or starch composition of the plant (i.e. amount and/or type of starch present in the plant). Preferably the method of altering the characteristics of the plant will also comprise the introduction of one or more further sequences, in addition to an effective portion of the sequence of the invention. The introduced sequence of the invention and the one or more further sequences (which may be sense or antisense sequences) may be operably linked to a single promoter (which would ensure both sequences were transcribed at essentially the same time), or may be operably linked to separate promoters (which may be necessary for optimal expression). Where separate promoters are employed they may be identical to each other or different. Suitable promoters are well known to those skilled in the art and include both constitutive and inducible types. Examples include the CaMV 35S promoter (e.g. single or tandem repeat) and the patatin promoter. Advantageously the promoter will be tissue-specific. Desirably the promoter will cause expression of the operably linked sequence at substantial levels only in the tissue of the plant where starch synthesis and/or starch storage mainly occurs. Thus, for example, where the sequence is introduced into a potato plant, the operably linked promoter may be tuber-specific, such as the patatin promoter.

Desirably, for example, the method will also comprise the introduction of an effective portion of a sequence encoding a class B SBE, operably linked in the antisense orientation to a suitable promoter active in the plant. Desirably the further sequence will comprise an effective portion of the sequence encoding the potato class B SBE molecule. Conveniently the further sequence will comprise an effective portion of the sequence described by Blennow & Johansson (1991 Phytochem. 30, 437-444) or that disclosed in

WO92/11375. More preferably, the further sequence will comprise at least an effective portion of the sequence disclosed in International Patent Application No. WO 95/26407. Use of antisense sequences against both class A and class B SBE in combination has now been found by the present inventors to result in the production of starch having very greatly altered properties (see below). Those skilled in the art will appreciate the possibility that, if the plant already comprises a sense or antisense sequence which efficiently inhibits the class B SBE activity, introduction of a sense or antisense sequence to inhibit class A SBE activity (thereby producing a plant with inhibition of both class A and class B activity) might alter greatly the properties of the starch in the plant, without the need for introduction of one or more further sequences. Thus the sequence of the invention is conveniently introduced into plants already having low levels of class A and/or class B SBE activity, such that the inhibition resulting from the introduction of the sequence of the invention is likely to have a more pronounced effect.

The sequence of the invention, and the one or more further sequences if desired, can be introduced into the plant by any one of a number of well-known techniques (e.g. Agrobacterium-mediated transformation, or by "biolistic" methods). The sequences are likely to be most effective in inhibiting SBE activity in potato plants, but theoretically could be introduced into any plant. Desirable examples include pea, tomato, maize, wheat, rice, barley, sweet potato and cassava plants. Preferably the plant will comprise a natural gene encoding an SBE molecule which exhibits reasonable homology with the introduced nucleic acid sequence of the invention.

In another aspect, the invention provides a plant cell, or a plant or the progeny thereof, which has been altered by the method defined above. The progeny of the altered plant may be obtained, for example, by vegetative propagation, or by crossing the altered plant and reserving the seed so obtained. The invention also provides parts of the altered plant, such as storage organs. Conveniently, for example, the invention provides tubers comprising altered starch, said tubers being obtained from an altered plant or the progeny thereof. Potato tubers obtained from altered plants (or the progeny thereof) will be particularly useful materials in certain industrial applications and for the preparation and/or processing of foodstuffs and may be used, for example, to prepare low-fat waffles and

chips (amylose generally being used as a coating to prevent fat uptake), and to prepare mashed potato (especially "instant" mashed potato) having particular characteristics.

In particular relation to potato plants, the invention provides a potato plant or part thereof which, in its wild type possesses an effective SBE A gene, but which plant has been altered such that there is no effective expression of an SBE A polypeptide within the cells of at least part of the plant. The plant may have been altered by the method defined above, or may have been selected by conventional breeding to be deleted for the class A SBE gene, presence or absence of which can be readily determined by screening samples of the plants with a nucleic acid probe or antibody specific for the potato class A gene or gene product respectively.

The invention also provides starch extracted from a plant altered by the method defined above, or the progeny of such a plant, the starch having altered properties compared to starch extracted from equivalent, but unaltered, plants. The invention further provides a method of making altered starch, comprising altering a plant by the method defined above and extracting therefrom starch having altered properties compared to starch extracted from equivalent, but unaltered, plants. Use of nucleotide sequences in accordance with the invention has allowed the present inventors to produce potato starches having a wide variety of novel properties.

In particular the invention provides the following: a plant (especially a potato plant) altered by the method defined above, containing starch which, when extracted from the plant, has an elevated endotherm peak temperature as judged by DSC, compared to starch extracted from a similar, but unaltered, plant; a plant (especially a potato plant) altered by the method defined above, containing starch which, when extracted from the plant, has an elevated viscosity onset temperature (conveniently elevated by 10 - 25°C) as judged by viscoamylograph compared to starch extracted from a similar, but unaltered, plant; a plant (especially a potato plant) altered by the method defined above, containing starch which, when extracted from the plant, has a decreased peak viscosity (conveniently decreased by 240 - 700SNUs) as judged by viscoamylograph compared to starch extracted from a similar, but unaltered, plant; a plant (especially a potato plant) altered by the method

defined above, containing starch which, when extracted from the plant, has an increased pasting viscosity (conveniently increased by 37 - 260SNUs) as judged by viscoamylograph compared to starch extracted from a similar, but unaltered, plant; a plant (especially a potato plant) altered by the method defined above, containing starch which, when extracted from the plant, has an increased set-back viscosity (conveniently increased by 224 - 313 SNUs) as judged by viscoamylograph compared to starch extracted from a similar, but unaltered, plant; a plant (especially a potato plant) altered by the method defined above, containing starch which, when extracted from the plant, has a decreased set-back viscosity as judged by viscoamylograph compared to starch extracted from a similar, but unaltered, plant; and a plant (especially a potato plant) altered by the method defined above, containing starch which, when extracted from the plant, has an elevated amylose content as judged by iodometric assay (i.e. by the method of Morrison & Laignelet 1983, cited above) compared to starch extracted from a similar, but unaltered, plant. The invention also provides for starch obtainable or obtained from such plants as aforesaid.

In particular the invention provides for starch which, as extracted from a potato plant by wet milling at ambient temperature, has one or more of the following properties, as judged by viscoamylograph analysis performed according to the conditions defined below: viscosity onset temperature in the range 70-95°C (preferably 75-95°C); peak viscosity in the range 500 - 12 stirring number units; pasting viscosity in the range 214 - 434 stirring number units; set-back viscosity in the range 450 - 618 or 14 - 192 stirring number units; or displays no significant increase in viscosity during viscoamylograph. Peak, pasting and set-back viscosities are defined below. Viscosity onset temperature is the temperature at which there is a sudden, marked increase in viscosity from baseline levels during viscoamylograph, and is a term well-known to those skilled in the art.

In other particular embodiments, the invention provides starch which as extracted from a potato plant by wet milling at ambient temperature has a peak viscosity in the range 200 - 500 SNUs and a set-back viscosity in the range 275-618 SNUs as judged by viscoamylograph according to the protocol defined below; and starch which as extracted from a potato plant by wet milling at ambient temperature has a viscosity which does not decrease between the start of the heating phase (step 2) and the start of the final holding

phase (step 5) and has a set-back viscosity of 303 SNUs or less as judged by viscoamylograph according to the protocol defined below.

For the purposes of the present invention, viscoamylograph conditions are understood to pertain to analysis of a 10% (w/w) aqueous suspension of starch at atmospheric pressure, using a Newport Scientific Rapid Visco Analyser with a heating profile of: holding at 50°C for 2 minutes (step 1), heating from 50 to 95°C at a rate of 1.5°C per minute (step 2), holding at 95°C for 15 minutes (step 3), cooling from 95 to 50°C at a rate of 1.5°C per minute (step 4), and then holding at 50°C for 15 minutes (step 5). Peak viscosity may be defined for present purposes as the maximum viscosity attained during the heating phase (step 2) or the holding phase (step 3) of the viscoamylograph. Pasting viscosity may be defined as the viscosity attained by the starch suspensions at the end of the holding phase (step 3) of the viscoamylograph. Set-back viscosity may be defined as the viscosity of the starch suspension at the end of step 5 of the viscoamylograph.

In yet another aspect the invention provides starch from a potato plant having an apparent amylose content (% w/w) of at least 35%, as judged by iodometric assay according to the method described by Morrison & Laignelet (1983 J. Cereal Science 1, 9-20). Preferably the starch will have an amylose content of at least 40%, more preferably at least 50%, and most preferably at least 66%. Starch obtained directly from a potato plant and having such properties has not hitherto been produced. Indeed, as a result of the present invention, it is now possible to generate *in vivo* potato starch which has some properties analogous to the very high amylose starches (e.g. Hylon 7) obtainable from maize.

Starches with high (at least 35%) amylose contents find commercial application as, amongst other reasons, the amylose component of starch reassociates more strongly and rapidly than the amylopectin component during retrogradation processes. This may result, for example, in pastes with higher viscosities, gels of greater cohesion, or films of greater strength for starches with high (at least 35%) compared with normal (less than 35%) amylose contents. Alternatively, starches may be obtained with very high amylose contents, such that the granule structure is substantially preserved during heating, resulting in starch suspensions which demonstrate substantially no increase in viscosity during

cooking (i.e. there is no significant viscosity increase during viscoamylograph conditions defined above). Such starches typically exhibit a viscosity increase of less than 10% (preferably less than 5%) during viscoamylograph under the conditions defined above.

In commerce, these valuable properties are currently obtained from starches of high amylose content derived from maize plants. It would be of commercial value to have an alternative source of high amylose starches from potato as other characteristics such as granule size, organoleptic properties and textural qualities may distinguish application performances of high amylose starches from maize and potato plants.

Thus high amylose starch obtained by the method of the present invention may find application in many different technological fields, which may be broadly categorised into two groups: food products and processing; and "Industrial" applications. Under the heading of food products, the novel starches of the present invention may find application as, for example, films, barriers, coatings or gelling agents. In general, high amylose content starches absorb less fat during frying than starches with low amylose content, thus the high amylose content starches of the invention may be advantageously used in preparing low fat fried products (e.g. potato chips, crisps and the like). The novel starches may also be employed with advantage in preparing confectionery and in granular and retrograded "resistant" starches. "Resistant" starch is starch which is resistant to digestion by α -amylase. As such, resistant starch is not digested by α -amylases present in the human small intestine, but passes into the colon where it exhibits properties similar to soluble and insoluble dietary fibre. Resistant starch is thus of great benefit in foodstuffs due to its low calorific value and its high dietary fibre content. Resistant starch is formed by the retrogradation (akin to recrystallization) of amylose from starch gels. Such retrogradation is inhibited by amylopectin. Accordingly, the high amylose starches of the present invention are excellent starting materials for the preparation of resistant starch. Suitable methods for the preparation of resistant starch are well-known to those skilled in the art and include, for example, those described in US 5,051,271 and US 5,281,276. Conveniently the resistant starches provided by the present invention comprise at least 5% total dietary fibre, as judged by the method of Prosby *et al.*, (1985 J. Assoc. Off. Anal. Chem. 68, 677), mentioned in US 5,281, 276.

Under the heading of "Industrial" applications, the novel starches of the invention may be advantageously employed, for example, in corrugating adhesives, in biodegradable products such as loose fill packaging and foamed shapes, and in the production of glass fibers and textiles.

Those skilled in the art will appreciate that the novel starches of the invention may, if desired, be subjected *in vitro* to conventional enzymatic, physical and/or chemical modification, such as cross-linking, introduction of hydrophobic groups (e.g. octenyl succinic acid, dodecyl succinic acid), or derivatization (e.g. by means of esterification or etherification).

In yet another aspect the invention provides high (35% or more) amylose starches which generate paste viscosities greater than those obtained from high amylose starches from maize plants after processing at temperatures below 100°C. This provides the advantage of more economical starch gelatinisation and pasting treatments through the use of lower processing temperatures than are currently required for high amylose starches from maize plants.

The invention will now be further described by way of illustrative example and with reference to the drawings, of which:

Figure 1 shows a typical viscoamylograph for a 10% w/w suspension of potato starch;

Figure 2 shows viscoamylographs for 10% suspensions of starch from various maize varieties;

Figure 3 is a schematic representation of the cloning strategy used by the present inventors;

Figure 4a shows the amino acid alignment of the C-terminal portion of starch branching enzyme isoforms from various sources: amino acid residues matching the consensus

sequence are shaded;

Figure 4b shows the alignment of DNA sequences of various starch branching enzyme isoforms which encode a conserved amino acid sequence;

Figure 5 shows the DNA sequence (Seq ID No. 14) and predicted amino acid sequence (Seq ID No. 15) of a full length potato class A SBE cDNA clone obtained by PCR;

Figure 6 shows a comparison of the most highly conserved part of the amino acid sequences of potato class A (uppermost sequence) and class B (lowermost sequence) SBE molecules;

Figure 7 shows a comparison of the amino acid sequence of the full length potato class A (uppermost sequence) and pea (lowermost sequence) class A SBE molecules;

Figure 8 shows a DNA alignment of various full length potato class A SBE clones obtained by the inventors;

Figure 9 shows the DNA sequence of a potato class A SBE clone determined by direct sequencing of PCR products, together with the predicted amino acid sequence;

Figure 10 is a multiple DNA alignment of various full length potato SBE A clones obtained by the inventors;

Figure 11 is a schematic illustration of the plasmid pSJ64;

Figure 12 shows the DNA sequence and predicted amino acid sequence of the full length potato class A SBE clone as present in the plasmid pSJ90; and

Figure 13 shows viscoamylographs for 10% w/w suspensions of starch from various transgenic potato plants made by the relevant method aspect of the invention.

Examples

Example 1

Cloning of Potato class A SBE

The strategy for cloning the second form of starch branching enzyme from potato is shown in Figure 3. The small arrowheads represent primers used by the inventors in PCR and RACE protocols. The approximate size of the fragments isolated is indicated by the numerals on the right of the Figure. By way of explanation, a comparison of the amino acid sequences of several cloned plant starch branching enzymes (SBE) from maize (class A), pea (class A), maize (class B), rice (class B) and potato (class B), as well as human glycogen branching enzyme, allowed the inventors to identify a region in the carboxy-terminal one third of the protein which is almost completely conserved (GYLNFMGNEFGHPEWIDFPR) (Figure 4a). A multiple alignment of the DNA sequences (human, pea class A, potato class B, maize class B, maize class A and rice class B, respectively) corresponding to this region is shown in Figure 4b and was used to design an oligo which would potentially hybridize to all known plant starch branching enzymes: AATTT(C/T)ATGGGIAA(C/T)GA(A/G)TT(C/T)GG (Seq ID No. 20).

Library PCR

The initial isolation of a partial potato class A SBE cDNA clone was from an amplified potato tuber cDNA library in the λ Zap vector (Stratagene). One half μ L of a potato cDNA library (titre 2.3×10^9 pfu/mL) was used as template in a 50 μ L reaction containing 100 pmol of a 16 fold degenerate POTSBE primer and 25 pmol of a T7 primer (present in the λ Zap vector 3' to the cDNA sequences - see Figure 3), 100 μ M dNTPs, 2.5 U Taq polymerase and the buffer supplied with the Taq polymerase (Stratagene). All components except the enzyme were added to a 0.5 mL microcentrifuge tube, covered with mineral oil and incubated at 94°C for 7 minutes and then held at 55°C, while the Taq polymerase was added and mixed by pipetting. PCR was then performed by incubating for 1 min at 94°C, 1 min at 58°C and 3 minutes at 72°C, for 35 cycles. The PCR products were extracted with phenol/chloroform, ethanol precipitated and resuspended in TE pH 8.0 before cloning into the T/A cloning vector pT7BlueR (Invitrogen).

Several fragments between 600 and 1300 bp were amplified. These were isolated from an agarose gel and cloned into the pT7BlueR T/A cloning vector. Restriction mapping of 24 randomly selected clones showed that they belonged to several different groups (based on size and presence/absence of restriction sites). Initially four clones were chosen for sequencing. Of these four, two were found to correspond to the known potato class B SBE sequence, however the other two, although homologous, differed significantly and were more similar to the pea class A SBE sequence, suggesting that they belonged to the class A family of branching enzymes (Burton *et al.*, 1995 *The Plant Journal*, cited above). The latter two clones (~ 800bp) were sequenced fully. They both contained at the 5' end the sequence corresponding to the degenerate oligonucleotide used in the PCR and had a predicted open reading frame of 192 amino acids. The deduced amino acid sequence was highly homologous to that of the pea class A SBE.

The ~ 800 bp PCR derived cDNA fragment (corresponding to nucleotides 2281 to 3076 of the psbe2 con.seq sequence shown in Figure 8) was used as a probe to screen the potato tuber cDNA library. From one hundred and eighty thousand plaques, seven positives were obtained in the primary screen. PCR analysis showed that five of these clones were smaller than the original 800 bp cDNA clone, so these were not analysed further. The two other clones (designated 3.2.1 and 3.1.1) were approximately 1200 and 1500 bp in length respectively. These were sequenced from their 5' ends and the combined consensus sequence aligned with the sequence from the PCR generated clones. The cDNA clone 3.2.1 was excised from the phage vector and plasmid DNA was prepared and the insert fully sequenced. Several attempts to obtain longer clones from the library were unsuccessful, therefore clones containing the 5' end of the full length gene were obtained using RACE (rapid amplification of cDNA ends).

Rapid Amplification of cDNA ends (RACE) and PCR conditions

RACE was performed essentially according to Frohman (1992 *Amplifications* 11-15). Two µg of total RNA from mature potato tubers was heated to 65°C for 5 min and quick cooled on ice. The RNA was then reverse transcribed in a 20 µL reaction for 1 hour at 37°C using BRL's M-MLV reverse transcriptase and buffer with 1 mM DTT, 1 mM dNTPs, 1 U/µL RNAsin (Promega) and 500 pmol random hexamers (Pharmacia) as

primer. Excess primers were removed on a Centricon 100 column and cDNA was recovered and precipitated with isopropanol. cDNA was A-tailed in a volume of 20 μ l using 10 units terminal transferase (BRL), 200 μ M dATP for 10 min at 37°C, followed by 5 min at 65°C. The reaction was then diluted to 0.5 ml with TE pH 8 and stored at 4°C as the cDNA pool. cDNA clones were isolated by PCR amplification using the primers R_oR_idT₁₇, R_o and POTSBE24. The PCR was performed in 50 μ L using a hot start technique: 10 μ L of the cDNA pool was heated to 94°C in water for 5 min with 25 pmol POTSBE24, 25 pmol R_o and 2.5 pmol of R_oR_idT₁₇ and cooled to 75°C. Five μ L of 10 x PCR buffer (Stratagene), 200 μ M dNTPs and 1.25 units of Taq polymerase were added, the mixture heated at 45°C for 2 min and 72°C for 40 min followed by 35 cycles of 94°C for 45 sec, 50°C for 25 sec, 72°C for 1.5 min and a final incubation at 72°C for 10 min. PCR products were separated by electrophoresis on 1% low melting agarose gels and the smear covering the range 600-800 bp fragments was excised and used in a second PCR amplification with 25 pmol of R_i and POTSBE25 primers in a 50 μ L reaction (28 cycles of 94°C for 1 min, 50°C 1 min, 72°C 2 min). Products were purified by chloroform extraction and cloned into pT7 Blue. PCR was used to screen the colonies and the longest clones were sequenced.

The first round of RACE only extended the length of the SBE sequence approximately 100 bases, therefore a new A-tailed cDNA library was constructed using the class A SBE specific oligo POTSBE24 (10 pmol) in an attempt to recover longer RACE products. The first and second round PCR reactions were performed using new class A SBE primers (POTSBE 28 and 29 respectively) derived from the new sequence data. Conditions were as before except that the elongation step in the first PCR was for 3 min and the second PCR consisted of 28 cycles at 94 °C for 45 seconds, 55 °C for 25 sec and 72 °C for 1 min 45 sec.

Clones ranging in size from 400 bp to 1.4 kb were isolated and sequenced. The combined sequence of the longest RACE products and cDNA clones predicted a full length gene of about 3150 nucleotides, excluding the poly(A) tail (psbe 2con.seq in Fig. 8).

As the sequence of the 5' half of the gene was compiled from the sequence of several

RACE products generated using Taq polymerase, it was possible that the compiled sequence did not represent that of a single mRNA species and/or had nucleotide sequence changes. The 5' 1600 bases of the gene was therefore re-isolated by PCR using Ultma, a thermostable DNA polymerase which, because it possesses a 3'-5' exonuclease activity, has a lower error rate compared to Taq polymerase. Several PCR products were cloned and restriction mapped and found to differ in the number of *Hind* III, *Ssp* I, and *EcoR* I sites. These differences do not represent PCR artefacts as they were observed in clones obtained from independent PCR reactions (data not shown) and indicate that there are several forms of the class A SBE gene transcribed in potato tubers.

In order to ensure that the sequence of the full length cDNA clone was derived from a single mRNA species it was therefore necessary to PCR the entire gene in one piece. cDNA was prepared according to the RACE protocol except that the adaptor oligo R₉R₁dT₁₇ (5 pmol) was used as a primer and after synthesis the reaction was diluted to 200 μL with TE pH 8 and stored at 4°C. Two μL of the cDNA was used in a PCR reaction of 50 μL using 25 pmol of class A SBE specific primers PBER1 and PBERT (see below), and thirty cycles of 94° for 1 min, 60°C for 1 min and 72°C for 3 min. If Taq polymerase was used the PCR products were cloned into pT7Blue whereas if Ultma polymerase was used the PCR products were purified by chloroform extraction, ethanol precipitation and kinased in a volume of 20 μL (and then cloned into pBSSK IIP which had been cut with EcoRV and dephosphorylated). At least four classes of cDNA were isolated, which again differed in the presence or absence of *Hind* III, *Ssp* I and *EcoR* I sites. Three of these clones were sequenced fully, however one clone could not be isolated in sufficient quantity to sequence.

The sequence of one of the clones (number 19) is shown in Figure 5. The first methionine (initiation) codon starts a short open reading frame (ORF) of 7 amino acids which is out of frame with the next predicted ORF of 882 amino acids which has a molecular mass (Mr) of approximately 100 Kd. Nucleotides 6-2996 correspond to SBE sequence - the rest of the sequence shown is vector derived. Figure 6 shows a comparison of the most highly conserved part of the amino acid sequence of potato class A SBE (residues 180-871, top, row) and potato class B SBE (bottom row, residues 98-792); the middle row indicates the

degree of similarity, identical residues being denoted by the common letter, conservative changes by two dots and neutral changes by a single dot. Dashes indicate gaps introduced to optimise the alignment. The class A SBE protein has 44% identity over the entire length with potato class B SBE, and 56% identity therewith in the central conserved domain (Figure 6), as judged by the "Megalign" program (DNASTAR). However, Figure 7 shows a comparison between potato class A SBE (top row, residues 1-873) and pea class A SBE (bottom row, residues 1-861), from which it can be observed that cloned potato gene is more homologous to the class A pea enzyme, where the identity is 70 % over nearly the entire length, and this increases to 83 % over the central conserved region (starting at IPPP at position ~170). It is clear from this analysis that this cloned potato SBE gene belongs to the class A family of SBE genes.

An *E. coli* culture, containing the plasmid pSJ78 (which directs the expression of a full length potato SBE Class A gene), has been deposited (on 3rd January 1996) under the terms of the Budapest Treaty at The National Collections of Industrial and Marine Bacteria Limited (23 St Machar Drive, Aberdeen, AB2 1RY, United Kingdom), under accession number NCIMB 40781. Plasmid pSJ78 is equivalent to clone 19 described above. It represents a full length SBE A cDNA blunt-end ligated into the vector pBSSKIIIP.

Polymorphism of class A SBE genes

Sequence analysis of the other two full length class A SBE genes showed that they contain frameshift mutations and are therefore unable to encode full length proteins and indeed they were unable to complement the branching enzyme deficiency in the KV832 mutant (described below). An alignment of the full length DNA sequences is shown in Figure 8: "10con.seq" (Seq ID No. 12), "19con.seq" (Seq ID No. 14) and "11con.seq" (Seq ID No. 13) represent the sequence of full length clones 10, 19 and 11 obtained by PCR using the PBER1 and PBERT primers (see below), whilst "psbe2con.seq" (Seq ID No. 18) represents the consensus sequence of the RACE clones and cDNA clone 3.2.1. Those nucleotides which differ from the overall consensus sequence (not shown) are shaded. Dashes indicate gaps introduced to optimise the alignment. Apart from the frameshift mutations these clones are highly homologous. It should be noted that the 5' sequence of psbe2con is longer because this is the longest RACE product and it also contains several

changes compared to the other clones. The upstream methionine codon is still present in this clone but the upstream ORF is shortened to just 3 amino acids and in addition there is a 10 base deletion in the 5' untranslated leader.

The other significant area of variation is in the carboxy terminal region of the protein coding region. Closer examination of this area reveals a GAA trinucleotide repeat structure which varies in length between the four clones. These are typical characteristics of a microsatellite repeat region. The most divergent clone is #11 which has only one GAA triplet whereas clone 19 has eleven perfect repeats and the other two clones have five and seven GAA repeats. All of these deletions maintain the ORF but change the number of glutamic acid residues at the carboxy terminus of the protein.

Most of the other differences between the clones are single base changes. It is quite possible that some of these are PCR errors. To address this question direct sequencing of PCR fragments amplified from first strand cDNA was performed. Figure 9 shows the DNA sequence, and predicted amino acid sequence, obtained by such direct sequencing. Certain restriction sites are also marked. Nucleotides which could not be unambiguously assigned are indicated using standard IUPAC notation and, where this uncertainty affects the predicted amino acid sequence, a question mark is used. Sequence at the extreme 5' and 3' ends of the gene could not be determined because of the heterogeneity observed in the different cloned genes in these regions (see previous paragraph). However this can be taken as direct evidence that these differences are real and are not PCR or cloning artefacts.

There is absolutely no evidence for the frameshift mutations in the PCR derived sequence and it would appear that these mutations are an artefact of the cloning process, resulting from negative selection pressure in *E. coli*. This is supported by the fact that it proved extremely difficult to clone the full length PCR products intact as many large deletions were seen and the full length clones obtained were all cloned in one orientation (away from the LacZ promoter), perhaps suggesting that expression of the gene is toxic to the cells. Difficulties of this nature may have been responsible, at least in part, for the previous failure of other researchers to obtain the present invention.

A comparison of all the full length sequences is shown in Figure 10. In addition to clones 10, 11 and 19 are shown the sequences of a *Bgl* II - *Xho* I product cloned directly into the QE32 expression vector ("86CON.SEQ", Seq ID No. 16) and the consensus sequence of the directly sequenced PCR products ("pcrsbe2con.seq", Seq ID No. 17). Those nucleotides which differ from the consensus sequence (not shown) are shaded. Dashes indicate gaps introduced to optimise the alignment. There are 11 nucleotide differences predicted to be present in the mRNA population, which are indicated by asterisks above and below the sequence. The other differences are probably PCR artefacts or possibly sequencing errors.

Complementation of a branching enzyme deficient *E. coli* mutant

To determine if the isolated SBE gene encodes an active protein i.e. one that has branching enzyme activity, a complementation test was performed in the *E. coli* strain KV832. This strain is unable to make bacterial glycogen as the gene for the glycogen branching enzyme has been deleted (Keil *et al.*, 1987 Mol. Gen. Genet. 207, 294-301). When wild type cells are grown in the presence of glucose they synthesise glycogen (a highly branched glucose polymer) which stains a brown colour with iodine, whereas the KV832 cells make only a linear chain glucose polymer which stains blueish green with iodine. To determine if the cloned SBE gene could restore the ability of the KV832 cells to make a branched polymer, the clone pSJ90 (Seq ID No. 19) was used and constructed as below. The construct is a PCR-derived, substantially full length fragment (made using primers PBE 2B and PBE 2X, detailed below), which was cut with *Bgl* II and *Xho* I and cloned into the *Bam*H I / *Sal* I sites of the His-tag expression vector pQE32 (Qiagen). This clone, pSJ86, was sequenced and found to have a frameshift mutation of two bases in the 5' half of the gene. This frameshift was removed by digestion with *Nsi* I and *Sna*B I and replaced with the corresponding fragment from a Taq-generated PCR clone to produce the plasmid pSJ90 (sequence shown in Figure 12; the first 10 amino acids are derived from the expression vector). The polypeptide encoded by pSJ90 would be predicted to correspond to amino acids 46-882 of the full SBE coding sequence. The construct pSJ90 was transformed into the branching enzyme deficient KV832 cells and transformants were grown on solid PYG medium (0.85% KH₂PO₄, 1.1% K₂HPO₄, 0.6% yeast extract) containing 1.0% glucose. To test for complementation, a loop of cells was

scraped off and resuspended in 150 μ l of water, to which was added 15 μ l Lugol's solution (2g KI and 1g I₂ per 300ml water). It was found that the potato SBE fragment-transformed KV832 cells now stained a yellow-brown colour with iodine whereas control cells containing only the pQE32 vector continued to stain blue-green.

Expression of potato class A SBE in *E. coli*

Single colonies of KV832, containing one of the plasmids pQE32, pAGCR1 or pSJ90, were picked into 50ml of 2xYT medium containing carbenicillin, kanamycin and streptomycin as appropriate (100, 50 and 25 mg/L, respectively) in a 250ml flask and grown for 5 hours, with shaking, at 37°C. IPTG was then added to a final concentration of 1mM to induce expression and the flasks were further incubated overnight at 25°C. The cells were harvested by centrifugation and resuspended in 50 mM sodium phosphate buffer (pH 8.0), containing 300mM NaCl, 1mg/ml lysozyme and 1mM PMSF and left on ice for 1 hour. The cell lysates were then sonicated (3 pulses of 10 seconds at 40% power using a microprobe) and cleared by centrifugation at 12,000g for 10 minutes at 4°C. Cleared lysates were concentrated approximately 10 fold in a Centricon™ 30 filtration unit. Duplicate 10 μ l samples of the resulting extract were assayed for SBE activity by the phosphorylation stimulation method, as described in International Patent Application No. PCT/GB95/00634. In brief, the standard assay reaction mixture (0.2ml) was 200mM 2-(N-morpholino) ethanesulphonic acid (MES) buffer pH6.5, containing 100nCi of ¹⁴C glucose-1-phosphate at 50mM, 0.05 mg rabbit phosphorylase A, and *E. coli* lysate. The reaction mixture was incubated for 60 minutes at 30°C and the reaction terminated and glucan polymer precipitated by the addition of 1ml of 75% (v/v) methanol, 1% (w/v) potassium hydroxide, and then 0.1ml glycogen (10mg/ml). The results are presented below:

Construct	SBE Activity (cpm)
pQE32 (control)	1,829
pSJ90 (potato class A SBE)	14,327
pAGCR1 (pea class A SBE)	29,707

The potato class A SBE activity is 7-8 fold above background levels. It was concluded therefore that the potato class A SBE gene was able to complement the BE mutation in the

phosphorylation stimulation assay and that the cloned gene does indeed code for a protein with branching enzyme activity.

Oligonucleotides

The following synthetic oligonucleotides (Seq ID No.s 1-11 respectively) were used:

R _o R _{dT} ₁₇	AAGGATCCGTCGACATCGATAATACGACTCACTATAGGGA(T) ₁₇
R _o	AAGGATCCGTCGACATC
R _l	GACATCGATAATACGAC
POTSBE24	CATCCAACCACCATCTCGCA
POTSBE25	TTGAGAGAAGATACTAAAGT
POTSBE28	ATGTTCACTCCATCTAAAGT
POTSBE29	AGAACACAATTCTAGCTC
PBER 1	GGGGCCTTGAACTCAGCAAT
PBERT	CGTCCCAGCATTGACATAA
PBE 2B	CTTGGATCCTTGAACTCAGCAATTG
PBE 2X	TAACTCGAGCAACGCGATCACAGTTCGT

Example 2

Production of Transgenic Plants

Construction of plant transformation vectors with antisense starch branching enzyme genes

A 1200 bp *Sac I* - *Xho I* fragment, encoding approximately the -COOH half of the potato class A SBE (isolated from the rescued λZap clone 3.2.1), was cloned into the *Sac I* - *Sal I* sites of the plant transformation vector pSJ29 to create plasmid pSJ64, which is illustrated schematically in Figure 11. In the figure, the black line represents the DNA sequence. The broken line represents the bacterial plasmid backbone (containing the origin of replication and bacterial selection marker), which is not shown in full. The filled triangles on the line denote the T-DNA borders (RB = right border, LB = left border). Relevant restriction sites are shown above the black line, with the approximate distances (in kilobases) between the sites (marked by an asterisk) given by the numerals below the

line. The thinnest arrows indicate polyadenylation signals (pAnos = nopaline synthase, pAg7 = Agrobacterium gene 7), the arrows intermediate in thickness denote protein coding regions (SBE II = potato class A SBE, HYG = hygromycin resistance gene) and the thickest arrows represent promoter regions (P-2x35 = double CaMV 35S promoter, Pnos = nopaline synthase promoter). Thus pSJ64 contained the class A SBE gene fragment in an antisense orientation between the 2X 35S CaMV promoter and the nopaline synthase polyadenylation signal.

For information, pSJ29 is a derivative of the binary vector pGPTV-HYG (Becker *et al.*, 1992 Plant Molecular Biology 20, 1195-1197) modified as follows: an approximately 750 bp (*Sac* I, T4 DNA polymerase blunted - *Sal* I) fragment of pJIT60 (Guerineau *et al.*, 1992 Plant Mol. Biol. 18, 815-818) containing the duplicated cauliflower mosaic virus (CaMV) 35S promoter (Cabb-JI strain, equivalent to nucleotides 7040 to 7376 duplicated upstream of 7040 to 7433, Frank *et al.*, 1980 Cell 21, 285-294) was cloned into the *Hind* III (Klenow polymerase repaired) - *Sal* I sites of pGPTV-HYG to create pSJ29.

Plant transformation

Transformation was conducted on two types of potato plant explants; either wild type untransformed minitubers (in order to give single transformants containing the class A antisense construct alone) or minitubers from three tissue culture lines (which gave rise to plants #12, #15, #17 and #18 indicated in Table 1) which had already been successfully transformed with the class B (SBE I) antisense construct containing the tandem 35S promoter (so as to obtain double transformant plants, containing antisense sequences for both the class A and class B enzymes).

Details of the method of Agrobacterium transformation, and of the growth of transformed plants, are described in International Patent Application No. WO 95/26407, except that the medium used contained 3% sucrose (not 1%) until the final transfer and that the initial incubation with Agrobacterium (strain 3850) was performed in darkness. Transformants containing the class A antisense sequence were selected by growth in medium containing 15mg/L hygromycin (the class A antisense construct comprising the HYG gene, i.e. hygromycin phosphotransferase).

Transformation was confirmed in all cases by production of a DNA fragment from the antisense gene after PCR in the presence of appropriate primers and a crude extract of genomic DNA from each regenerated shoot.

Characterisation of starch from potato plants

Starch was extracted from plants as follows: potato tubers were homogenised in water for 2 minutes in a Waring blender operating at high speed. The homogenate was washed and filtered (initially through 2mm, then through 1mm filters) using about 4 litres of water per 100gms of tubers (6 extractions). Washed starch granules were finally extracted with acetone and air dried.

Starch extracted from singly transformed potato plants (class A/SBE II antisense, or class B/SBE I antisense), or from double transformants (class A/SBE II and class B/SBE I antisense), or from untransformed control plants, was partially characterised. The results are shown in Table 1. The table shows the amount of SBE activity (units/gram tissue) in tubers from each transformed plant. The endotherm peak temperature ($^{\circ}$ C) of starch extracted from several plants was determined by DSC, and the onset temperature ($^{\circ}$ C) of pasting was determined by reference to a viscoamylograph ("RVA"), as described in WO 95/26407. The viscoamylograph profile was as follows: step 1 - 50 $^{\circ}$ C for 2 minutes; step 2 - increase in temperature from 50 $^{\circ}$ C to 95 $^{\circ}$ C at a rate of 1.5 $^{\circ}$ C per minute; step 3 - holding at 95 $^{\circ}$ C for 15 minutes; step 4 - cooling from 95 $^{\circ}$ C to 50 $^{\circ}$ C at a rate of 1.5 $^{\circ}$ C per minute; and finally, step 5 - holding at 50 $^{\circ}$ C for 15 minutes. Table 1 shows the peak, pasting and set-back viscosities in stirring number units (SNUs), which is a measure of the amount of torque required to stir the suspensions. Peak viscosity may be defined for present purposes as the maximum viscosity attained during the heating phase (step 2) or the holding phase (step 3) of the viscoamylograph. Pasting viscosity may be defined as the viscosity attained by the starch suspensions at the end of the holding phase (step 3) of the viscoamylograph. Set-back viscosity may be defined as the viscosity of the starch suspension at the end of step 5 of the viscoamylograph.

A determination of apparent amylose content (% w/w) was also performed, using the iodometric assay method of Morrison & Laignelet (1983 J. Cereal Sci. 1, 9-20). The

results (percentage apparent amylose) are shown in Table 1. The untransformed and transformed control plants gave rise to starches having apparent amylose contents in the range 29(+/-3)%.

Generally similar values for amylose content were obtained for starch extracted from most of the singly transformed plants containing the class A (SBE II) antisense sequence. However, some plants (#152, 249) gave rise to starch having an apparent amylose content of 37-38%, notably higher than the control value. Starch extracted from these plants had markedly elevated pasting onset temperatures, and starch from plant 152 also exhibited an elevated endotherm peak temperature (starch from plant 249 was not tested by DSC).

Table 1

Sample description	Sample number	DSC			Viscometerograph			(RVA)		Apparent amylose content (% w/w)	Phosphorus content (mg/100g)
		Tuber SBE activity ([U/g starch])	Peak temperature (°C)	Onset temperature (°C)	Peak viscosity (SNU)	Pasting viscosity (SNU)	Set-back viscosity (SNU)				
Untransformed control	146 243	7.6 22.2	65.6 nd	65.5 62.0	56 761	161 135	260 241	31.2 29.1	68	68	
AS-Class A SBE	152 249	12.7 13.9	69.5 nd	70.0 70.0	497 497	300 434	529 519	37.5 36.5	68	68	
AS-Class B SBE (17) (control)	145	0.7	68.9	68.6	698	177	305	29.6	111	111	
AS-Class B SBE (17) + AS-Class A SBE	150 161	0.6 0.5	74.0 73.0	69.0 76.0	214 349	214 324	303 610	53.1 40.9	198 208	198	
AS-Class B SBE (18) (control)	144	1.6	64.5	64.7	714	154	253	29.0	67	67	
AS-Class B SBE (19) + AS-Class A SBE	149	3.0	68.5	66.9	474	287	452	35.6	127	127	
AS-Class B SBE (19) (control)	172	0.22	nd	65.4	707	167	260	26.8	130	130	
AS-Class B SBE (19) + AS-Class A SBE	201 208a 208 202 212 220	0.10 0.10 0.30 0.02 1.40 1.40	nd nd 72.8-80.5 nd nd nd	>65 >65 68.4 70.0 75.6 75.6	no peak no peak no peak no peak no peak no peak	12 15 14 172 308 345	13 17 19 215 541 560	66.4 64.1 62.6 57.9 49.5 44.1	210	210	
AS-Class B SBE (19) (control)	170	0.2	nd	60.5	768	212	303	27.8			
AS-Class B SBE (19) + AS-Class A SBE	238 238a 239a	0.7 0.9 0.6	nd nd nd	95.0 91.2 77.6	no peak no peak no peak	23 139 244	14 192 239	60.4 56.7 46.2			

50°C (2 min), 50/85°C (1.5°C/min), 85°C (15 min), 85-80°C (1.5°C/min), 80°C (15 min)

at end of 50°C (2 min), 50/85°C (1.5°C/min), 85°C (15 min).

at end of profile

Starch Branching Enzymes

Instrument "Staining Number Units" (arbitrary units)

nd

29/1

Table 1

Sample description	Sample number	Tuber SBE activity (U/g starch)	DSC		Onset temperature (°C)
			Peak temperature (°C)	Onset temperature (°C)	
Untransformed control	146	7.6	65.8	65.5	
	243	22.2	nd	62.6	
AS-Class A SBE	152	12.7	69.5	70.9	
	249	13.9	nd	70.0	
AS-Class B SBE (17) (control)	145	0.7	68.9	66.8	
	150	0.6	74.0	86.0	
AS-Class B SBE (17) + AS-Class A SBE	161	0.5	73.0	76.6	
	144	1.6	64.5	64.7	
AS-Class B SBE (18) (control)	149	3.0	68.5	69.9	

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Viscoamylograph Viscosity (RVA)		Set-back viscosity (SNU)	Apparent amylose content (% w/w)	Phosphorus content (mg/100g)
Peak viscosity (SNU)	Pasting viscosity (SNU)			
545	161	280	31.2	68
761	135	241	29.1	
467	380	529	37.5	89
497	434	518	38.5	
659	177	305	29.8	111
214	214	303	53.1	198
349	324	618	40.9	206
714	154	258	29.0	97
474	267	482	35.6	127

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	172	0.22	nd	65.4
AS-Class B SBE (15) (control)				
AS-Class B SBE (15) + AS-Class A SBE				
201	0.10	nd	>95	
208a	0.10	nd	>95	
208	0.30	72.8-80.5	>95	
202	0.02	nd	89.4	
212	1.40	nd	78.0	
220	1.40	nd	75.8	
AS-Class B SBE (12) (control)	170	0.2	nd	66.5
AS-Class B SBE (12) + AS-Class A SBE				
238	0.7	nd	95.0	
238a	0.9	nd	91.2	
230a	0.6	nd	77.6	

50°C (2 min), 50-95°C (1.5°C/min), 85°C (15 min), 85-50°C (1.5°C/min), 50°C (15 min)

at end of 50°C (2min), 50-95°C (1.5°C/min), 85°C (15 min)

at end of profile

Starch Branching Enzyme

Instrument "Sizing Number Units" (arbitrary units)

not determined

RVA profile

Pasting viscosity (47 min)

Set-back viscosity (92 min)

SBE

SNU

nd

29/4

	707	167	290	28.8	130
no peak		12	13	66.4	
no peak		15	17	64.1	
no peak		14	19	62.8	
no peak		172	245	240	
308		296	541	57.9	
355		345	593	49.5	
				44.1	
	768	202	303	27.8	
no peak				14	60.4
no peak		23		192	56.7
244		139		450	48.2
		239			

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It should be noted that, even if other single transformants were not to provide starch with an altered amylose/amyopectin ratio, the starch from such plants might still have different properties relative to starch from conventional plants (e.g. different average molecular weight or different amylopectin branching patterns), which might be useful.

Double transformant plants, containing antisense sequences for both the class A and class B enzymes, had greatly reduced SBE activity (units/gm) compared to untransformed plants or single anti-sense class A transformants, (as shown in Table 1). Moreover, certain of the double transformant plants contained starch having very significantly altered properties. For example, starch extracted from plants #201, 202, 208, 208a, 236 and 236a had drastically altered amylose/amyopectin ratios, to the extent that amylose was the main constituent of starch from these plants. The pasting onset temperatures of starch from these plants were also the most greatly increased (by about 25-30°C). Starch from plants such as #150, 161, 212, 220 and 230a represented a range of intermediates, in that such starch displayed a more modest rise in both amylose content and pasting onset temperature. The results would tend to suggest that there is generally a correlation between % amylose content and pasting onset temperature, which is in agreement with the known behaviour of starches from other sources, notably maize.

The marked increase in amylose content obtained by inhibition of class A SBE alone, compared to inhibition of class B SBE alone (see PCT/GB95/00634) might suggest that it would be advantageous to transform plants first with a construct to suppress class A SBE expression (probably, in practice, an antisense construct), select those plants giving rise to starch with the most altered properties, and then to re-transform with a construct to suppress class B SBE expression (again, in practice, probably an antisense construct), so as to maximise the degree of starch modification.

In addition to pasting onset temperatures, other features of the viscoamylograph profile e.g. for starches from plants #149, 150, 152, 161, 201, 236 and 236a showed significant differences to starches from control plants, as illustrated in Figure 13. Referring to Figure 13, a number of viscoamylograph traces are shown. The legend is as follows: shaded box - normal potato starch control (29.8% amylose content); shaded circle - starch from plant

149 (35.6% amylose): shaded triangle, pointing upwards - plant 152 (37.5%); shaded triangle, pointing downwards - plant 161 (40.9%); shaded diamond - plant 150 (53.1%); unshaded box - plant 236a (56.7%); unshaded circle - plant 236 (60.4%); unshaded triangle, pointing upwards - plant 201 (66.4%); unshaded triangle, pointing downwards - Hylon V starch, from maize (44.9 % amylose). The thin line denotes the heating profile.

With increasing amylose content, peak viscosities during processing to 95°C decrease, and the drop in viscosity from the peak until the end of the holding period at 95°C also generally decreases (indeed, for some of the starch samples there is an increase in viscosity during this period). Both of these results are indicative of reduced granule fragmentation, and hence *increased* granule stability during pasting. This property has not previously been available in potato starch without extensive prior chemical or physical modification. For applications where a maximal viscosity after processing to 95°C is desirable (i.e. corresponding to the viscosity after 47 minutes in the viscoamylograph test), starch from plant #152 would be selected as starches with both lower (Controls, #149) and higher (#161, #150) amylose contents have lower viscosities following this gelatinisation and pasting regime (Figure 13 and Table 1). It is believed that the viscosity at this stage is determined by a combination of the extent of granule swelling and the resistance of swollen granules to mechanical fragmentation. For any desired viscosity behaviour, one skilled in the art would select a potato starch from a range containing different amylose contents produced according to the invention by performing suitable standard viscosity tests.

Upon cooling pastes from 95°C to 50°C, potato starches from most plants transformed in accordance with the invention showed an increase in viscoamylograph viscosity as expected for partial reassociation of amylose. Starches from plants #149, 152 and 161 all show viscosities at 50°C significantly in excess of those for starches from control plants (Figure 13 and Table 1). This contrasts with the effect of elevated amylose contents in starches from maize plants (Figure 2) which show very low viscosities throughout the viscoamylograph test. Of particular note is the fact that, for similar amylose contents, starch from potato plant 150 (53% amylose) shows markedly increased viscosity compared with Hylon 5 starch (44.9% amylose) as illustrated in Figure 13. This demonstrates that

useful properties which require elevated (35% or greater) amylose levels can be obtained by processing starches from potato plants below 100°C, whereas more energy-intensive processing is required in order to generate similarly useful properties from high amylose starches derived from maize plants.

Final viscosity in the viscoamylograph test (set-back viscosity after 92 minutes) is greatest for starch from plant #161 (40.9% amylose) amongst those tested (Figure 13 and Table 1). Decreasing final viscosities are obtained for starches from plant #152 (37.5% amylose), #149 (35.6% amylose) and #150 (53.1% amylose). Set-back viscosity occurs where amylose molecules, exuded from the starch granule during pasting, start to re-associate outside the granule and form a viscous gel-like substance. It is believed that the set-back viscosity values of starches from transgenic potato plants represent a balance between the inherent amylose content of the starches and the ability of the amylose fraction to be exuded from the granule during pasting and therefore be available for the reassociation process which results in viscosity increase. For starches with low amylose content, increasing the amylose content tends to make more amylose available for re-association, thus increasing the set-back viscosity. However, above a threshold value, increased amylose content is thought to inhibit granule swelling, thus preventing exudation of amylose from the starch granule and reducing the amount of amylose available for re-association. This is supported by the RVA results obtained for the very high amylose content potato starches seen in the viscoamylograph profiles in Figure 13. For any desired viscosity behaviour following set-back or retrogradation to any desired temperature over any desired timescale, one skilled in the art would select a potato starch from a range containing different amylose contents produced according to the invention by performing standard viscosity tests.

Further experiments with starch from plants #201 and 208 showed that this had an apparent amylose content of over 62% (see Table 1). Viscoamylograph studies showed that starch from these plants had radically altered properties and behaved in a manner similar to hylon 5 starch from maize plants (Figure 13). Under the conditions employed in the viscoamylograph, this starch exhibited extremely limited (nearly undetectable) granule swelling. Thus, for example, unlike starch from control plants, starch from plants

201, 208 and 208a did not display a clearly defined pasting viscosity peak during the heating phase. Microscopic analysis confirmed that the starch granule structure underwent only minor swelling during the experimental heating process. This property may well be particularly useful in certain applications, as will be apparent to those skilled in the art.

Some re-grown plants have so far been found to increase still further the apparent amylose content of starch extracted therefrom. Such increases may be due to:-

- i) Growth and development of the first generation transformed plants may have been affected to some degree by the exogenous growth hormones present in the tissue culture system, which exogenous hormones were not present during growth of the second generation plants; and
- ii) Subsequent generations were grown under field conditions, which may allow for attainment of greater maturity than growth under laboratory conditions, it being generally held that amylose content of potato starch increases with maturity of the potato tuber. Accordingly, it should be possible to obtain potato plants giving rise to tubers with starch having an amylose content in excess of the 66% level so far attained, simply by analysing a greater number of transformed plants and/or by re-growing transgenic plants through one or more generations under field conditions.

Table 1 shows that another characteristic of starch which is affected by the presence of anti-sense sequences to SBE is the phosphorus content. Starch from untransformed control plants had a phosphorus content of about 60-70mg/100gram dry weight (as determined according to the AOAC Official Methods of Analysis, 15th Edition, Method 948.09 "Phosphorus in Flour"). Introduction into the plant of an anti-sense SBE B sequence was found to cause a modest increase (about two-fold) in phosphorus content, which is in agreement with the previous findings reported at scientific meetings. Similarly, anti-sense to SBE A alone causes only a small rise in phosphorus content relative to untransformed controls. However, use of anti-sense to both SBE A and B in combination results in up to a four-fold increase in phosphorus content, which is far greater than any *in planta* phosphorus content previously demonstrated for potato starch.

This is useful in that, for certain applications, starch must be phosphorylated *in vitro* by

chemical modification. The ability to obtain potato starch which, as extracted from the plant, already has a high phosphorus content will reduce the amount of *in vitro* phosphorylation required suitably to modify the starch. Thus, in another aspect the invention provides potato starch which, as extracted from the plant, has a phosphorus content in excess of 200mg/100gram dry weight starch. Typically the starch will have a phosphorus content in the range 200 - 240mg/100gram dry weight starch.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

- (A) NAME: National Starch and Chemical Investment Holding Corporation
- (B) STREET: 501 Silverside Road, Suite 27
- (C) CITY: Wilmington
- (D) STATE: Delaware
- (E) COUNTRY: United States of America
- (F) POSTAL CODE (ZIP): 19809

(ii) TITLE OF INVENTION: Improvements in or Relating to Plant Starch Composition

(iii) NUMBER OF SEQUENCES: 20

(iv) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 57 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

AAGGATCCGT CGACATCGAT AATACGACTC ACTATAGGGA TTTTTTTTTT TTTTTTTT

57

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 17 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

AAGGATCCGT CGACATC

17

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 17 base pairs

36

- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

GACATCGATA ATACGAC

17

(2) INFORMATION FOR SEQ ID NO: 4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

CATCCAACCA CCATCTCGCA

20

(2) INFORMATION FOR SEQ ID NO: 5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

TTGAGAGAAG ATACCTAAAGT

20

(2) INFORMATION FOR SEQ ID NO: 6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

ATGTTCAAGTC CATCTAAAGT

20

(2) INFORMATION FOR SEQ ID NO: 7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

AGAACAAACAA TTCCTAGCTC

20

(2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

GGGGCCTTGA ACTCAGCAAT

20

(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

CGTCCCAGCA TTCGACATAA

20

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 26 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

CTTGGATCCT TGAACTCAGC AATTG

26

(2) INFORMATION FOR SEQ ID NO: 11:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 29 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:

TAACTCGAGC AACGCGATCA CAAGTTCGT

29

(2) INFORMATION FOR SEQ ID NO: 12:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3003 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 12:

GATGGGGCCT	TGAACTCAGC	AATTGACAC	TCAGTTAGTT	ACACTGCCAT	CACTTATCAG	60
ATCTCTATT	TTTCTCTAA	TTCCAACCAA	GGAATGAATA	AAAAGATAGA	TTTGAAAAAA	120
CCCTAAGGAG	AGAAGAAGAA	AGATGGTGTA	TACACTCTCT	GGAGTTCGTT	TTCTACTGT	180
TCCATCAGTG	TACAAATCTA	ATGGATTCA	CAGTAATGGT	GATCGGAGGA	ATGCTAATAT	240
TTCTGTATT	TTGAAAAAAC	ACTCTTTTC	ACGGAAGATC	TTGGCTGAAA	AGTCTTCTTA	300
CAATTCCGAA	TCCCGACCTT	CTACAATTGC	AGCATCGGGG	AAAGTCCTTG	TGCCTGGAAT	360
CCAGAGTGAT	AGCTCCTCAT	CCTCAACAGA	TCAATTGAG	TTCGCTGAGA	CATCTCCAGA	420
AAATTCCCCA	GCATCAACTG	ATGTAGATAG	TTCAACAATG	GAACACGCTA	GCCAGATTAA	480
AACTGAGAAC	GATGACGTTG	AGCCGTCAAG	TGATCTTACA	GGAAAGTGTG	AAGAGCTGGA	540
TTTTGCTTCA	TCACTACAAC	TACAAGAAGG	TGGTAAACTG	GAGGAGTCTA	AAACATTAAA	600
TACTTCTGAA	GAGACAATT	TTGATGAATC	TGATAGGATC	AGAGAGAGGG	GCATCCCTCC	660
ACCTGGACTT	GGTCAGAAGA	TTTATGAAAT	AGACCCCTT	TTGACAAACT	ATCGTCAACA	720
CCTTGATTAC	AGGTATTAC	AGTACAAGAA	ACTGAGGGAG	GCAATTGACA	AGTATGAGGG	780
TGGTTGGAA	GCTTTTCTC	GTGGTTATGA	AAGAATGGGT	TTCACTCGTA	GTGCTACAGG	840
TATCACTTAC	CGTGAGTGGG	CTCCTGGTGC	CCAGTCAGCT	GCCCTCATTG	GGGATTTCAA	900
CAATTGGGAC	GCAAATGCTG	ACTTATGAC	TCGGAATGAA	TTTGGTGTCT	GAGAGATTT	960
TCTGCCAAAT	AATGTGGATG	GTTCTCCTGC	AATTCTCAT	GGGTCCAGAG	TGAAGATACG	1020
TATGGACACT	CCATCAGGTG	TTAAGGATT	CATTCTGCT	TGGATCAACT	ACTCTTTACA	1080
GCTTCCTGAT	GAAATTCCAT	ATAATGGAAT	ATATTATGAT	CCACCCGAAG	AGGAGAGGTA	1140
TATCTTCAA	CACCCACGGC	CAAAGAAACC	AAAGTCGGTG	AGAATATATG	AATCTCATAT	1200
TGGAATGAGT	AGTCCGGAGC	CTAAAATTAA	CTCATACTG	AATTAGAG	ATGAAGTTCT	1260
TCCTCGCATA	AAAAAAGCTT	GGGTACAATG	CGGTGCAAAT	TATGGCTATT	CAAGAGCATT	1320
CTTATTATGC	TAGTTTGGT	TATCATGTCA	CAAATTTTT	TGCACCAAGC	AGCCGTTTG	1380

GAACGCCCGA CGACCTTAAG TCTTGATTG ATAAAGCTCA TGAGCTAGGA ATTGTTGTTC	1440
TCATGGACAT TGTCACAGC CATGCATCAA ATAATACCTT AGATGGACTG AACATGTTG	1500
ACGGCACAGA TAGTTGTTAC TTTCACTCTG GAGCTCGTGG TTATCATTGG ATGTGGGATT	1560
TCCGCCTCTT TAACTATGGA AACTGGGAGG TACTTAGGTA TCTTCTCTCA AATGCGAGAT	1620
GGTGGTTGGA TGAGTTCAA TTTGATGGAT TTAGATTGA TGGTGTGACA TCAATGATGT	1680
GTACTCACCA CGGATTATCG GTGGGATTCA CTGGGAACTA CGAGGAATAC TTTGGACTCG	1740
CAACTGATGT GGATGCTGTT GTGTATCTGA TGCTGGTCAA CGATCTTATT CATGGGCTTT	1800
TCCCAGATGC AATTACCATT GGTGAAGATG TTAGCGGAAT GCCGACATT TGTGTTCCCG	1860
TTCAAGATGG GGGTGGTGGC TTTGACTATC GGCTGCATAT GGCAATTGCT GATAAATGGA	1920
TTGAGTTGCT CAAGAACCG GATGAGGATT GGAGAGTGGG TGATATTGTT CATAACTGA	1980
CAAATAGAAG ATGGTCGGAA AAGTGTGTTT CATACTGTA AAGTCATGAT CAAGCTCTAG	2040
TCGGTGATAA AACTATAGCA TTCTGGCTGA TGGACAAGGA TATGTATGAT TTTATGGCTC	2100
TGGATAGACC GTCAACATCA TTAATAGATC GTGGGATAGC ATTACACAAG ATGATTAGGC	2160
TTGTAACATAT GGGATTAGGA GGAGAAGGGT ACCTAAATT CATGGGAAAT GAATTGGCC	2220
ACCCCTGAGTG GATTGATTTC CCTAGGGCTG AACAAACACCT CTCTGATGGC TCAGTAATT	2280
CCAGAAACCA ATTCAGTTAT GATAATGCA GACGGAGATT TGACCTGGGA GATGCAGAAT	2340
ATTTAAGATA CCGTGGTTG CAAGAATTG ACCGGGCTAT GCAGTATCTT GAAGATAAT	2400
ATGAGTTTAT GACTTCAGAA CACCAAGTTCA TATCACGAAA GGATGAAGGA GATAGGATGA	2460
TTGTATTGAA AAAAGGAAAC CTAGTTTGT TCTTTAATT TCACGGACA AAAGGCTATT	2520
CAGACTATCG CATAGGCTGC CTGAAGCCTG GAAAATACAA GGTTGCCTTG GACTCAGATG	2580
ATCCACTTTT TGGTGGCTTC GGGAGAATTG ATCATAATGC CGAATATTTC ACCTTTGAAG	2640
GATGGTATGA TGATCGTCCT CGTTCAATT TGGTGTATGC ACCTAGTAGA ACAGCAGTGG	2700
TCTATGCACT AGTAGACAAA GAAGAAGAAG AAGAAGAAGA AGTAGCAGTA GTAGAAGAAG	2760
TAGTAGTAGA AGAAGAATGA ACGAACTTGT GATCGCGTTG AAAGATTGAA ACGCCACATA	2820
GAGCTTCTTG ACGTATCTGG CAATATTGCA TTAGTCTTGG CGGAATTCA TGTGACAACA	2880
GGTTGCAAT TCTTCCACT ATTAGTAGTG CAACGATATA CGCAGAGATG AAGTGCTGAA	2940
CAAAACATA TGTAAAATCG ATGAATTAT GTCGAATGCT GGGACGATCG AATTCTGCA	3000
GCC	3003

40

(2) INFORMATION FOR SEQ ID NO: 13:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2975 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:

TTGATGGGCC TTGAACTCAG CAATTGACA CTCAGTTAGT TACACTCCTA TCACATTATCA	60
GATCTCTATT TTTTCTCTTA ATTCCAACCA GGGGAATGAA TAAAAGGATA GATTTGTAAA	120
AACCCCTAAGG AGAGAAGAAG AAAGATGGTG TATATACTCT CTGGAGTTCG TTTTCTACT	180
GTTCCATCAG TGTACAAATC TAATGGATT GCAGTAATG GTGATCGGAG GAATGCTAAT	240
GTTTCTGTAT TCTTGAAAAA GCACTCTCTT TCACGGAAGA TCTTGGCTGA AAAGTCTTCT	300
TACAATTCCG AATTCCGACC TTCTACAGTT GCAGCATCGG GGAAAGTCCT TGTGCCTGGA	360
ACCCAGAGTG ATAGCTCCTC ATCCTCAACA GACCAATTG AGTTCACTGA GACATCTCCA	420
GAAAATTCCC CAGCATCAAC TGATGTAGAT AGTTCAACAA TGGAACACGC TAGCCAGATT	480
AAAACGTAGA ACGATGACGT TGAGCCGTCA AGTGATCTTA CAGGAAGTGT TGAAGAGCTG	540
GATTTGCTT CATCACTACA ACTACAAGAA GGTGGTAAAC TGGAGGAGTC TAAAACATTA	600
AATACTTCTG AAGAGACAAT TATTGATGAA TCTGATAGGA TCAGAGAGAG GGGCATCCCT	660
CCACCTGGAC TTGGTCAGAA GATTATGAA ATAGACCCCC TTTGACAAA CTATCGTCAA	720
CACCTTGATT ACAGGTATTAC ACAGTACAAG AAACTGAGGG AGGCAATTGA CAAGTATGAG	780
GGTGGTTTGG AAGCTTTCT CGTGGTTATG AAAAATGGG TTTCACTCGT AGTGCTACAG	840
GTATCACTTA CCGTGAGTGG GCTCCTGGTG CCCAGTCAGC TGCCCTCATT GGAGATTC	900
ACAATTGGGA CGCAAATGCT GACATTATGA CTCGGAATGA ATTTGGTGT TGAGGAGATT	960
TTCTGCCAAA TAATGTGGAT GGTTCTCCTG CAATTCTCA TGGGTCCAGA GTGAAGATAC	1020
GTATGGACAC TCCATCAGGT GTTAAGGATT CCATTCTGC TTGGATCAAC TACTCTTAC	1080
AGCTTCTGA TGAAATTCCA TATAATGGAA TATATTATGA TCCACCCGAA GAGGAGAGGT	1140
ATATCTTCCA ACACCCACGG CCAAAGAAC CAAAGTCGCT GAGAATATAT GAATCTCATA	1200
TTGGAATGAG TAGTCCGGAG CCTAAAATTA ACTCATACTG GAATTTAGA GATGAAGTTC	1260
TTCCTCGCAT AAAAAGCTT GGGTACAATG CGCTGCGAAT TATGGCTATT CAAGAGCATT	1320
CTTATTATGC TAGTTTGGT TATCATGTCA CAAATTTTT TGACCCAAGC AGCCGTTTG	1380

GAACGCCCGA CGACCTTAAG TCTTCGATTG ATAAAGCTCA TGAGCTAGGA ATTGTTGTTC	1440
TCATGGACAT CGTTCACAGC CATGCATCAA ATAATACTTT AGATGGACTG AACATGTTG	1500
ACGGCACCGA TAGTTGTTAC TTTCACTCTG GAGCTCGTGG TTATCATTGG ATGTGGGATT	1560
CCGCCTCTTT AACTATGGAA ACTGGGAGGT ACTTAGGTAT CTTCTCTCAA ATGCGAGATG	1620
GTGGTTGGAT GAGTTCAAAT TTGATGGATT TAGATTCGAT GGTGTGACAT CAATGATGTA	1680
TACTCACCAC GGATTATCGG TGGGATTCAC TGGGAACACTAC GAGGAATACT TTGGACTCGC	1740
AACTGATGTG GATGCTGTTG TGTATCTGAT GCTGGTCAAC GATCTTATTTC ATAGGCTTTT	1800
CCCAGATGCA ATTACCATTG GTGAAGATGT TAGCGGAATG CCGACATTTT GTATTCCCGT	1860
TCAAGATGGG GGTGTTGGCT TTGACTATCG GCTGCATATG GCAATTGCTG ATAAATGGAT	1920
TGAGTTGCTC AAGAACCGGG ATGAGGATTG GAGAGTGGGT GATATTGTTC ATACACTGAC	1980
AAATAGAAGA TGGTCGGAAA AGTGTGTTTC ATACGCTGAA AGTCATGATC AAGCTCTAGT	2040
CGGTGATAAA ACTATAGCAT TCTGGCTGAT GGACAAGGAT ATGTATGATT TTATGGCTCT	2100
GGATAGACCG CCAACATCAT TAATAGATCG TGGGATAGCA TTGCACAAGA TGATTAGGCT	2160
TGTAACTATG GGATTAGGAG GAGAAGGGTA CCTAAATTTC ATGGGAAATG AATTGGCCA	2220
CCCTGAGTGG ATTGATTTCCT CTAGGGCTGA GCCACACCTT TCTGATGGCT CAGTAATTCC	2280
CGGAAACCAA TTCAGTTATG ATAAATGCAG ACGGAGATT GACCTGGGAG ATGCAGAATA	2340
TTTAAGATAC CATGGGTTAC AAGAATTGATCTGGCTATG CAGTATCTTG AAGATAAATA	2400
TGAGTTTATG ACTTCAGAAC ACCAGTTCAT ATCACGAAAG GATGAAGGAG ATAGGATGAT	2460
TGTATTGAA AGAGGAAACC TAGTTTCGT CTTTAATTTC CACTGGACAA ATAGCTATTTC	2520
AGACTATCGC ATAGGCTGCC TGAAGCCTGG AAAATACAAG GTTGTCTGG ACTCAGATGA	2580
TCCACTTTT GGTGGCTTCG GGAGAATTGA TCATAATGCC GAATATTCA CCTCTGAAGG	2640
ATCGTATGAT GATCGTCCTT GTTCAATTAT GGTGTATGCA CCTAGTAGAA CAGCAGTGGT	2700
CTATGCACTA GTAGACAAAC TAGAAGTAGC AGTAGTAGAA GAACCCATTG AAGAATGAAC	2760
GAACTTGTGA TCGCGTTGAA AGATTGAAC GTTACTTGGT CATCCACATA GAGCTTCTTG	2820
ACATCAGTCT TGGCGGAATT GCATGTGACA ACAAGGTTTG CAGTTCTTTC CACTATTAGT	2880
AGTCCACCGA TATACGCAGA GATGAAGTGC TGAACAAACA TATGTAAAAT CGATGAATT	2940
ATGTCGAATG CTGGGACGAT CGAATTCTG CAGCC	2975

(2) INFORMATION FOR SEQ ID NO: 14:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3033 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ix) FEATURE:

- (A) NAME/KEY: CDS
(B) LOCATION: 145..2790

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:

CTA CAA CTA CAA GAA GGT AAA CTG GAG GAG TCT AAA ACA TTA AAT Leu Gin Leu Glu Glu Gly Gly Lys Leu Glu Glu Ser Lys Thr Leu Asn 140 145 150	603
ACT TCT GAA GAG ACA ATT ATT GAT GAA TCT GAT AGG ATC AGA GAG AGG Thr Ser Glu Glu Thr Ile Ile Asp Glu Ser Asp Arg Ile Arg Glu Arg 155 160 165	651
GGC ATC CCT CCA CCT GGA CTT GGT CAG AAG ATT TAT GAA ATA GAC CCC Gly Ile Pro Pro Pro Gly Leu Gly Glu Lys Ile Tyr Glu Ile Asp Pro 170 175 180 185	699
CTT TTG ACA AAC TAT CGT CAA CAC CTT GAT TAC AGG TAT TCA CAG TAC Leu Leu Thr Asn Tyr Arg Glu His Leu Asp Tyr Arg Tyr Ser Glu Tyr 190 195 200	747
AAG AAA CTG AGG GAG GCA ATT GAC AAG TAT GAG GGT GGT TTG GAA GCC Lys Lys Leu Arg Glu Ala Ile Asp Lys Tyr Glu Gly Gly Leu Glu Ala 205 210 215	795
TTT TCT CGT GGT TAT GAA AAA ATG GGT TTC ACT CGT AGT GCT ACA GGT Phe Ser Arg Gly Tyr Glu Lys Met Gly Phe Thr Arg Ser Ala Thr Gly 220 225 230	843
ATC ACT TAC CGT GAG TGG GCT CTT GGT GCC CAG TCA GCT GCC CTC ATT Ile Thr Tyr Arg Glu Trp Ala Leu Gly Ala Glu Ser Ala Ala Leu Ile 235 240 245	891
GGA GAT TTC AAC AAT TGG GAC GCA AAT GCT GAC ATT ATG ACT CGG AAT Gly Asp Phe Asn Asn Trp Asp Ala Asn Ala Asp Ile Met Thr Arg Asn 250 255 260 265	939
GAA TTT GGT GTC TGG GAG ATT TTT CTG CCA AAT AAT GTG GAT GGT TCT Glu Phe Gly Val Trp Glu Ile Phe Leu Pro Asn Asn Val Asp Gly Ser 270 275 280	987
CCT GCA ATT CCT CAT GGG TCC AGA GTG AAG ATA CGT ATG GAC ACT CCA Pro Ala Ile Pro His Gly Ser Arg Val Lys Ile Arg Met Asp Thr Pro 285 290 295	1035
TCA GGT GTT AAG GAT TCC ATT CCT GCT TGG ATC AAC TAC TCT TTA CAG Ser Gly Val Lys Asp Ser Ile Pro Ala Trp Ile Asn Tyr Ser Leu Glu 300 305 310	1083
CTT CCT GAT GAA ATT CCA TAT AAT GGA ATA CAT TAT GAT CCA CCC GAA Leu Pro Asp Glu Ile Pro Tyr Asn Gly Ile His Tyr Asp Pro Pro Glu 315 320 325	1131
GAG GAG AGG TAT ATC TTC CAA CAC CCA CGG CCA AAG AAA CCA AAG TCG Glu Glu Arg Tyr Ile Phe Gin His Pro Arg Pro Lys Lys Pro Lys Ser 330 335 340 345	1179
CTG AGA ATA TAT GAA TCT CAT ATT GGA ATG AGT AGT CCG GAG CCT AAA Leu Arg Ile Tyr Glu Ser His Ile Gly Met Ser Ser Pro Glu Pro Lys 350 355 360	1227

ATT AAC TCA TAC GTG AAT TTT AGA GAT GAA GTT CTT CCT CGC ATA AAA Ile Asn Ser Tyr Val Asn Phe Arg Asp Glu Val Leu Pro Arg Ile Lys 365 370 375	1275
AAG CTT GGG TAC AAT GCG CTG CAA ATT ATG GCT ATT CAA GAG CAT TCT Lys Leu Gly Tyr Asn Ala Leu Gln Ile Met Ala Ile Gln Glu His Ser 380 385 390	1323
TAT TAC GCT AGT TTT GGT TAT CAT GTC ACA AAT TTT TTT GCA CCA AGC Tyr Tyr Ala Ser Phe Gly Tyr His Val Thr Asn Phe Phe Ala Pro Ser 395 400 405	1371
AGC CGT TTT GGA ACG CCC GAC GAC CTT AAG TCT TTG ATT GAT AAA GCT Ser Arg Phe Gly Thr Pro Asp Asp Leu Lys Ser Leu Ile Asp Lys Ala 410 415 420 425	1419
CAT GAG CTA GGA ATT GTT GTT CTC ATG GAC ATT GTT CAC AGC CAT GCA His Glu Leu Gly Ile Val Val Leu Met Asp Ile Val His Ser His Ala 430 435 440	1467
TCA AAT AAT ACT TTA GAT GGA CTG AAC ATG TTT GAC TGC ACC GAT AGT Ser Asn Asn Thr Leu Asp Gly Leu Asn Met Phe Asp Cys Thr Asp Ser 445 450 455	1515
TGT TAC TTT CAC TCT GGA GCT CGT GGT TAT CAT TGG ATG TGG GAT TCC Cys Tyr Phe His Ser Gly Ala Arg Gly Tyr His Trp Met Trp Asp Ser 460 465 470	1563
CGC CTC TTT AAC TAT GGA AAC TGG GAG GTA CTT AGG TAT CTT CTC TCA Arg Leu Phe Asn Tyr Gly Asn Trp Glu Val Leu Arg Tyr Leu Leu Ser 475 480 485	1611
AAT GCG AGA TGG TGG TTG GAT GCG TTC AAA TTT GAT GGA TTT AGA TTT Asn Ala Arg Trp Trp Leu Asp Ala Phe Lys Phe Asp Gly Phe Arg Phe 490 495 500 505	1659
GAT GGT GTG ACA TCA ATG ATG TAT ATT CAC CAC GGA TTA TCG GTG GGA Asp Gly Val Thr Ser Met Met Tyr Ile His His Gly Leu Ser Val Gly 510 515 520	1707
TTC ACT GGG AAC TAC GAG GAA TAC TTT GGA CTC GCA ACT GAT GTG GAT Phe Thr Gly Asn Tyr Glu Glu Tyr Phe Gly Leu Ala Thr Asp Val Asp 525 530 535	1755
GCT GTT GTG TAT CTG ATG CTG GTC AAC GAT CTT ATT CAT GGG CTT TTC Ala Val Val Tyr Leu Met Leu Val Asn Asp Leu Ile His Gly Leu Phe 540 545 550	1803
CCA GAT GCA ATT ACC ATT GGT GAA GAT GTT AGC GGA ATG CCG ACA TTT Pro Asp Ala Ile Thr Ile Gly Glu Asp Val Ser Gly Met Pro Thr Phe 555 560 565	1851
TGT ATT CCC GTC CAA GAG GGG GGT GTT GGC TTT GAC TAT CGG CTG CAT Cys Ile Pro Val Gln Glu Gly Gly Val Gly Phe Asp Tyr Arg Leu His 570 575 580 585	1899

ATG GCA ATT GCT GAT AAA CGG ATT GAG TTG CTC AAG AAA CGG GAT GAG Met Ala Ile Ala Asp Lys Arg Ile Glu Leu Leu Lys Lys Arg Asp Glu 590 595 600	1947
GAT TGG AGA GTG GGT GAT ATT GTT CAT ACA CTG ACA AAT AGA AGA TGG Asp Trp Arg Val Gly Asp Ile Val His Thr Leu Thr Asn Arg Arg Trp 605 610 615	1995
TCG GAA AAG TGT GTT TCA TAC GCT GAA AGT CAT GAT CAA GCT CTA GTC Ser Glu Lys Cys Val Ser Tyr Ala Glu Ser His Asp Gln Ala Leu Val 620 625 630	2043
GGT GAT AAA ACT ATA GCA TTC TGG CTG ATG GAC AAG GAT ATG TAT GAT Gly Asp Lys Thr Ile Ala Phe Trp Leu Met Asp Lys Asp Met Tyr Asp 635 640 645	2091
TTT ATG GCT CTG GAT AGA CCG TCA ACA TCA TTA ATA GAT CGT GGG ATA Phe Met Ala Leu Asp Arg Pro Ser Thr Ser Leu Ile Asp Arg Gly Ile 650 655 660 665	2139
GCA TTG CAC AAG ATG ATT AGG CTT GTA ACT ATG GGA TTA GGA GGA GAA Ala Leu His Lys Met Ile Arg Leu Val Thr Met Gly Leu Gly Glu 670 675 680	2187
GGG TAC CTA AAT TTC ATG GGA AAT GAA TTC GGC CAC CCT GAG TGG ATT Gly Tyr Leu Asn Phe Met Gly Asn Glu Phe Gly His Pro Glu Trp Ile 685 690 695	2235
GAT TTC CCT AGG GCT GAA CAA CAC CTC TCT GAT GGC TCA GTA ATC CCC Asp Phe Pro Arg Ala Glu Gln His Leu Ser Asp Gly Ser Val Ile Pro 700 705 710	2283
GGA AAC CAA TTC AGT TAT GAT AAA TGC AGA CGG AGA TTT GAC CTG GGA Gly Asn Gln Phe Ser Tyr Asp Lys Cys Arg Arg Phe Asp Leu Gly 715 720 725	2331
GAT GCA GAA TAT TTA AGA TAC CGT GGG TTG CAA GAA TTT GAC CGG CCT Asp Ala Glu Tyr Leu Arg Tyr Arg Gly Leu Gln Glu Phe Asp Arg Pro 730 735 740 745	2379
ATG CAG TAT CTT GAA GAT AAA TAT GAG TTT ATG ACT TCA GAA CAC CAG Met Gln Tyr Leu Glu Asp Lys Tyr Glu Phe Met Thr Ser Glu His Gln 750 755 760	2427
TTC ATA TCA CGA AAG GAT GAA GGA GAT AGG ATG ATT GTA TTT GAA AAA Phe Ile Ser Arg Lys Asp Glu Gly Asp Arg Met Ile Val Phe Glu Lys 765 770 775	2475
GGA AAC CTA GTT TTT GTC TTT AAT TTT CAC TGG ACA AAA AGC TAT TCA Gly Asn Leu Val Phe Val Phe Asn Phe His Trp Thr Lys Ser Tyr Ser 780 785 790	2523
GAC TAT CGC ATA GCC TGC CTG AAG CCT GGA AAA TAC AAG GTT GCC TTG Asp Tyr Arg Ile Ala Cys Leu Lys Pro Gly Lys Tyr Lys Val Ala Leu 795 800 805	2571

GAC TCA GAT GAT CCA CTT TTT GGT GGC TTC GGG AGA ATT GAT CAT AAT Asp Ser Asp Asp Pro Leu Phe Gly Gly Phe Gly Arg Ile Asp His Asn 810 815 820 825	2619
GCC GAA TAT TTC ACC TTT GAA GGA TGG TAT GAT GAT CGT CCT CGT TCA Ala Glu Tyr Phe Thr Phe Glu Gly Trp Tyr Asp Asp Arg Pro Arg Ser 830 835 840	2667
ATT ATG GTG TAT GCA CCT TGT AAA ACA GCA GTG GTC TAT GCA CTA GTA Ile Met Val Tyr Ala Pro Cys Lys Thr Ala Val Val Tyr Ala Leu Val 845 850 855	2715
GAC AAA GAA GTA GCA GCA Asp Lys Glu Glu Glu Glu Glu Glu Glu Glu Glu Val Ala Ala 860 865 870	2763
GTA GAA GAA GTA GTA GAA GAA GAA TGAACGAACT TGTGATCGCG Val Glu Glu Val Val Glu Glu Glu 875 880	2810
TTGAAAGATT TGAACGCTAC ATAGAGCTTC TTGACGTATC TGGCAATATT GCATCAGTCT TGGCGGAATT TCATGTGACA CAAGGTTGC AATTCTTCC ACTATTAGTA GTGCAACGAT ATACGCAGAG ATGAAGTGCT GAACAAACAT ATGTAAAATC GATGAATTAA TGTCGAATGC TGGGACGATC GAATTCTGC AGGCCGGGGG ACCCCTTAGT TCT	2870 2930 2990 3033

(2) INFORMATION FOR SEQ ID NO: 15:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 882 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:

Met Val Tyr Thr Leu Ser Gly Val Arg Phe Pro Thr Val Pro Ser Val 1 5 10 15
Tyr Lys Ser Asn Gly Phe Ser Ser Asn Gly Asp Arg Arg Asn Ala Asn 20 25 30
Val Ser Val Phe Leu Lys Lys His Ser Leu Ser Arg Lys Ile Leu Ala 35 40 45
Glu Lys Ser Ser Tyr Asn Ser Glu Phe Arg Pro Ser Thr Val Ala Ala 50 55 60
Ser Gly Lys Val Leu Val Pro Gly Thr Gln Ser Asp Ser Ser Ser 65 70 75 80
Ser Thr Asp Gln Phe Glu Phe Thr Glu Thr Ser Pro Glu Asn Ser Pro 85 90 95

47

Ala Ser Thr Asp Val Asp Ser Ser Thr Met Glu His Ala Ser Gln Ile
100 105 110

Lys Thr Glu Asn Asp Asp Val Glu Pro Ser Ser Asp Leu Thr Gly Ser
115 120 125

Val Glu Glu Leu Asp Phe Ala Ser Ser Leu Gln Leu Gln Glu Gly Gly
130 135 140

Lys Leu Glu Glu Ser Lys Thr Leu Asn Thr Ser Glu Glu Thr Ile Ile
145 150 155 160

Asp Glu Ser Asp Arg Ile Arg Glu Arg Gly Ile Pro Pro Pro Gly Leu
165 170 175

Gly Gln Lys Ile Tyr Glu Ile Asp Pro Leu Leu Thr Asn Tyr Arg Gln
180 185 190

His Leu Asp Tyr Arg Tyr Ser Gln Tyr Lys Lys Leu Arg Glu Ala Ile
195 200 205

Asp Lys Tyr Glu Gly Gly Leu Glu Ala Phe Ser Arg Gly Tyr Glu Lys
210 215 220

Met Gly Phe Thr Arg Ser Ala Thr Gly Ile Thr Tyr Arg Glu Trp Ala
225 230 235 240

Leu Gly Ala Gln Ser Ala Ala Leu Ile Gly Asp Phe Asn Asn Trp Asp
245 250 255

Ala Asn Ala Asp Ile Met Thr Arg Asn Glu Phe Gly Val Trp Glu Ile
260 265 270

Phe Leu Pro Asn Asn Val Asp Gly Ser Pro Ala Ile Pro His Gly Ser
275 280 285

Arg Val Lys Ile Arg Met Asp Thr Pro Ser Gly Val Lys Asp Ser Ile
290 295 300

Pro Ala Trp Ile Asn Tyr Ser Leu Gln Leu Pro Asp Glu Ile Pro Tyr
305 310 315 320

Asn Gly Ile His Tyr Asp Pro Pro Glu Glu Glu Arg Tyr Ile Phe Gln
325 330 335

His Pro Arg Pro Lys Lys Pro Lys Ser Leu Arg Ile Tyr Glu Ser His
340 345 350

Ile Gly Met Ser Ser Pro Glu Pro Lys Ile Asn Ser Tyr Val Asn Phe
355 360 365

Arg Asp Glu Val Leu Pro Arg Ile Lys Lys Leu Gly Tyr Asn Ala Leu
370 375 380

Gln Ile Met Ala Ile Gln Glu His Ser Tyr Tyr Ala Ser Phe Gly Tyr
385 390 395 400

His Val Thr Asn Phe Phe Ala Pro Ser Ser Arg Phe Gly Thr Pro Asp
 405 410 415
 Asp Leu Lys Ser Leu Ile Asp Lys Ala His Glu Leu Gly Ile Val Val
 420 425 430
 Leu Met Asp Ile Val His Ser His Ala Ser Asn Asn Thr Leu Asp Gly
 435 440 445
 Leu Asn Met Phe Asp Cys Thr Asp Ser Cys Tyr Phe His Ser Gly Ala
 450 455 460
 Arg Gly Tyr His Trp Met Trp Asp Ser Arg Leu Phe Asn Tyr Gly Asn
 465 470 475 480
 Trp Glu Val Leu Arg Tyr Leu Leu Ser Asn Ala Arg Trp Trp Leu Asp
 485 490 495
 Ala Phe Lys Phe Asp Gly Phe Arg Phe Asp Gly Val Thr Ser Met Met
 500 505 510
 Tyr Ile His His Gly Leu Ser Val Gly Phe Thr Gly Asn Tyr Glu Glu
 515 520 525
 Tyr Phe Gly Leu Ala Thr Asp Val Asp Ala Val Val Tyr Leu Met Leu
 530 535 540
 Val Asn Asp Leu Ile His Gly Leu Phe Pro Asp Ala Ile Thr Ile Gly
 545 550 555 560
 Glu Asp Val Ser Gly Met Pro Thr Phe Cys Ile Pro Val Gln Glu Gly
 565 570 575
 Gly Val Gly Phe Asp Tyr Arg Leu His Met Ala Ile Ala Asp Lys Arg
 580 585 590
 Ile Glu Leu Leu Lys Lys Arg Asp Glu Asp Trp Arg Val Gly Asp Ile
 595 600 605
 Val His Thr Leu Thr Asn Arg Arg Trp Ser Glu Lys Cys Val Ser Tyr
 610 615 620
 Ala Glu Ser His Asp Gln Ala Leu Val Gly Asp Lys Thr Ile Ala Phe
 625 630 635 640
 Trp Leu Met Asp Lys Asp Met Tyr Asp Phe Met Ala Leu Asp Arg Pro
 645 650 655
 Ser Thr Ser Leu Ile Asp Arg Gly Ile Ala Leu His Lys Met Ile Arg
 660 665 670
 Leu Val Thr Met Gly Leu Gly Gly Glu Gly Tyr Leu Asn Phe Met Gly
 675 680 685
 Asn Glu Phe Gly His Pro Glu Trp Ile Asp Phe Pro Arg Ala Glu Gln
 690 695 700

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His Leu Ser Asp Gly Ser Val Ile Pro Gly Asn Gln Phe Ser Tyr Asp
 705 710 715 720
 Lys Cys Arg Arg Arg Phe Asp Leu Gly Asp Ala Glu Tyr Leu Arg Tyr
 725 730 735
 Arg Gly Leu Gln Glu Phe Asp Arg Pro Met Gln Tyr Leu Glu Asp Lys
 740 745 750
 Tyr Glu Phe Met Thr Ser Glu His Gln Phe Ile Ser Arg Lys Asp Glu
 755 760 765
 Gly Asp Arg Met Ile Val Phe Glu Lys Gly Asn Leu Val Phe Val Phe
 770 775 780
 Asn Phe His Trp Thr Lys Ser Tyr Ser Asp Tyr Arg Ile Ala Cys Leu
 785 790 795 800
 Lys Pro Gly Lys Tyr Lys Val Ala Leu Asp Ser Asp Asp Pro Leu Phe
 805 810 815
 Gly Gly Phe Gly Arg Ile Asp His Asn Ala Glu Tyr Phe Thr Phe Glu
 820 825 830
 Gly Trp Tyr Asp Asp Arg Pro Arg Ser Ile Met Val Tyr Ala Pro Cys
 835 840 845
 Lys Thr Ala Val Val Tyr Ala Leu Val Asp Lys Glu Glu Glu Glu
 850 855 860
 Glu Glu Glu Glu Glu Glu Val Ala Ala Val Glu Glu Val Val Val Glu
 865 870 875 880
 Glu Glu

(2) INFORMATION FOR SEQ ID NO: 16:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2576 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:

TCATTAAGA GGAGAAATT	ACTATGAGAG GATCTCACCA	TCACCATCAC CATGGGATCT	60
TGGCTGAAAA GTCTTCTTAC	AATTCCGAAT TCCGACCTTC	TACAGTTGCA GCATCGGGGA	120
AAGTCCTTGT GCCTGGAACC	CAGAGTGATA GCTCCTCATC	CTAACAAAC CAATTTGAGT	180
TCACTGAGAC ATCTCCAGAA	AATTCCCCAG CATCAACTGA	TGTAGATAGT TCAACAATGG	240
AACACGCTAG CCAGATTAAA	ACTGAGAACG ATGACGTTGA	GCCGTCAAGT GATCTTACAG	300

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GAAGTGTGAA AGAGCTGGAT TTTGCTTCAT CACTACAAC ACAAGAAGGT GGTAAACTGG 360
AGGAGTCTAA AACATTAAAT ACTTCTGAAG AGACAATTAT TGATGAATCT GATAGGATCA 420
GAGAGAGGGG CATCCCTCCA CCTGGACTTG GTCAGAAGAT TTATGAAATA GACCCCTTT 480
TGACAAACTA TCGTCAACAC CTTGATTACA GGTATTACA GTACAAGAAA CTGAGGGAGG 540
CAATTGACAA GTATGAGGGT GGTTTGGAAAG CTTTTCTCG TGGTTATGAA AAAATGGGTT 600
TCACTCGTAG TGCTACAGGT ATCACTTACC GTGAGTGGGC TCCTGGTGCC CAGTCAGCTG 660
CCCTCATTGG AGATTCAAC AATTGGGACG CAAATGCTGA CATTATGACT CGGAATGAAT 720
TTGGTGTCTG GGAGATTTT CTGCCAAATA ATGTGGATGG TTCTCCTGCA ATTCCCTCATG 780
GGTCCAGAGT GAAGATACT ATGGACACTC CATCAGGTGT TAAGGATTCC ATTCCCTGCTT 840
GGATCAACTA CTCTACAGCT TCCTGATGAA ATTCCATATA ATGGAATATA TTATGATCCA 900
CCCGAAGAGG AGAGGTATAT CTTCCAACAC CCACGGCCAA AGAAACCAAA GTCGCTGAGA 960
ATATATGAAT CTCATATTGG AATGAGTAGT CCGGAGCCTA AAATTAACCTC ATACGTGAAT 1020
TTTAGAGATG AAGTTCTTCC TCGCATAAAA AAGCTTGGGT ACAATGCGCT GCAAATTATG 1080
GCTATTCAAG AGCATTCTTA TTATGCTAGT TTTGGTTATC ATGTCACAAA TTTTTTGCA 1140
CCAAGCAGCC GTTTTGGAAC GCCCGACGAC CTTAAGTCTT TGATTGATAA AGCTCATGAG 1200
CTAGGAATTG TTGTTCTCAT GGACATTGTT CACAGCCATG CATCAAATAA TACTTTAGAT 1260
GGACTGAACA TGTTTGACGG CACCGATAGT TGTTACTTC ACTCTGGAGC TCGTGGTTAT 1320
CATTGGATGT GGGATTCCCG CCTTTTAAC TATGGAAACT GGGAGGTACT TAGGTATCTT 1380
CTCTCAAATG CGAGATGGTG GTGGATGAG TTCAAATTG ATGGATTAG ATTTGATGGT 1440
GTGACATCAA TGATGTATAC TCACCACGGA TTATCGGTGG GATTCACTGG GAACTACGAG 1500
GAATACTTG GACTCGAAC TGATGTGGAT GCTGTTGTGT ATCTGATGCT GGTCAACGAT 1560
CTTATTCTAG GGCTTTCCC AGATGCAATT ACCATTGGTG AAGATGTTAG CGGAATGCCG 1620
ACATTTGTA TTCCCGTTCA AGATGGGGGT GTTGGCTTTG ACTATCGGCT GCATATGGCA 1680
ATTGCTGATA AATGGATTGA GTTGCTCAAG AAACGGGATG AGGATTGGAG AGTGGGTGAT 1740
ATTGTTCTATA CACTGACAAA TAGAAGATGG TCGGAAAAGT GTGTTCTATA CGCTGAAAGT 1800
CATGATCAAG CTCTAGTCGG TGATAAAACT ATAGCATTCT GGCTGATGGA CAAGGATATG 1860
TATGATTITA TGGCTCTGGA TAGACCGCCA ACATCATTAA TAGATCGTGG GATAGCATTG 1920
CACAAGATGA TTAGGCTTGT AACTATGGGA TTAGGAGGGAG AAGGGTACCT AAATTCATG 1980

GGAAATGAAT TCGGCCACCC TGAGTGGATT GATTTCCCTA GGGCTGAACA ACACCTCTCT	2040
GATGACTCAG TAATTCCCGG AAACCAATT AGTTATGATA AATGCAGACG GAGATTTGAC	2100
CTGGGAGATG CAGAATATTT AAGATACCGT GGGTTGCAAG AATTTGACCG GGCTATGCAG	2160
TATCTTGAAG ATAAATATGA GTTTATGACT TCAGAACACC AGTTCATATC ACGAAAGGAT	2220
GAAGGAGATA GGATGATTGT ATTTGAAAAA GGAAACCTAG TTTTGTCTT TAATTTTCAC	2280
TGGACAAAAA GCTATTAGA CTATCGCATA GGCTGCCTGA AGCCTGGAAA ATACAAGGTT	2340
GCCTTGGACT CAGATGATCC ACTTTTGTT GGCTTCGGGA GAATTGATCA TAATGCCGAA	2400
TATTCACCT TTGAAGGATG GTATGATGAT CGTCCTCGTT CAATTATGGT GTATGCACCT	2460
TGTAGAACAG CAGTGGTCTA TGCACTAGTA GACAAAGAAG AAGAAGAAGA AGAAGAAGAA	2520
GAAGAAGTAG CAGTAGTADA AGAAGTAGTA GTAGAAGAAG AATGAACGAA CTTGTG	2576

(2) INFORMATION FOR SEQ ID NO: 17:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2529 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:

GGATGCTAAT GTTTCTGTAT TCTTGAAAAA GCACTCTCTT TCACGGAAGA TCTTGGCTGA	60
AAAGTCTTCT TACAATTCCG AATCCCGACC TTCTACAGTT GCAGCATCGG GGAAAGTCCT	120
TGTGCCTGGA AYCCAGAGTG ATAGCTCCTC ATCCTCAACA GACCAATTG AGTTCACTGA	180
GACATCTCCA GAAAATTCCC CAGCATCAAC TGATGTAGAT AGTTCAACAA TGGAACACGC	240
TAGCCAGATT AAAACTGAGA ACGATGACGT TGAGCCGTCA AGTGTCTTA CAGGAAGTGT	300
TGAAGAGCTG GATTTTGCTT CATCACTACA ACTACAAGAA GGTGGTAAAC TGGAGGAGTC	360
TAAAACATTA AATACTTCTG AAGAGACAAT TATTGATGAA TCTGATAGGA TCAGAGAGAG	420
GGGCATCCCT CCACCTGGAC TTGGTCAGAA GATTTATGAA ATAGACCCCC TTTTGACAAA	480
CTATCGTCAA CACCTTGATT ACAGGTATT ACAGTACAAG AAACTGAGGG AGGCAATTGA	540
CAAGTATGAG GGTGGTTGG AAGCTTTTC TCGTGGTTAT GAAAAAAATGG GTTTCACTCG	600
TAGTGCTACA GGTATCACTT ACCGTGAGTG GGCTCCTGGT GCCCAGTCAG CTGCCCTCAT	660
TGGAGATTTC AACAAATTGGG ACGCAAATGC TGACATTATG ACTCGGAATG AATTTGGTGT	720
CTGGGAGATT TTTCTGCCAA ATAATGTGGA TGGTTCTCCT GCAATTCCCTC ATGGGTCCAG	780

AGTGAAGATA CGYATGGACA CTCCATCAGG TGTAAAGGAT TCCATTCTG CTTGGATCAA	840
CTACTCTTA CAGCTTCCTG ATGAAATTCC ATATAATGGA ATATATTATG ATCCACCCGA	900
AGAGGAGAGG TATRTCTTCC AACACCCACG GCCAAAGAAA CCAAAGTCGC TGAGAATATA	960
TGAATCTCAT ATTGGAATGA GTAGTCCGGA GCCTAAAATT AACTCATAACG TGAATTTAG	1020
AGATGAAGTT CTTCCTCGCA TAAAAAASCT TGGGTACAAT GCGGTGCAA TTATGGCTAT	1080
TCAAGAGCAT TCTTATTATG CTAGTTTGG TTATCATGTC ACAAAATTTT TTGCACCAAG	1140
CAGCCGTTT GGAACGCCCG ACGACCTAA GTCTTGATT GATAAAGCTC ATGAGCTAGG	1200
AATTGTTGTT CTCATGGACA TTGTTCACAG CCATGCATCA AATAATACCT TAGATGGACT	1260
GAACATGTTT GACGGCACAG ATAGTTGTTA CTTTCACTCT GGAGCTCGTG GTTATCATTG	1320
GATGTGGGAT TCCCAGCTCT TTAACATATGG AAACCTGGGAG GTACTTAGGT ATCTTCTCTC	1380
AAATGCGAGA TGGTGGTTGG ATGAGTTCAA ATTTGATGGA TTTAGATTTG ATGGTGTGAC	1440
ATCAATGATG TATACTCACC ACGGATTATC GGTGGGATTG ACTGGGAACG ACGAGGAATA	1500
CTTGGACTC GCAACTGATG TGGATGCTGT TGTGTATCTG ATGCTGGTCA ACGATCTTAT	1560
TCACGGGCTT TTCCCAGATG CAATTACCAT TGGTGAAGAT GTAGCGGAA TGCCGACATT	1620
TTGTATTCCC GTTCAAGATG GGGGTGGTGG CTTTGAATAT CGGCTGCATA TGGCAATTGC	1680
TGATAATGG ATTGAGTTGC TCAAGAACG GGATGAGGAT TGGAGAGTGG GTGATATTGT	1740
TCATACACTG ACAAAATAGAA GATGGTCGGA AAAGTGTGTT TCATMCGCTG AAAGTCATGA	1800
TCAAGCTCTA GTCGGTGATA AAACTATAGC ATYCTGGCTG ATGGACAAGG ATATGTATGA	1860
TTTTATGGCT CTGGATAGAC CGYCAACAYC ATTAATAGAT CGTGGGATAG CATTGCACAA	1920
GATGATTAGG CTTGTAACTA TGGGATTAGG AGGAGAAGGG TACCTAAATT TCATGGGAAA	1980
TGAATTGGC CACCTGAGT GGATTGATT CCCTAGGGCT GARCAACACC TCTCTGATGG	2040
CTCAGTAATT CCCGGAAACC AATTCACTTA TGATAATGC AGACGGAGAT TTGACCTGGG	2100
AGATGCAGAA TATTTAAGAT ACCATGGGTT GCAAGAATTG GACCGGGCTA TGCACTATCT	2160
TGAAGATAAA TATGAGTTA TGACTTCAGA ACACCAAGTC ATATCACGAA AGGATGAAGG	2220
AGATAGGATG ATTGTATTTG AAARAGGAAA CCTAGTTTT GTCTTAATT TTCACTGGAC	2280
AAATAGCTAT TCAGACTATC GCATAGGCTG CCTGAAGCCT GGAAAATACA AGGTTGGCTT	2340
GGACTCAGAT GATCCACTTT TTGGTGGCTT CGGGAGAATT GATCATAATG CCGAATATT	2400
CACCTCTGAA GGATCGTATG ATGATCGTCC TCGTTCAATT ATGGTGTATG CACCTAGTAG	2460

AACAGCAGTG GTCTATGCAC TAGTAGACAA ANTAGAAGNA GAAGAAGAAG AAGAACCGN	2520
NGAAGAATT	2529

(2) INFORMATION FOR SEQ ID NO: 18:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 3231 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:

GATTTAATAC GACTCACTAT AGGGATTTT TTTTTTTTTT TTTTAAAAAC CTCCCTCCACT	60
CAGTCTGGG ATCTCTCTCT CTCTTCACGC TTCTCTGGG GCCTTGAACt CAGCAATTG	120
ACACTCAGTT AGTTACACTC CTATCACTCA TCAGATCTCT ATTttttCTC TTAATTCCAA	180
CCAAGGAATG AATTAAAAGA TTAGATTTGA AGGAGAGAAG AAGAAAGATG GTGTATACAC	240
TCTCTGGAGT TCGTTTCCt ACTGTTCCAT CAGTGTACAA ATCTAATGGA TTCAGCAGTA	300
ATGGTGATCG GAGGAATGCT AATGTTCTG TATTCTGAA AAAGCACTCT CTTTCACGGA	360
AGATCTTGGC TGAAAAGTCT TCTTACGATT CCGAATCCCG ACCTTCTACA GTTGCAGCAT	420
CGGGGAAAGT CCTTGTACCT GGAATCCAGA GTGATAGCTC CTCATCCTCA ACAGACCAAT	480
TTGAGTTCAC TGAGACAGCT CCAGAAAATT CCCCAGCATH AACTGATGTG GATAGTTCAA	540
CAATGGAACA CGCTAGCCAG ATTAAAATG AGAACGATGA CGTTGAGCCG TCAAGTGATC	600
TTACAGGAAG TGTTGAAGAG TTGGATTTG CTTCATCACT ACAACTACAA GAAGGTGGTA	660
AACTGGAGGA GTCTAAAACA TTAAATACTT CTGAAGAGAC AATTATTGAT GAATCTGATA	720
GGATCAGAGA GAGGGGCATC CCTCCACCTG GACTTGGTCA GAAGATTTAT GAAATAGACC	780
CCCTTTGAC AAACTATCGT CAACACCTTG ATTACAGGTa TTCACAGTAC AAGAAAATGA	840
GGGAGGCAAT TGACAAGTAT GAGGGTGGTT TGGAAGCTTT TTCTCGTGGT TATGAAAAAA	900
TGGGTTTCAC TCGTAGTGCT ACAGGTATCA CTTACCGTGA GTGGGCTCCT GGTGCCAGT	960
CAGCTGCTCT CATTGGAGAT TTCAACAATT GGGACGCAA TGCTGACATT ATGACTCGGA	1020
ATGAATTGG TGTCTGGGAG ATTtTTCTGC CAAATAATGT GGATGGTTCT CCTGCAATTc	1080
CTCATGGTC CAGAGTGAAG ATACGCATGG ACACHTCATC AGGTGTTAAG GATTCCATTc	1140
CTGCTTGGAT CAACTACTCT TTACAGCTTC CTGATGAAAT TCCATATAAT GGAATATATT	1200
ATGATCCACC CGAAGAGGAG AGGTATGTCT TCCAACACCC ACGGCCAAAG AAACCAAAGT	1260

CGCTGAGAAT ATATGAATCT CATATTGGAA TGAGTAGTCC GGAGCCTAAA ATTAACATCAT	1320
ACGTGAATT TAGAGATGAA GTTCTTCCTC GCATAAAAAA CCTTGGGTAC AATGCGGTGC	1380
AAATTATGGC TATTCAAGAG CATTCTTATT ATGCTAGTTT TGGTTATCAT GTCACAAATT	1440
TTTTGCACC AAGCAGCCGT TTTGGAACGC CCGACGACCT TAAGTCTTG ATTGATAAAG	1500
CTCATGAGCT AGGAATTGTT GTTCTCATGG ACATTGTTCA CAGCCATGCA TCAAATAATA	1560
CTTTAGATGG ACTGAACATG TTTGACGGCA CAGATAGTTG TTACTTCAC TCTGGAGCTC	1620
GTGGTTATCA TTGGATGTGG GATTCCGCC TCTTTAACTA TGGAAACTGG GAGGTACTTA	1680
GGTATCTTCT CTCAAATGCG AGATGGTGGT TGGATGAGTG CAAATTTGRT GGATTTAGAT	1740
TTGATGGTGT GACATCAATG ATGTATACTC ACCACGGATT ATCGGTGGGA TTCACTGGGA	1800
ACTACGAGGA ATACTTGGA CTCGCAACTG ATGTRGATGC TGCGTGTAT CTGATGCTGG	1860
CCAACGATCT TATTGATGGG CTTTCCCAG ATGCAATTAC CATTGGTCAA GATGTTAGCG	1920
GAATGCCGAC ATTTTGATT CCCGTTCAAG ATGGGGGTGT TGGCTTGAC TATCGGCTGC	1980
ATATGGCAAT TGCTGATAAA TGGATTGAGT TGCTCAAGAA ACAGGGATGAG GATTGGAGAG	2040
TGGGTGATAT TGTTCATACA CTGACAAATA GAAGATGGTC GGAAAAGTGT GTTTCATACG	2100
CTGAAAGTCA TGATCAAGCT CTAGTCGGTG ATAAAACAT ATGCAATTAC CATTGGTCAA GATGTTAGCG	2160
AGGATATGTA TGATTTATG GCTTGGATA GACCGTCAAC ATCATTAAATA GATCGTGGGA	2220
TAGCATTGCA CAAGATGATT AGGCTTGAA CTATGGGATT AGGAGGAGAA GGGTACCTAA	2280
ATTCATGGG AAATGAATTC GGCCACCCCTG AGTGGATTGA TTTCCTAGG GCTGAACAAC	2340
ACCTCTCTGA TGGCTCAGTA ATTCCCGGAA ACCAATTCAAG TTATGATAAA TGCAGACGGAA	2400
GATTTGACCT GGGAGATGCA GAATATTTAA GATACCGTGG GTTGCAAGAA TTTGACCGGG	2460
CTATGCAGTA TCTTGAAAGAT AAATATGAGT TTATGACTTC AGAACACCAG TTCATATCAC	2520
GAAAGGATGTA AGGAGATAGG ATGATTGTAT TTGAAAAAGG AACACCTAGTT TTTGTCTTTA	2580
ATTTCACTG GACAAAAGC TATTCAACT ATCGCATAGG CTGGCTGAAG CCTGGAAAAT	2640
ACAAGGTTGC CTTGGACTCA GATGATCCAC TTTTTGGTGG CTTCGGGAGA ATTGATCATA	2700
ATGCCGAATG TTTCACCTTT GAAGGGATGGT ATGATGATCG TCCTCGTTCA ATTATGGTGT	2760
ATGCCACCTAG TAGAACAGCA GTGGTCTATG CACTAGTAGA CAAAGAAGAA GAAGAAGAAG	2820
AAGTAGCAGT AGTAGAAGAA GTAGTAGTAG AAGAAGAATG AACGAACCTTG TGATCGCGTT	2880
GAAAGATTTG AACGCTACAT AGAGCTTCTT GACGTATCTG GCAATATTGC ATCAGTCTTG	2940

GCGGAATTC ATGTGACAAA AGGTTGCAA TTCTTCCAC TATTAGTAGT GCAACGATAT	3000
ACGCAGAGAT GAAGTGCTGA ACAAACATAT GTAAAATCGA TGAATTATG TCGAATGCTG	3060
GGACGGGCTT CAGCAGGTTT TGCTTAGTGA GTTCTGTAAA TTGTCATCTC TTTANATGTA	3120
CAGCCCCTA GAAATCAATT ATGTGAGACC TAAAAAAACAA TAACCATAAA ATGGAAATAG	3180
TGCTGATCTA ATGATGTTT AANCCNNNA AAAAAAAAAA AAAAACTCGA G	3231

(2) INFORMATION FOR SEQ ID NO: 19:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 2578 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:

TCATTAAAAGA GGAGAAATTA ACTATGAGAG GATCTCACCA TCACCATCAC CATGGGATCT	60
TGGCTGAAAA GTCTTCTTAC AATTCCGAAT TCCGACCTTC TACAGTTGCA GCATCGGGGA	120
AAGTCCTTGT GCCTGGAACC CAGAGTGATA GCTCCTCATC CTCAACAAAC CAATTTGAGT	180
TCACTGAGAC ATCTCCAGAA AATTCCCCAG CATCAACTGA TGTAGATAGT TCAACAATGG	240
AACACGCTAG CCAGATTAAA ACTGAGAACG ATGACGTTGA GCCGTCAGT GATCTTACAG	300
GAAGTGTGTA AGAGCTGGAT TTTGCTTCAT CACTACAAC ACAAGAAGGT GGTAAACTGG	360
AGGAGTCTAA AACATTAAAT ACTTCTGAAG AGACAATTAT TGATGAATCT GATAGGATCA	420
GAGAGAGGGG CATCCCTCCA CCTGGACTTG GTCAGAAAGAT TTATGAAATA GACCCCTTT	480
TGACAAACTA TCGTCAACAC CTTGATTACA GGTATTACCA GTACAAGAAA CTGAGGGAGG	540
CAATTGACAA GTATGAGGGT GGTTTGGAAAG CTTTTCTCG TGGTTATGAA AAAATGGGTT	600
TCACTCGTAG TGCTACAGGT ATCACTTACC GTGAGTGGGC TCCTGGTGCC CAGTCAGCTG	660
CCCTCATTGG AGATTCAAC AATTGGGACG CAAATGCTGA CATTATGACT CGGAATGAAT	720
TTGGTGTCTG GGAGATTTT CTGCCAAATA ATGTGGATGG TTCTCCTGCA ATTCCCTCATG	780
GGTCCAGAGT GAAGATACGT ATGGACACTC CATCAGGTGT TAAGGATTCC ATTCCCTGCTT	840
GGATCAACTA CTCTTCACAG CTTCCTGATG AAATTCCATA TAATGGAATA TATTATGATC	900
CACCCGAAGA GGAGAGGTAT ATCTTCCAAC ACCCACGGCC AAAGAAACCA AAGTCGCTGA	960
GAATATATGA ATCTCATATT GGAATGAGTA GTCCGGAGCC TAAAATTAAC TCATACGTGA	1020
ATTTAGAGA TGAAGTTCTT CCTCGCATAA AAAAGCTTGG GTACAATGCG GTGCAAATTA	1080

TGGCTATTCA AGAGCATTCT TATTATGCTA GTTTGGTTA TCATGTCACA AATTTTTTG	1140
CACCAAGCAG CCGTTTGGA ACGCCCGACG ACCTTAAGTC TTTGATTGAT AAAGCTCATG	1200
AGCTAGGAAT TGTTGTTCTC ATGGACATTG TTCACAGCCA TGCATCAAAT AATACTTAG	1260
ATGGACTGAA CATGTTGAC GGCACCGATA GTTGTTACTT TCACTCTGGA GCTCGTGGTT	1320
ATCATTGGAT GTGGGATTCC CGCCTTTTA ACTATGGAAA CTGGGAGGTA CTTAGGTATC	1380
TTCTCTCAA TGCGAGATGG TGGTTGGATG AGTTCAAATT TGATGGATT AGATTTGATG	1440
GTGTGACATC AATGATGTAT ACTCACCAACG GATTATCGGT GGGATTCACT GGGAACTACG	1500
AGGAATACTT TGGACTCGCA ACTGATGTGG ATGCTGTTGT GTATCTGATG CTGGTCAACG	1560
ATCTTATTCA TGGGCTTTTC CCAGATGCAA TTACCATTGG TGAAGATGTT AGCGGAATGC	1620
CGACATTTG TATTCCCCTT CAAGATGGGG GTGTTGGCTT TGACTATCGG CTGCATATGG	1680
CAATTGCTGA TAAATGGATT GAGTTGCTCA AGAACCGGGA TGAGGATTGG AGAGTGGGTG	1740
ATATTGTTCA TACACTGACA AATAGAAGAT GGTCGGAAAA GTGTGTTCA TACGCTGAAA	1800
GTCATGATCA AGCTCTAGTC GGTGATAAAA CTATAGCATT CTGGCTGATG GACAAGGATA	1860
TGTATGATTT TATGGCTCTG GATAGACCGC CAACATCATT AATAGATCGT GGGATAGCAT	1920
TGCACAAGAT GATTAGGCTT GTAATATGG GATTAGGAGG AGAAGGGTAC CTAATTTC	1980
TGGGAAATGA ATTGGCCAC CCTGAGTGGA TTGATTTCCC TAGGGCTGAA CAACACCTCT	2040
CTGATGACTC AGTAATTCCC GGAAACCAAT TCAGTTATGA TAAATGCAGA CGGAGATTG	2100
ACCTGGGAGA TGCAGAATAT TTAAGATACC GTGGGTTGCA AGAATTGAC CGGGCTATGC	2160
AGTATCTTGA AGATAAATAT GAGTTTATGA CTTCAGAACCA CCAGTTCATC TCACGAAAGG	2220
ATGAAGGAGA TAGGATGATT GTATTGAAA AAGGAAACCT AGTTTTGTC TTTAATTTC	2280
ACTGGACAAA AAGCTATTCA GACTATCGCA TAGGCTGCCT GAAGCCTGGA AAATACAAGG	2340
TTGCCCTGGGA CTCAGATGAT CCACTTTTG GTGGCTTCGG GAGAATTGAT CATAATGCCG	2400
AATATTTCAC CTTTGAAGGA TGGTATGATG ATCGTCCTCG TTCAATTATG GTGTATGCAC	2460
CTTGTAGAAC AGCAGTGGTC TATGCACTAG TAGACAAAGA AGAAGAAGAA GAAGAAGAAG	2520
AAGAAGAAGT AGCAGTAGTA GAAGAAGTAG TAGTAGAAGA AGAATGAACG AACATTGTG	2578

(2) INFORMATION FOR SEQ ID NO: 20:

- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 23 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

AATTTYATGG GNAAYGARTT YGG

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CLAIMS

1. Starch extracted from a potato plant and having an amylose content of at least 35%, as judged by the iodometric assay method of Morrison & Laignelet (1983 J. Cereal Science 1, 9-20).
2. Starch according to claim 1, having an amylose content of at least 37%, as judged by the method defined in claim 1.
3. Starch according to claim 1, having an amylose content of at least 40%, as judged by the method defined in claim 1.
4. Starch according to claim 1, having an amylose content of at least 50%, as judged by the method defined in claim 1.
5. Starch according to claim 1, having an amylose content of at least 66%, as judged by the method defined in claim 1.
6. Starch according to any one of claims 1-5, having an amylose content of 35 - 66%, as judged by the method defined in claim 1.
7. Starch which as extracted from a potato plant by wet milling at ambient temperature has a viscosity onset temperature in the range 70 - 95°C, as judged by viscoamylograph of a 10% w/w aqueous suspension thereof, performed at atmospheric pressure using the Newport Scientific Rapid Visco Analyser 3C with a heating profile of holding at 50°C for 2 minutes, heating from 50 to 95°C at a rate of 1.5°C per minute, holding at 95°C for 15 minutes, cooling from 95 to 50°C at a rate of 1.5°C per minute, and then holding at 50°C for 15 minutes.
8. Starch which as extracted from a potato plant by wet milling at ambient temperature has peak viscosity in the range 500 - 12 stirring number units (SNUs), as judged by viscoamylograph conducted according to the protocol defined in claim 7.

9. Starch which as extracted from a potato plant by wet milling at ambient temperature has a pasting viscosity in the range 214 - 434 SNUs, as judged by viscoamylograph conducted according to the protocol defined in claim 7.
10. Starch which as extracted from a potato plant by wet milling at ambient temperature has a set-back viscosity in the range 450 - 618 SNUs, as judged by viscoamylograph conducted according to the protocol defined in claim 7.
11. Starch which as extracted from a potato plant by wet milling at ambient temperature has a set-back viscosity in the range 14 - 192 SNUs, as judged by viscoamylograph conducted according to the protocol defined in claim 7.
12. Starch which as extracted from a potato plant by wet milling at ambient temperature has a peak viscosity in the range 200 - 500 SNUs and a set-back viscosity in the range 275-618 SNUs as judged by viscoamylograph according to the protocol defined in claim 7.
13. Starch which as extracted from a potato plant by wet milling at ambient temperature has a viscosity which does not decrease between the start of the heating phase (step 2) and the start of the final holding phase (step 5) and has a set-back viscosity of 303 SNUs or less as judged by viscoamylograph according to the protocol defined in claim 7.
14. Starch which as extracted from a potato plant by wet milling at ambient temperature displays no significant increase in viscosity as judged by viscoamylograph conducted according to the protocol defined in claim 7.
15. Starch which as extracted from a potato plant by wet milling at ambient temperature, is in accordance with claim 7 and in accordance with any one of claims 8 to 14.
16. Starch according to any one of claims 7 to 15, having an amylose content in the range 35 - 66%, as judged by the method of Morrison & Laignelet defined in claim 1.

17. Starch which as extracted from a potato plant, has a phosphorus content in excess of 200mg/100grams dry weight starch.
18. Starch according to claim 17, having a phosphorus content in the range 200 - 240mg/100grams dry weight starch.
19. Starch according to claim 17 or 18, further in accordance with any one of claims 1 to 16.
20. Starch prepared by physical, chemical and/or enzymatic treatment of a starch initially having properties in accordance with any one of claims 1-19.
21. Starch according to claim 20, being resistant starch prepared by physical, chemical and/or enzymatic treatment of a starch initially having properties in accordance with any one of claims 1-19.
22. Starch according to claim 21, comprising in excess of 5% total dietary fibre, as determined according to the method of Prosky *et al.*, (1985 J. Assoc. Off. Anal. Chem. 68, 677).
23. Use of starch according to any one of claims 1-22 in the preparation or processing of a foodstuff.
24. Use of starch according to claim 23, wherein the starch is used to provide a film, barrier, coating or as a gelling agent.
25. Use of starch according to claim 23, to prepare resistant starch compositions.
26. Use of starch according to any one of claims 1-22 in the preparation or processing of corrugating adhesives, biodegradable products, packaging, glass fibers and textiles.
27. A nucleotide sequence encoding an effective portion of a class A starch branching

enzyme (SBE) obtainable from potato plants.

28. A nucleotide sequence according to claim 27, encoding a polypeptide comprising substantially the amino acid sequence of residues 49 to 882 of the sequence shown in Figure 5.
29. A nucleotide sequence according to claim 27 or 28, comprising substantially the sequence of nucleotides 289 to 2790 of the sequence shown in Figure 5, or a functional equivalent thereof.
30. A nucleotide sequence according to claim 29, further comprising the sequence of nucleotides 145 to 288 of the sequence shown in Figure 5, or a functional equivalent thereof.
31. A nucleotide sequence according to claim 27, comprising the sequence of nucleotides 228 to 2855 of the sequence labelled psbe2con.seq in Figure 8, or a functional equivalent thereof.
32. A nucleotide sequence according to claim 27, comprising the sequence of nucleotides 57 to 2564 of the sequence labelled as psbe2con.seq in Figure 12, or a functional equivalent thereof.
33. A nucleotide sequence according to any one of claims 27 to 32, comprising an in-frame ATG start codon, and optionally including a 5' and/or a 3' untranslated region.
34. A nucleotide sequence according to claim 27, comprising the sequence of nucleotides 45 to 3200 of the sequence labelled as psbe2con.seq in Figure 8, or a functional equivalent thereof.
35. A nucleic acid construct comprising a sequence in accordance with any one of claims 27 to 34.

36. An expression vector comprising a nucleic acid construct according to claim 35.
37. A host cell into which has been introduced a sequence in accordance with any one of claims 27 to 36.
38. An effective portion of a class A SBE polypeptide obtainable from potato plants and encoded by a nucleotide sequence in accordance with any one of claims 27 to 36.
39. A polypeptide according to claim 38, comprising substantially the sequence of amino acids 49 to 882 of the sequence shown in Figure 5, or a functional equivalent thereof.
40. A polypeptide according to claim 38 or 39, comprising the sequence of amino acids 1 to 48 of the sequence shown in Figure 5.
41. A polypeptide in accordance with any one of claims 38, 39 or 40 in substantial isolation from other plant-derived constituents.
42. A method of altering the characteristics of a plant, comprising introducing into the plant a portion of a nucleotide sequence in accordance with any one of claims 27 to 36, operably linked to a suitable promoter active in the plant, so as to affect the expression of a gene present in the plant.
43. A method according to claim 42, wherein the nucleotide sequence is operably linked in the anti-sense orientation to a suitable promoter active in the plant.
44. A method according to claim 42, wherein the introduced sequence comprises one or more of the following operably linked in the sense orientation to a promoter active in the plant, so as to cause sense suppression of an enzyme naturally expressed in the plant: a 5' untranslated region, a 3' untranslated region, or a coding region of the potato SBE class A starch branching enzyme.
45. A method according to any one of claims 42, 43 or 44, further comprising

introducing into the plant one or more further sequences.

46. A method according to claim 45, wherein one or more of the further sequences are operably linked in the anti-sense orientation to a suitable promoter active in the plant.

47. A method according to claim 45 or 46, wherein the further sequence comprises a portion of a class B SBE nucleotide sequence.

48. A method according to any one of claims 42 to 47, effective in altering the starch composition of a plant.

49. A plant or plant cell having characteristics altered by the method of any one of claims 42 to 48, or the progeny of such a plant, or part of such a plant.

50. A plant according to claim 49, selected from one of the following: potato, pea, tomato, maize, wheat, rice, barley, sweet potato, and cassava.

51. A tuber or other storage organ from a plant according to claim 49 or 50.

52. Use of a tuber or other storage organ according to claim 51, in the preparation and/or processing of a foodstuff.

53. A plant according to claim 49 or 50, containing starch which, as extracted from the plant by wet milling at ambient temperature, has an elevated viscosity onset temperature as judged by viscoamylograph conducted according to the protocol defined in claim 7, compared to starch extracted from a similar, but unaltered, plant.

54. A plant according to claim 53, wherein the viscosity onset temperature is elevated by an amount in the range of 10 to 25°C.

55. A plant according to claim 49 or 50, containing starch which, as extracted from the plant by wet milling at ambient temperature, has a decreased peak viscosity as judged by

viscoamylograph conducted according to the protocol defined in claim 7, compared to starch extracted from a similar, but unaltered, plant.

56. A plant according to claim 55, wherein the peak viscosity is decreased by an amount in the range of 240 to 700 SNUs.

57. A plant according to claim 49 or 50, containing starch which, as extracted from the plant by wet milling at ambient temperature, has an increased pasting viscosity as judged by viscoamylograph conducted according to the protocol defined in claim 7, compared to starch extracted from a similar, but unaltered, plant.

58. A plant according to claim 57, wherein the pasting viscosity is increased by an amount in the range of 37 to 260 SNUs.

59. A plant according to claim 49 or 50, containing starch which, as extracted from the plant by wet milling at ambient temperature, has an increased set-back viscosity as judged by viscoamylograph conducted according to the protocol defined in claim 7, compared to starch extracted from a similar, but unaltered, plant.

60. A plant according to claim 59, wherein the set-back viscosity is increased by an amount in the range of 224 to 313 SNUs.

61. A plant according to claim 49 or 50, containing starch which, as extracted from the plant by wet milling at ambient temperature, has a decreased set-back viscosity as judged by viscoamylograph conducted according to the protocol defined in claim 7, compared to starch extracted from a similar, but unaltered, plant.

62. A plant according to claim 49 or 50, containing starch which, as extracted from the plant by wet milling at ambient temperature, has an elevated apparent amylose content as judged by iodometric assay according to the method of Morrison & Laignelet, compared to starch extracted from a similar, but unaltered, plant.

63. A plant according to claim 49 or 50, containing starch which, as extracted from the plant, has a phosphorus content in excess of 200mg/100grams dry weight starch.
64. Starch obtainable from a plant according to any one of claims 49, 50 or 53 - 63.
65. Starch according to claim 64 and further in accordance with any one of claims 1 - 22.
66. A method of modifying starch *in vitro*, comprising treating starch under suitable conditions with an effective amount of a polypeptide in accordance with any one of claims 38 to 41.
67. A potato plant or part thereof which, in its wild type possesses an effective SBE A gene, but which plant has been altered such that there is no effective expression of an SBE A polypeptide within the cells of at least part of the plant.
68. A potato plant according to claim 67, wherein the alteration is effected by a method according to any one of claims 42-48.

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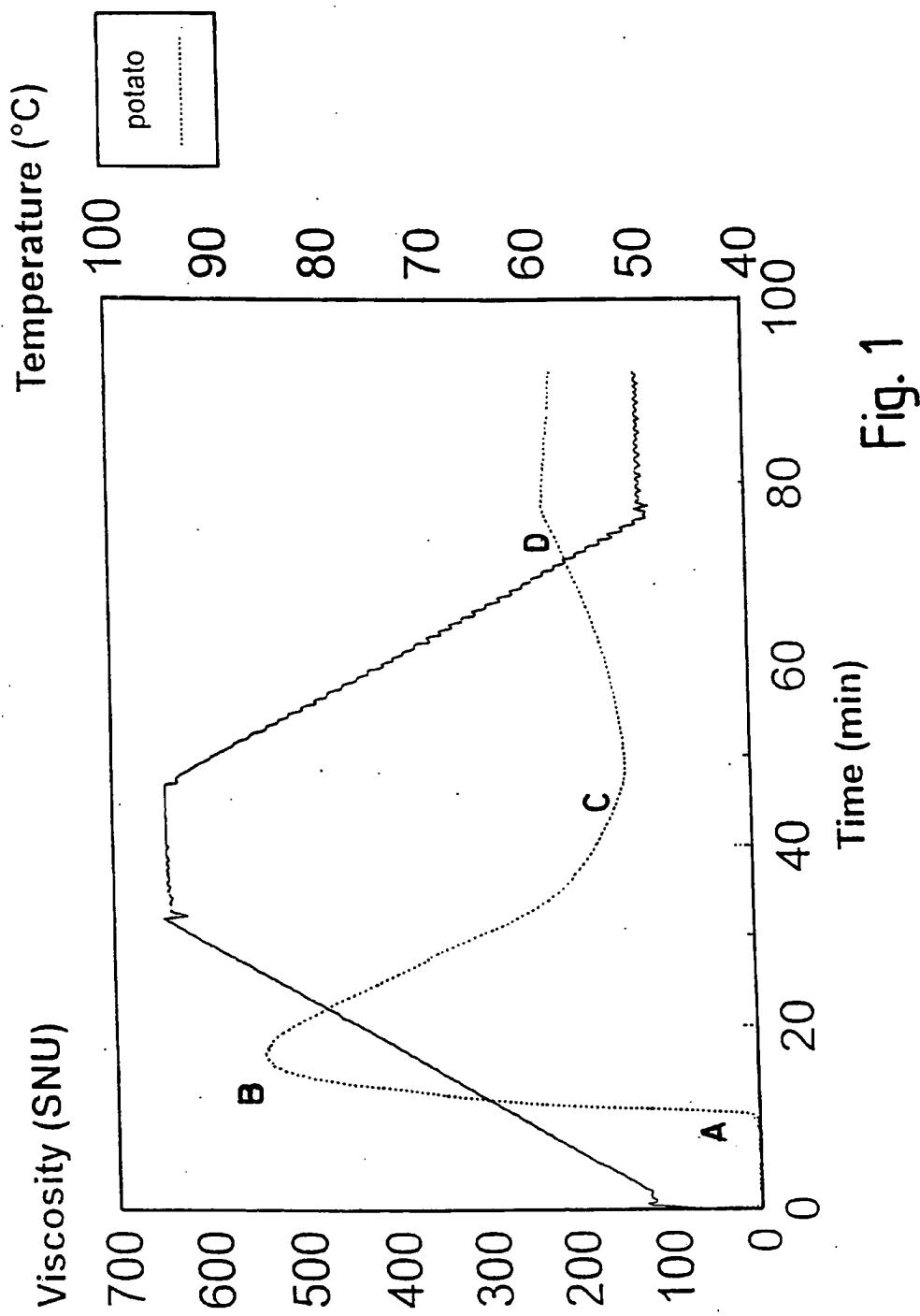


Fig. 1

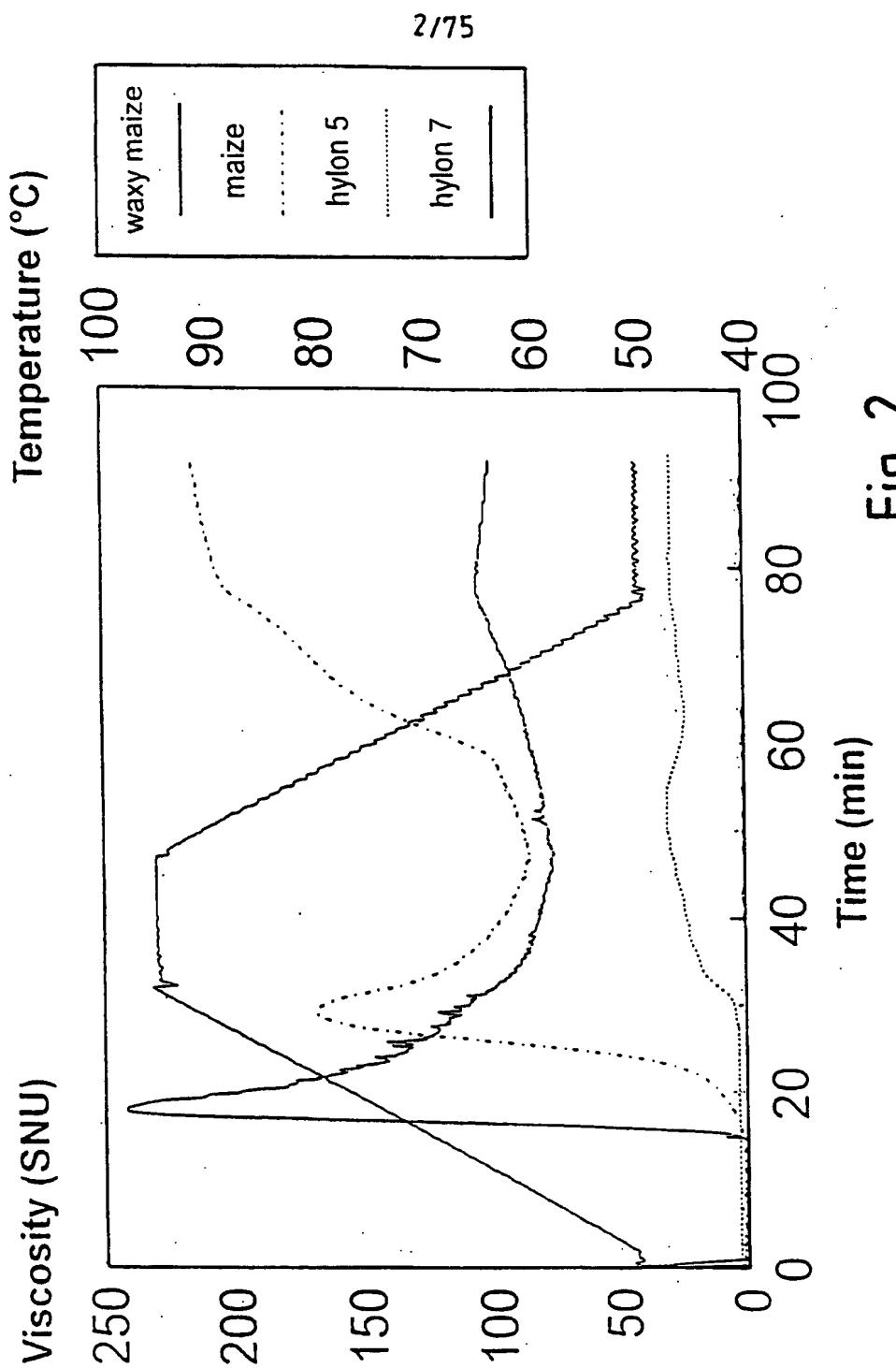


Fig. 2

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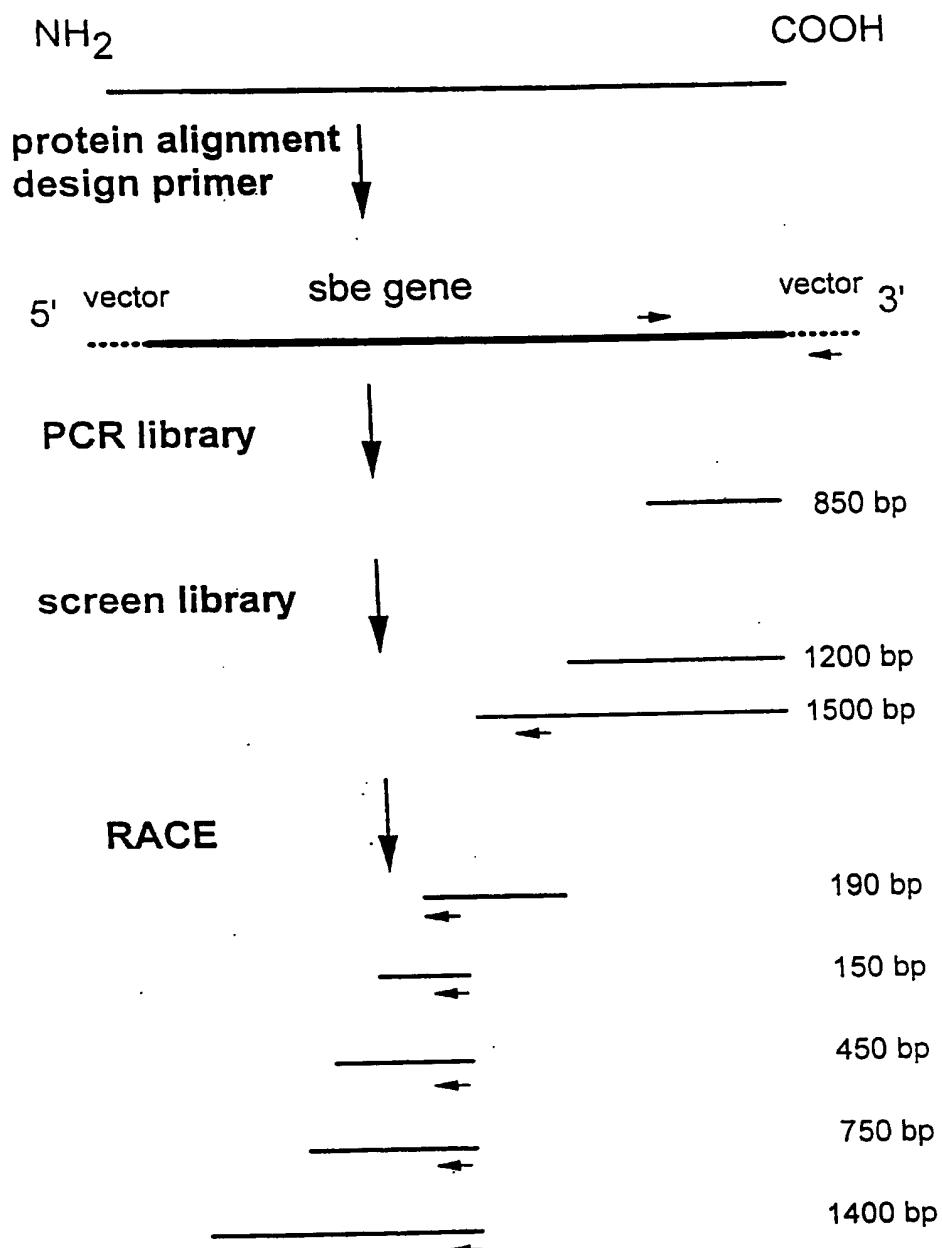


Fig. 3

	Majority P A S P T I D R G I A L H K M I H L I T M G L G G E G Y L N F M G N											
maize 2	P	S	T	P	T	I	D	R	G	I	A	L
pea 1	P	S	T	P	L	I	D	R	G	I	A	H
maize 1	P	A	S	P	T	I	N	R	G	I	A	L
rice 1	P	A	S	P	T	V	V	D	A	G	I	L
potato1	D	A	S	P	V	V	I	D	R	G	I	Q
human	P	F	T	P	V	I	D	R	L	I	T	H
	Majority F S L G D A D . H L R Y K G M N A F D Q A M N A L E E K F S F L A S S											
maize 2	F	D	L	G	D	A	D	Y	H	G	M	Q
pea 1	F	D	L	G	D	A	D	Y	H	G	M	Q
maize 1	W	S	L	V	D	T	D	H	Y	M	N	A
rice 1	W	S	L	V	D	S	E	H	K	F	M	N
potato1	W	N	L	A	D	D	L	H	R	Y	K	F
human	F	H	L	T	D	D	L	L	R	Y	K	F
	Majority K V G C O L P G K Y K V A L D S D A L V F G G H G R V G H D V D H F											
maize 2	R	I	G	C	R	K	P	G	V	Y	K	V
pea 1	K	V	G	C	L	K	P	G	K	Y	K	I
maize 1	K	V	G	C	D	L	P	G	K	Y	R	V
rice 1	K	V	G	C	D	L	P	G	K	Y	R	V
potato1	R	V	G	T	A	L	P	G	K	F	K	I
human	R	V	G	T	A	L	P	G	K	F	K	I

Fig. 4a Sheet 2
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Fig. 4a SHEET 1

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E	F	G	H	P	E	W	I	D	F	P	R	G	P	O	R	L	P	S	G	K	F	I	P	G	N	N	S	Y	D	K	C	R	R	R	666		
E	F	G	H	P	E	W	I	D	F	P	R	G	E	Q	H	L	P	N	G	K	I	V	P	G	N	N	S	Y	D	K	C	R	R	R	713		
E	F	G	H	P	E	W	I	D	F	P	R	G	E	P	R	E	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	624				
E	F	G	H	P	E	W	I	D	F	P	R	G	E	P	R	E	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	618					
E	F	G	H	P	E	W	I	D	F	P	R	G	E	P	R	E	-	-	-	-	-	-	-	-	-	-	-	-	-	-	638						
E	F	G	H	P	E	W	I	D	F	P	R	G	E	P	R	E	-	-	-	-	-	-	-	-	-	-	-	-	-	-	566						
K A I V S D K N E G D K V I V F E R G D L V F N F H P N N S Y E G Y																																					
H	O	Y	I	S	R	K	H	E	E	D	K	V	I	V	F	E	K	G	D	L	V	F	V	F	N	F	H	C	N	N	S	Y	F	D	Y	736	
H	O	Y	I	S	R	K	N	E	G	D	R	V	I	I	F	E	R	D	N	L	V	F	V	F	N	F	H	W	T	N	S	Y	S	D	Y	783	
K	O	Y	I	S	R	M	M	N	E	E	K	V	I	V	F	E	R	G	D	L	V	F	V	F	N	F	H	P	K	T	Y	E	G	Y	694		
K	O	Y	I	S	R	M	M	N	E	K	D	K	V	I	V	F	E	R	G	D	L	V	F	V	F	N	F	H	P	N	K	T	Y	K	G	Y	688
K	O	Y	I	S	R	M	M	N	E	K	D	K	V	I	V	F	E	R	G	D	L	V	F	V	F	N	F	H	P	N	N	T	Y	E	G	Y	708
Q	A	Y	V	S	E	K	H	E	G	N	K	I	I	A	F	E	R	A	G	L	L	F	T	F	N	F	H	P	S	K	S	Y	T	D	Y	636	
T S P E G - P G V P E T N F N N R P N S F K V L S P S R T C V A Y Y R V																																					
T	A	-	-	-	-	-	D	C	S	H	D	N	R	P	Y	S	F	S	V	Y	I	P	S	R	T	C	V	V	Y	A	P	V	798				
T	S	-	-	-	-	-	E	G	W	Y	D	D	R	R	P	R	S	F	L	V	Y	A	P	S	R	T	A	V	V	Y	A	L	A	845			
T	S	P	E	G	Y	P	G	V	P	E	T	N	F	N	F	N	R	P	N	S	F	K	V	L	S	P	P	R	T	C	V	A	Y	Y	R	V	764
T	S	P	E	G	M	P	G	V	P	E	T	N	F	N	R	P	N	S	F	K	V	L	S	P	P	R	T	C	V	A	Y	Y	R	V	758		
T	S	P	E	G	I	P	G	V	P	E	T	N	F	N	G	R	P	N	S	F	K	V	L	S	P	A	R	T	C	V	A	Y	Y	R	V	778	
S	E	-	-	-	-	-	A	F	E	H	N	G	R	P	Y	S	L	L	V	Y	I	P	S	R	V	A	L	I	L	O	N	V	698				

Fig. 4a SHEET 2

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Majority DEXXSE-R-----
Majorize 2 E G V E S P I E L S
pea 1 E G V E S P I E L S
maize 1 H A K A E T G K T S P A E S I D V K A S R A S S K
rice 1 R G G
potato 1 R G G
human 1 R G G

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800
856
824
769
789
703

E D K E A T A G G K K G W K F A R Q P S D O D T K .

Fig. 4a SHEET 3

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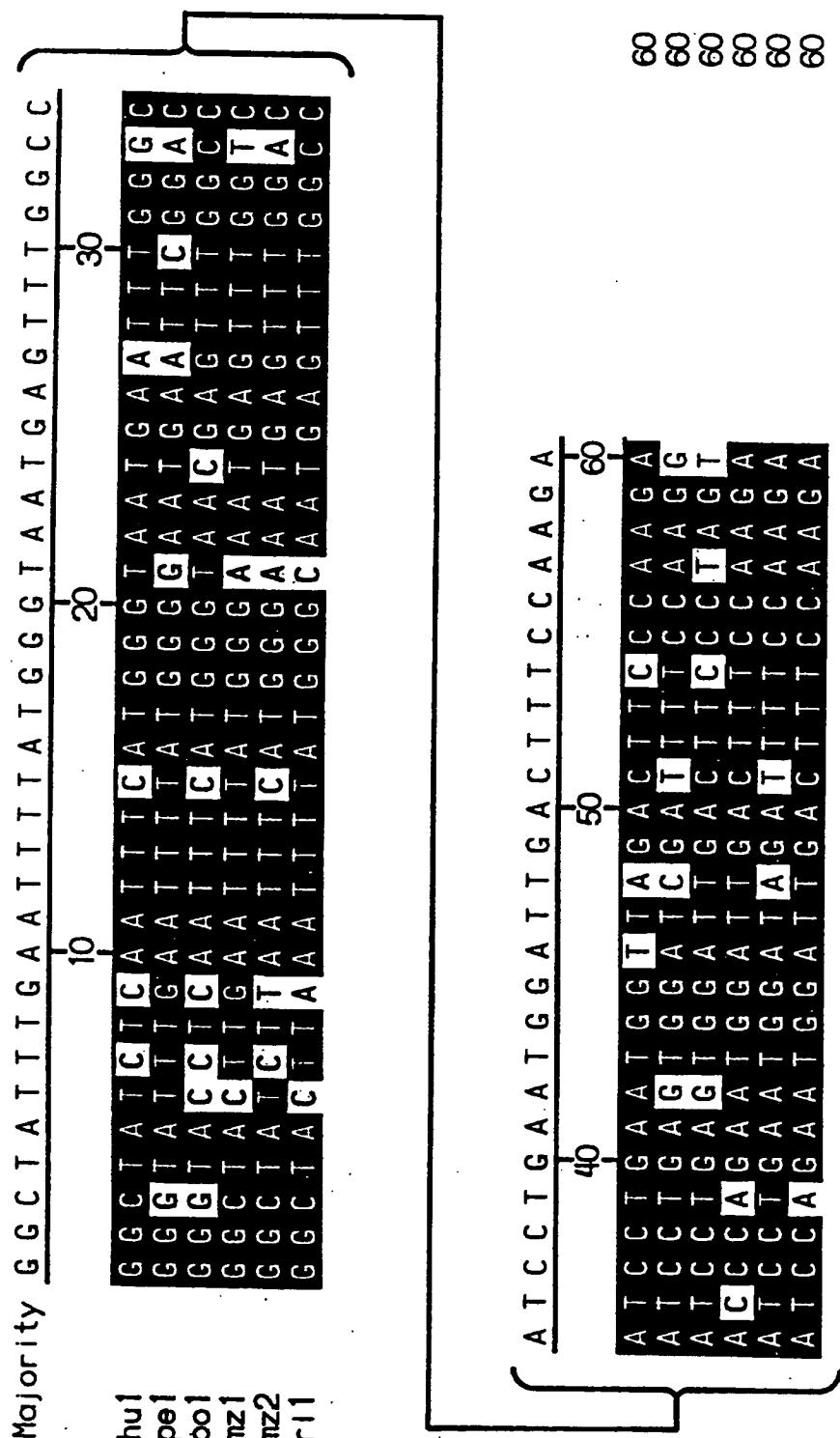


Fig. 4b

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TGTGATGGGGCCTTGAACCTAGCAATTGACACTCAGTTAGTTACA
 AACTACCCCGGAACCTTGAGTCGTTAAACTGTGAGTCATCAATGTA
 AAGGAATGAATAAAAGGATAGATTGTAAAAACCTAAGGAGAGA
 TTCCTTACTTATTTCTATCTAACATTTGGGATTCCCTCTCT
 M N K R I D L
 GTTCCCATCAGTGTACAAATCTAATGGATTCAAGCAGTAATGGTGAT
 CAAGGTAGTCACATGTTAGATTACCTAACGTCGTCAATTACCACTA
 V P S V Y K S N G F S S N G D
 Bgl II EcoR I
 TCACGGAAAGATCTTGGCTGAAAAGTCTTCTTACAATTCCGAATT
 AGTGCCTTCTAGAACCGACTTTCAAGAAGAATGTTAAGGCTTAAG
 S R K I L A E K S S Y N S E F
 ACCCAGAGTGATAGCTCCTCATCCTAACAGACCAATTGAGTTC
 TGGGTCTCACTATCGAGGGAGTAGGAGTTGTCGGTTAAACTCAAG
 T Q S D S S S S T D Q F E F
 AGTTCAACAATGGAACACGCTAGCCAGATTAAACTGAGAACGAT
 TCAAGTTGTTACCTTGTGCGATCGGTCTAACATTGACTCTTGCTA
 S S T M E H A S Q I K T E N D
 GATTTGCTTCACTACAACATAAGAAGGTGGTAAACTGGAG
 CTAAAACGAAGTAGTGATGTTGATGTTCTTCCACCATTGACCTC
 D F A S S L Q L Q E G G K L E

Fig 5
Sheet 2

Fig. 5 SHEET 1

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Bgl II

CTCCTATCACTTATCAGATCTCTATTCTCTTAATTCCAACC
 GAGGATAGTGAATAGTCTAGAGATAAAAAAGAGAATTAGGTTGG 90

AGAAGAAAGATGGTGTATACACTCTCTGGAGTTCGTTTCTACT
 TCTTCTTCTACACATATGTGAGAGACCTCAAGCAAAAGGATGA 180

M V Y T L S G V R F P T

CGGAGGAATGCTAATGTTCTGTATTCTGAAAAAGCACTCTCTT
 GCCTCCTTACGATTACAAAGACATAAGAACCTTTCTGTGAGAGAA 270

R R N A N V S V F L K K H S L

CGACCTTCTACAGTTGCAGCATGGGAAAGTCCTTGTCCTGGAA
 GCTGGAAGATGTCAACGTCGTAGCCCCCTTCAGGAACACGGACCT 360

R P S T V A A S G K V L V P G

ACTGAGACATCTCCAGAAAATTCCCCAGCATCAACTGATGTAGAT
 TGACTCTGTAGAGGTCTTTAAGGGTCTAGTTGACTACATCTA 450

T E T S P E N S P A S T D V D

GACGTTGAGCCGTCAAGTGATCTTACAGGAAGTGTGAAGAGCTG
 CTGCAAACCTGGCAGTTCACTAGAAATGTCCTTCACAACCTCTCGAC 540

D V E P S S D L T G S V E E L

GAGTCTAAAACATTAATCTGAAGAGACAATTATTGATGAA
 CTCAGATTTGTAATTATGAAGACTTCTGTGTTAATAACTACTT 630

E S K T L N T S E E T I I D E

Fig. 5 SHEET 2

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TCTGATAGGATCAGAGAGAGGGCATCCCTCCACCTGGACTTGGT
AGACTATCCTAGTCTCTCCCCGTAGGGAGGTGGACCTGAACCA
S D R I R E R G I P P P P G L G

CACCTTGATTACAGGTATTACACAGTACAAGAAACTGAGGGAGGCA
GTGGAACTAATGTCCATAAGTGTATGTTCTTGACTCCCTCCGT
H L D Y R Y S Q Y K K L R E A

GAAAAAAATGGGTTTCACTCGTAGTGCTACAGGTATCACTTACCGT
CTTTTTACCCAAAGTGAGCATCACGATGTCCATAGTGAATGGCA
E K M G F T R S A T G I T Y R

AACAATTGGGACGCAAATGCTGACATTATGACTCGGAATGAATT
TTGTTAACCTGCCTTACGACTGTAATACTGAGCCTTACTTAAA
N N W D A N A D I M T R N E F

GCAATTCCATGGGTCCAGAGTGAAGATACTGATGGACACTCCA
CGTTAAGGAGTACCCAGGTCTCACTTCTATGCATAACCTGTGAGGT
A I P H G S R V K I R M D T P

Fig.5
Sheet 4

Fig. 5 SHEET 3

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Hinc II

CAGAAGATTATGAAATAGACCCCTTTGACAAACTATCGTCAA
 GTCTTCTAAATACTTATCTGGGGAAAACGTGGATAGCAGTT

Q K I Y E I D P L L T N Y R Q

ATTGACAAGTATGAGGGTGGTTGGAACGCCTTCCTCGTGGTTAT
 TAACCTGTTCATACTCCCACCAAAACCTCGGAAAAGAGCACCAATA

I D K Y E G G L E A F S R G Y

Pvu II

GAGTGGGCTTTGGTGCCCAGTCAGCTGCCCTCATTGGAGATTTC
 CTCACCCGAGAACACGGGTCACTCGACGGGAGTAACCTCTAAAG

E W A L G A Q S A A L I G D F

GGTGTCTGGGAGATTTCTGCCAAATAATGTGGATGGTTCTCCT
 CCACAGACCCCTCTAAAAAGACGGTTATTACACCTACCAAGAGGA

G V W E I F L P N N V D G S P

TCAGGTGTTAAGGATTCCATTCTGCTGGATCAACTACTCTTA
 AGTCCACAAATTCTTAAGGTAAGGACGAACCTAGTTGATGAGAAAT

S G V K D S I P A W I N Y S L

Fig. 5 SHEET 4

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CAGCTTCCTGATGAAATTCCATATAATGGAATACATTATGATCCA
 GTCGAAGGACTACTTTAAGGTATATTACCTTATGTAATACTAGGT
 Q L P D E I P Y N G I H Y D P

CCAAGTCGCTGAGAATATGAATCTCATATTGGAATGAGTAGT
 GGTTTCAGCGACTCTTATATACTTAGAGTATAACCTTACTCATCA
 P K S L R I Y E S H I G M S S

HinD III

CTTCCTCGCATAAAAAAGCTTGGGTACAATGCGCTGCAAATTATG
 GAAGGAGCGTATTTTCGAACCCATGTTACGCGACGTTAACATAC
 L P R I K K L G Y N A L Q I M

ACAAATTTTGACCAAGCAGCCGTTTGGAACGCCGACGAC
 TGTTTAAAAAACGTGGTCGTGGCAAAACCTTGCAGGGCTGCTG
 T N F F A P S S R F G T P D D

CTCATGGACATTGTTCACAGCCATGCATCAAATAACTTTAGAT
 GAGTACCTGTAACAAGTGTGGTACGTAGTTATTATGAAATCTA
 L M D I V H S H A S N N T L D

Fig.5
Sheet
6

Fig. 5 SHEET 5

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CCCGAAGAGGAGAGGTATATCTTCAACACCCACGGCCAAGAAA
1170
GGGCTTCTCCTCTCCATATAGAAGGTTGTGGGTGCCGGTTCTT
P E E E R Y I F Q H P R P K K

Xmn I

CCGGAGCCTAAAATTAACTCATACGTGAATTTAGAGATGAAGTT
1260
GGCCTCGGATTTAATTGAGTATGCACTTAAATCTCTACTTC
P E P K I N S Y V N F R D E V

GCTATTCAAGAGCATTCTTATTACGCTAGTTTGTTATCATGTC
1350
CGATAAGTTCTCGTAAGAATAATGCGATAAAACCAATAGTACAG
A I Q E H S Y Y A S F G Y H V

CTTAAGTCTTGATTGATAAAGCTCATGAGCTAGGAATTGTTGTT
1440
GAATTAGAAAACAACTATTGAGTACTCGATCCTAACCAA
L K S L I D K A H E L G I V V

GGACTGAACATGTTGACTGCACCGATAAGTTGTTACTTCACTCT
1530
CCTGACTTGTACAAACTGACGTGGCTATCAACAAATGAAAGTGAGA
G L N M F D C T D S C Y F H S

Fig. 5 SHEET 6

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Sac I

GGAGCTCGTGGTTATCATTGGATGTGGGATTCCCGCTCTTAAC
 CCTCGAGCACCAATAGTAACCTACACCCCTAAGGGCGGAGAAATTG
 G A R G Y H W M W D S R L F N

TGGTGGTTGGATGCGTTCAAATTGATGGATTAGATTGATGGT
 ACCACCAACCTACGCAAGTTAAACTACCTAAATCTAAACTACCA
 W W L D A F K F D G F R F D G

ACTGGGAACTACGAGGAATACTTTGGACTCGCAACTGATGTGGAT
 TGACCCCTTGATGCTCCTTATGAAACCTGAGCGTTGACTACACCTA
 T G N Y E E Y F G L A T D V D

TTCCCAGATGCAATTACCATTGGTGAAGATGTTAGCGGAATGCCG
 AAGGGTCTACGTTAACGTAACCACTTCTACAATCGCCTACGGC
 F P D A I T I G E D V S G M P

CGGCTGCATATGGCAATTGCTGATAAACGGATTGAGTTGCTCAAG
 GCCGACGTATACCGTTAACGACTATTGCTAACTCAACGAGTTC
 R L H M A I A D K R I E L L K

ACAAATAGAAGATGGTCGGAAAAGTGTGTTCATACGCTGAAAGT
 TGTTTATCTTCTACCGCTTTCACACAAAGTATGCGACTTTCA
 T N R R W S E K C V S Y A E S

Fig 5
Sheet 8

Fig. 5 SHEET 7

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TATGGAAACTGGGAGGTACCTAGGTATCTTCTCTCAAATGCGAGA
 ATACCTTGACCCCTCCATGAATCCATAGAACAGAGAGTTACGCTCT
 Y G N W E V L R Y L L S N A R

1620

GTGACATCAATGATGTATATTACACCACGGATTATCGGTGGGATTC
 CACTGTAGTTACTACATATAAGTGGTGCCTAATAGCCACCCCTAAG
 V T S M M Y I H H G L S V G F

1710

Hinc II

GCTGTTGTATCTGATGCTGGTCAACGATCTTATTATGGGCTT
 CGACAACACATAGACTACGACCAGTTGCTAGAATAAGTACCCGAA
 A V V Y L M L V N D L I H G L

1800

ACATTTGTATTCCCGTCCAAGAGGGGGTGTTGGCTTGACTAT
 TGTAAAACATAAGGGCAGGTTCTCCCCCACAACCGAAACTGATA
 T F C I P V Q E G G V G F D Y

1890

AAACGGGATGAGGATTGGAGAGTGGGTGATATTGTTCATACACTG
 TTTGCCCTACTCCTAACCTCTCACCCACTATAACAAGTATGTGAC
 K R D E D W R V G D I V H T L

1980

CATGATCAAGCTCTAGTCGGTGATAAAACTATAGCATTCTGGCTG
 GTACTAGTTCGAGATCAGCCACTATTTGATATCGTAAGACCGAC
 H D Q A L V G D K T I A F W L

2070

Fig. 5 SHEET 8

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Hinc II

ATGGACAAGGATATGTATGATTATGGCTCTGGATAGACCGTCA	
TACCTGTTCCCTATACATACTAAAATACCGAGACCTATCTGGCACT	
M D K D M Y D F M A L D R P S	

Asp 718
Kpn I

CTTGTAACTATGGGATTAGGAGGAGAAGGGTACCTAAATTCTATG	
GAACATTGATACCCCTAACCTCCTCTTCCCATGGATTAAAGTAC	
L V T M G L G G E G Y L N F M	

GAACAAACACCTCTCTGATGGCTCAGTAATCCCCGGAAACCAATTG	
CTTGTGAGAGACTACCGAGTCATTAGGGGCCTTGGTTAAG	
E Q H L S D G S V I P G N O F	

Ssp I

TATTAAGATAACCGTGGGTTGCAAGAATTGACC GG CCT ATGCAG	
ATAAATTCTATGGCACCCAACGTTCTAAACTGGCCGGATACGTC	
Y L R Y R G L Q E F D R P M Q	

ATATCACGAAAGGATGAAGGAGATAGGATGATTGTATTTGAAAAAA	
TATAGTGCTTCCCTACTTCCTCTATCCTACTAACATAAACTTTT	
I S R K D E G D R M I V F E K	

TCAGACTATCGCATAGCCTGCCCTGAAGCCTGGAAAATACAAGGTT	
AGTCTGATAGCGTATCGGACGGACTTCGGACCTTTATGTTCAA	
S D Y R I A C L K P G K Y K V	

Fig.5
Sheet 10

Fig.5 SHEET 9

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ACATCATTAAATAGATCGTGGGATAGCATTGCACAAGATGATTAGG
 TGTAGTAATTATCTAGCACCCCTATCGTAACGTGTTCTACTAATCC
 T S L I D R G I A L H K M I R

EcoR I

GGAAATGAATT CGGCCAC CCTGAGTGGATTGATTCCCTAGGGCT
 CCTTTACTTAAGCCGGTGGGACTCACCTAACTAAAGGGATCCCGA
 G N E F G H P E W I D F P R A

AGTTATGATAAAATGCAGACGGAGATTGACCTGGAGATGCAGAA
 TCAATACTATTTACGTCTGCCTCTAAACTGGACCCCTACGTCTT
 S Y D K C R R R F D L G D A E

TATCTTGAAGATAAAATGAGTTATGACTTCAGAACACCAGTTC
 ATAGAACTTCTATTTATACTCAAATACTGAAGTCTTGTGGTCAAG
 Y L E D K Y E F M T S E H Q F

GGAAACCTAGTTTGTC TTAATTTCACTGGACAAAAAGCTAT
 CCTTTGGATCAAAAACAGAAATTAAAAGTGAACCTGTTTTCGATA
 G N L V F V F N F H W T K S Y

GCCTTGGACTCAGATGATCCACTTTGGTGGCTTCGGGAGAATT
 CGGAACCTGAGTCTACTAGGTGAAAAACCACCGAAGCCCTCTAA
 A L D S D D P L F G G F G R I

Fig. 5 SHEET 10

SUBSTITUTE SHEET (RULE 26)

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Ssp I

GATCATAATGCCGAATATTCACCTTGAAGGGATGGTATGATGAT
CTAGTATTACGGCTTATAAAGTGGAAACTTCCTACCATACTACTA
D H N A E Y F T F E G W Y D D

GTCTATGCACTAGTAGACAAGAAGAAGAAGAAGAAGAAGAAGAAGAA
CAGATACGTGATCATCTGTTCTTCTTCTTCTTCTTCTTCTTCTTCTT
V Y A L V D K E E E E E E E E E E

TGAACGAACCTGTGATCGCGTTGAAAGATTGAACGCTACATAGA
ACTTGCTTGAACACTAGCGCAACTTCTAAACTTGCGATGTATCT

TCATGTGACACAAGGTTGCAATTCTTCCACTATTAGTAGTGCA
AGTACACTGTGTTCAAACGTTAAGAAAGGTGATAATCATCACGT

EcoR I Pst I

GATGAATTATGTCGAATGCTGGGACGATCGAATT CCTGCAGGCC
CTACTTAAATACAGCTTACGACCCCTGCTAGCTTAAGGACGTCGGG

Fig 5
Sheet
12

Fig. 5 SHEET 11

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CGTCCTCGTTCAATTATGGTGTATGCACCTTGTAAAACAGCAGTG
2700
GCAGGAGCAAGTTAACCAACATACTGGAACATTTGTCGTAC
R P R S I M V Y A P C K T A V

GAAGAAGAAGTAGCAGCAGTAGAAGAAGTAGTAGTAGAAGAAGAA
2790
CTTCTTCTTCATCGTCGTACATCTTCTTCATCATCATCTTCTTCTT
E E E V A A V E E V V V E E E

Ssp I

GCTTCTTGACGTATCTGGCAATATTGCATCAGTCTTGGCGGAATT
2880
CGAAGAACTGCATAGACCGTTATAACGTAGTCAGAACCGCCTTAA

Cla I

ACGATATAACGAGAGATGAAGTGCTGAACAAACATATGTAACATC
2970
TGCTATATGCGTCTACTTCACGACTTGTGTATACATTAG

GGGGGACCCCTTAGTTCT
3033
CCCCCTGGGAATCAAGA

Fig. 5 SHEET 12

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v180 v190 v200 v210 v220
 IYEIDPLL TNÝROHLDYRYSQYKKLREAIKYEGGLEAFSRGYEKMGFTR
 : : DP L. Y : H. R .: Y . : I: KYEG LE. F: : GY K. GF. R
 LLNLDPTEPYLDHFRHRMKRYVDQKMLIEKYEGPLEEFAQGYLKFGFNR
 ^100 ^110 ^120 ^130 ^140
 v230 v240 v250 v260 v270
 SATGITYREWALGAQSAALIGDFNNWDANADIMTRNEFGVWEIFLPNNVD
 . . . I. YREWA : AQ. A. : IGDFN. W. : : : M. : : FGVW. I : P: VD
 EDGCIVYREWAPAAQEAEVIGDFNGWNGSNHMMEKDQFGVWSIRIPD-VD
 ^150 ^160 ^170 ^180 ^190
 v280 v290 v300 v310 v320
 GSPAIPHGSRVKIRMDTPSGV-KDSIPAWINYSLQLPDEI--PYNGIHYD
 : . P. IPH. SRVK: R. . : GV D. IPAWI: Y: . . . PY: G: . D
 SKPVIPHNSRVKFRFKHGNGVWVDRIPAWIKYATADATKFAAPYDGVYWD
 ^200 ^210 ^220 ^230 ^240
 v330 v340 v350 v360 v370
 PPEEERYIFQHPRPKPKSLRIYESHI GMSSPEPKINSYVNFRDEVLPRI
 PP. ERY F. PRP KP: RIYE: H: GMSS: EP: NSY : F D: VLPRI
 PPPSERYHFKYPRPPKPRAPRIYEAHVGMSSEPRVNSYREFADDVLPRI
 ^250 ^260 ^270 ^280 ^290
 v380 v390 v400 v410 v420
 KKLGYNALQIMAIQEHSYYASFGYHVTNFFAPSSRGTPDDLKSLIDKAH
 K . YN: : Q: MAI EHSYY: SFGYHVTNFFA S: R: G. P: DLK LIDKAH
 KANNYNTVQLMAIMEHSYYGSFGYHVTNFFAVSNRYGNPEDLKYLIDKAH
 ^300 ^310 ^320 ^330 ^340
 v430 v440 v450 v460 v470
 ELGI VVLMIDIVHSHASNNTLDGLNMFDCTDSCYFHSGARGYHWMWDS
 . LG: VL: D: VHSHASNNDGLNFD : : : YFH: G. RGYH : WDS
 SLGLQVLVDVVHSHASNNTDGLNGFDIGQGSQESYFHAGERGYHKLWDS
 ^350 ^360 ^370 ^380 ^390
 v480 v490 v500 v510 v520
 RLFNYGNWEVLRYLLSNARWWLDAFKFDGFRTGVTSMMYIHHLGSVGFT
 RLFNY: NWEVLR: LLSN RWWL: . : FDGFRTDG: TSM: Y: HHG: : GFT
 RLFNYANWEVLRFLLSNLRWWLEEYNFDFRFDGITSMLYVHHGINMNGFT
 ^400 ^410 ^420 ^430 ^440
 v530 v540 v550 v560 v570
 GNYEEYFGLATDVDAVVYLMVNDLIHGLFPDAITIGEDVSGMPFCIPV
 GNY: EYF: ATDVDAVVYLM. N: LIH : FPDA. . I: EDVSGMP. . PV
 GNYNEYFSEATDVDAVVYLMLANNLHKIFPDATVIAEDVSGMPGLSRPV
 ^450 ^460 ^470 ^480 ^490
 v580 v590 v600 v610 v620
 QEGGVGF DYRLHMAIADKRIELLK-KRDEDWRVGDIVHTLTNRRWSEKCV
 EGG: GFDYRL MAI: DK: I: LK K. DEDW. : : : LTNRR. : EKC:
 SEGGIGFDYRLAMAIPDKWIDYLKNKNDedWSMKEVTSSLTNRRYTEKC
 ^500 ^510 ^520 ^530 ^540

Fig. 6 SHEET 1

SUBSTITUTE SHEET (RULE 26)

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↗630 ↗640 ↗650 ↗660 ↗670
 SYAESHDQAL VGDKTIAFWLMDKDMYDFMALDRPSTSL IDRGI ALHKMIR
 : YAESHDO: : VGDKTIAF LMDK: MY. M: : : : DRGIALHKMI:
 AYAESHDSQ I VGDKTIAFL LMDKEMSGMSCL TDASP VVDRG I ALHKMIH
 ↗550 ↗560 ↗570 ↗580 ↗590
 ↗680 ↗690 ↗700 ↗710 ↗720
 LVTMGLGGEGYLNFMGNEFGHPEWIDFPRAEQHLSDGSI PGNQFSYDKC
 : TM: LGGEGLNFMGNEFGHPEWIDFPR GN: . SYDKC
 FFTMALGGEGYLNFMGNEFGHPEWIDFPR ----- EGNNWSYDKC
 ↗600 ↗610 ↗620 ↗630
 ↗730 ↗740 ↗750 ↗760 ↗770
 RRRFDLGDAEYLRYRGLQEFDRPMQYLEDKYEFMTSEHQF ISRKDEGDRM
 RR: . : L: D: E: LRY: : : FDR: M: L: : K: . F: : S: . Q: : S: . D: : : :
 RRQWNLADSEHLRYKFMNAFDRAMNSLDEKFSFLASGKQIVSSMDDDNKV
 ↗640 ↗650 ↗660 ↗670 ↗680
 ↗780 ↗790 ↗800 ↗810 ↗820
 IVFEKGNLVFVFNFHWTKSYSYR IACLKPGKYKVALDSDPLFGGFGR I
 : VFE: G: LVFVFNFH . : Y: : Y: : C PGKY: VAL: SD. FGG GR
 VVFERGDLVVFNFHPNNTYEGYKVGCDLPGK YRVALGSDAWEFGGHGRA
 ↗690 ↗700 ↗710 ↗720 ↗730
 ↗830 ↗840 ↗850 ↗860
 DHNAEYFT ----- FEGWYDDRPRSIMVYAPCKTAVVYALVDKEEEEEE
 : H: . : FT E: . : RP. S: . V: P: T V. Y: VD. . E.
 GHDV DHFTSPEGIPGVPETNFNGRPNSFK VLSPARTCVAYYR VDERMSET
 ↗740 ↗750 ↗760 ↗770 ↗780
 ↗870
 EEEEEEV
 E: . : :
 EDYQTDI
 ↗790

Fig. 6 SHEET 2

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f10 f20 f30 f40
 MVYTLSGVRFPTVPSVYKSNGFSSNGDRRNANVSFLKKH--SLSRKILA
 MVYT: SG: RFP.: PS: . KS : . DRR.: : S FLK: : S: SR. L
 MVYTISGIRFPVLPSLHK---TLRCDRRASSHSFFLKNNSSSFRTSLY
 ^10 ^20 ^30 ^40
 f50 f60 f70 f80 f90
 EKSSYNSEFRPSTVAASGKVLPVGTDQSDSSSSSTDQFEFTETSPENSPAS
 . K S : SE : : ST: A. S: KVL: P. . Q D: S S : DQ: E . . . : E: . .
 AKFSRDSETKSSTIAESDKVLIPEPDQ-DNSVSLADQLENPDITSEDAQNL
 ^50 ^60 ^70 ^80 ^90
 f100 f110 f120 f130 f140
 TDVDSSTMHASQIKTENDDVEPSSDLTGSVEELDFASSLQLQEGGKLEE
 . D: TM. : : : : : : : : : : : : : : S : : : : : :
 EDL---TMKDGKYNID-ESTSSYREVGDEKGSVTSSLVDVNTDTQ--A
 ^100 ^110 ^120 ^130 ^140
 f150 f160 f170 f180 f190
 SKTLNTSEETIIDESDRIRERGIPPPGLGQKIYEIDPLLTNYRQHLDYRY
 . KT S: . : . : I IPPPG GQKIYEIDPLL . RQHLD: RY
 KKTSVHSDKVKVVDKPKI---IPPPGSGQKIYEIDPLLQAHRQHLDFRY
 ^150 ^160 ^170 ^180 ^190
 f200 f210 f220 f230 f240
 SQYKKLREAIDKYEGGLEAFSRGYEKMGFTRSATGITYREWALGAQSAAL
 : QYK: : RE. IDKYEGGL: AFSRGYEK. GFTRSATGITYREW: GA: SAAL
 GQYKRIREEIDKYEGGLDAFSRGYEFKGFTRSATGITYREWGPAGAKSAAL
 ^190 ^200 ^210 ^220 ^230
 f250 f260 f270 f280 f290
 1GDFNNWDANADIMTRNEFGWEIFLPNNVDGSPAIPHGSRVKIRMDTPS
 : GDFNNW: : NAD: MT: : FGWEIFLPNN. DGSP: IPHGSRVKI: MDTPS
 VGDFFNNWNPNADEVMTKDAFGWEIFLPNNADGSPPIPHGSRVKIHMDDTPS
 ^240 ^250 ^260 ^270 ^280
 f300 f310 f320 f330 f340
 GVKDSIPAWINYSQLQPDIEPYNGIHYPPEEERYIFQHPRPKPKPSLRI
 G: KDSIPAWI: : S: Q P: EIPYNGI. YDPPEEE: Y: F: HP: PK: P: S: RI
 GIKDSIPAWIKFSVQAPGEIPYNGIYYDPPEEEKYVFKHPOPKRPOSIRI
 ^290 ^300 ^310 ^320 ^330
 f350 f360 f370 f380 f390
 YESHIGMSSPEPKINSYVNFRDEVLPRIKKLGYNALQIMAIQEHSYYASF
 YESHIGMSSPEPKIN: Y. NFRD: VLPRIKKLGYN: QIMAIQEHSYYASF
 YESHIGMSSPEPKINTYANFRDDVLPRIKKLGYNQAVQIMAIQEHSYYASF
 ^340 ^350 ^360 ^370 ^380
 f400 f410 f420 f430 f440
 GHVHTNFFAPSSRFGTLPDDLKSLIDKAHELGI VVLMIDVHSHASNNTLDG
 GHVHTNFFAPSSRFGT: DLKSLID: AHELG: : VLMDIVHSH: SNNTLDG
 GHVHTNFFAPSSRFGTLPEDLKSLIDRAHELGLLVLMIDVHSHSSNNTLDG
 ^390 ^400 ^410 ^420 ^430

Fig. 7 SHEET 1

SUBSTITUTE SHEET (RULE 26)

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↗450 ↘460 ↗470 ↗480 ↗490
 LNMF DCTDSCYFHSGARGYHWMWDSRLFNYGNWEVLRYLLSNARWWLD.
 LNMF TD: YFH: G: RGYHWMWDSRLFNYG: WEVLRYLLSNARWWLD.
 LNMF DGTDGHYFHPGSRGYHWMWDSRLFNYGSWEVLRYLLSNARWWLDEY
 ↗440 ↗450 ↗460 ↗470 ↗480
 ↗500 ↗510 ↗520 ↗530 ↗540
 KFDGFRFDGVTSMMYIHGLSVGFTGNYEYFGLATDVDAVVYLMVNDL
 KFDGFRFDGVTSMMY. HHGL V: FTGNY. EYFGLATDV: AVVY: MLVNDL
 KFDGFRFDGVTSMMYTHHGLQVSFTGNYSEYFGLATDVEAVVYMLVNDL
 ↗490 ↗500 ↗510 ↗520 ↗530
 ↗550 ↗560 ↗570 ↗580 ↗590
 IHGLFPDAITIGEDVSGMPTFCIPVQEggVGFDYRLHMAIADKRIELLKK
 IHGLFP: A: : IGEDVSGMPTFC: P. Q: GG: GF: YRLHMA: ADK: IELLKK
 IHGLFPEAVSIGEDVSGMPTFCCLPTQDGIGFNYRLHMAVADKWIELLKK
 ↗540 ↗550 ↗560 ↗570 ↗580
 ↗600 ↗610 ↗620 ↗630 ↗640
 RDEDWRVGDIVHTLTNRRWSEKCVSYAESHDOALVGDKTIAFWLMDKDMY
 : DEDWR: GDIVHTLTNRRW EKCV YAESHDOALVGDKT: AFWLMDKDMY
 QDEDWRMGDIVHTLTNRRWLEKCVVYAESHDOALVGDKTLAFWLMDKDMY
 ↗590 ↗600 ↗610 ↗620 ↗630
 ↗650 ↗660 ↗670 ↗680 ↗690
 DFMALDRPSTSLIDRGIALHKMIRLVTMGLGEGYLNFMGNEFGHPEWID
 DFMALDRPST: LIDRGIALHKMIRL: TMGLGEGYLNFMGNEFGHPEWID
 DFMALDRPSTPLIDRGIALHKMIRLITMGLGEGYLNFMGNEFGHPEWID
 ↗640 ↗650 ↗660 ↗670 ↗680
 ↗700 ↗710 ↗720 ↗730 ↗740
 FPRAEQHLSDGSVIPGNQFSYDKCRRRFDLGDAEYLRYRGLQEFDRPMQY
 FPR: EQHL: : G. : : PGN: SYDKCRRRFDLGDA: YLRY: G: QEFDR: MQ.
 FPRGEQHLPNGKIVPGNNNSYDKCRRRFDLGDADYLRYHGMQEFDRAMQH
 ↗690 ↗700 ↗710 ↗720 ↗730
 ↗750 ↗760 ↗770 ↗780 ↗790
 LEDKYEFMTSEHQFISRKDEGDRMIYFEKGNLVFVFNFWTKSYSODYRIA
 LE: . Y. FMTSEHQ: ISRK: EGDR: I: FE: : NLVVFNFHWT: SYSDY: :
 LEETYGFMTSEHQYISRKNEGDRVIIFERDNLVFVFNFWTNYSYSDYKVG
 ↗740 ↗750 ↗760 ↗770 ↗780
 ↗800 ↗810 ↗820 ↗830 ↗840
 CLKPGKYKVALDSDPLFGGFGRIDHNAEYFTFEGWYDDRPRSIMVYAPC
 CLKPGKYK: . LDSDD. LFGGF. R: : H. AEYFT EGWYDDRPRS: : VYAP.
 CLKPGKYKIVLDSDDTLFGGFNRLNHTAEYFTSEGWYDDRPRSFLVYAPS
 ↗790 ↗800 ↗810 ↗820 ↗830
 ↗850 ↗860 ↗870
 KTAVVYALVDKEEEEEEEEEEVA
 : TAVVYAL. D E. E E . . V. :
 RTAVVYALADGVESEPIELSDGVES
 ↗840 ↗850 ↗860

Fig. 7 SHEET 2

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1	-----TTG-	AT
1	-----TTGA-	
1	-----GA-	
45	AAAAACCTCCTCCACTCAGTCTGGGATCTCTCTCTCT	
72	TTTCTTTAATTCCAACCAGCGAATGAATAAAAGGAT-A	
73	TTTCTTTAATTCCAACCAAGG-AATGAATAAAAGGAT-A	
71	TTTCTTTAATTCCAACCAAGG-AATGAATAAAAAGAT-A	
165	TTTCTTTAATTCCAACCAAGG-AATGAAT[AAAAGAT]A	
191	TGTACAAATCTAATGGATTCAAGCAGTAATGGTGATCGGAG	
191	TGTACAAATCTAATGGATTCAAGCAGTAATGGTGATCGGAG	
189	TGTACAAATCTAATGGATTCAAGCAGTAATGGTGATCGGAG	
274	TGTACAAATCTAATGGATTCAAGCAGTAATGGTGATCGGAG.	
311	AATTCCGACCTTCTACAGTTGCAGCATCGGGAAAGTCCT	
311	AATTCCGACCTTCTACAGTTGCAGCATCGGGAAAGTCCT	
309	AATCCGACCTTCTACAATTGCAGCATCGGGAAAGTCCT	
394	AATCCGACCTTCTACAGTTGCAGCATCGGGAAAGTCCT	
431	CAGCATCAACTGATGTAGATAGTTCAACAATGGAACACGC	
431	CAGCATCAACTGATGTAGATAGTTCAACAATGGAACACGC	
429	CAGCATCAACTGATGTAGATAGTTCAACAATGGAACACGC	
514	CAGCATCAACTGATGT[GATAGTTCAACAATGGAACACGC	
551	CATCACTACAACATACAAGAACGGTGGTAAACTGGAGGAGTC	
551	CATCACTACAACATACAAGAACGGTGGTAAACTGGAGGAGTC	
549	CATCACTACAACATACAAGAACGGTGGTAAACTGGAGGAGTC	
634	CATCACTACAACATACAAGAACGGTGGTAAACTGGAGGAGTC	
671	TTGGTCAGAAGATTATGAAATAGACCCCTTTGACAAA	
671	TTGGTCAGAAGATTATGAAATAGACCCCTTTGACAAA	
669	TTGGTCAGAAGATTATGAAATAGACCCCTTTGACAAA	
754	TTGGTCAGAAGATTATGAAATAGACCCCTTTGACAAA	
791	AAGC-TTTCTCGTGGTTATGAAAAAATGGGTTCACTCG	
791	AAGCCTTTCTCGTGGTTATGAAAAAATGGGTTCACTCG	
789	AAGCTTTCTCGTGGTTATGAAA[GAAATGGGTTCACTCG	
874	AAGCTTTCTCGTGGTTATGAAAAAATGGGTTCACTCG	

Fig.8
Sheet 2

Fig.8 SHEET 1

SUBSTITUTE SHEET (RULE 26)

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-----[REDACTED]-----GGGCCTTGAACTCAGCAATTGACACTCAGTTAGTTAC
 -----TGGGGCCTTGAACTCAGCAATTGACACTCAGTTAGTTAC
 -----TGGGGCCTTGAACTCAGCAATTGACACTCAGTTAGTTAC
[REDACTED]TACCGCTTCTCT[REDACTED]TGGGGCCTTGAACTCAGCAATTGACACTCAGTTAGTTAC

 GATTGTAAAAACCTAACGGAGAGAAGAAGAAAGATGGGTATA[REDACTED]ACTCTCT
 GATTGTAAAAACCTAACGGAGAGAAGAAGAAAGATGGGTATAACACTCTCT
 GATTGTAAAAACCTAACGGAGAGAAGAAGAAAGATGGGTATAACACTCTCT
 GATTG[REDACTED]AAGGAGAGAAGAAGAAAGATGGGTATAACACTCTCT

 GAATGCTAATGTTCTGTATTCTTGAAAAAGCACTCTCTTACCGGAAGATC
 GAATGCTAATGTTCTGTATTCTTGAAAAAGCACTCTCTTACCGGAAGATC
 GAATGCTAAT[REDACTED]CTGTATTCTTGAAAAAGCACTCTCTTACCGGAAGATC
 GAATGCTAATGTTCTGTATTCTTGAAAAAGCACTCTCTTACCGGAAGATC

 TGTGCCTGGAACCCAGAGTGTAGCTCCTCATCCTAACAGACCAATTGAG
 TGTGCCTGGAACCCAGAGTGTAGCTCCTCATCCTAACAGACCAATTGAG
 TGTGCCTGGAATCCAGAGTGTAGCTCCTCATCCTAACAGA[REDACTED]CAATTGAG
TGTACCTGGAATCCAGAGTGTAGCTCCTCATCCTAACAGACCAATTGAG

 TAGCCAGATTAAAAGTGAGAACGATGACGTTGAGCCGTCAGTGTATCTTACA
 TAGCCAGATTAAAAGTGAGAACGATGACGTTGAGCCGTCAGTGTATCTTACA
 TAGCCAGATTAAAAGTGAGAACGATGACGTTGAGCCGTCAGTGTATCTTACA
 TAGCCAGATTAAAAGTGAGAACGATGACGTTGAGCCGTCAGTGTATCTTACA

 TAAAACATTAATACCTCTGAAGAGACAATTATTGATGAATCTGATAGGATC
 TAAAACATTAATACCTCTGAAGAGACAATTATTGATGAATCTGATAGGATC
 TAAAACATTAATACCTCTGAAGAGACAATTATTGATGAATCTGATAGGATC
 TAAAACATTAATACCTCTGAAGAGACAATTATTGATGAATCTGATAGGATC

 CTATCGTCAACACCTTGATTACAGGTATTACAGTACAAGAAACTGAGGGAG
 CTATCGTCAACACCTTGATTACAGGTATTACAGTACAAGAAACTGAGGGAG
 CTATCGTCAACACCTTGATTACAGGTATTACAGTACAAGAAACTGAGGGAG
 CTATCGTCAACACCTTGATTACAGGTATTACAGTACAAGAAATGAGGGAG

 TAGTGCTACAGGTATCACTTACCGTGAGTGGGCTCTGGTGCCAGTCAGCT
 TAGTGCTACAGGTATCACTTACCGTGAGTGGGCT[REDACTED]GGTGCCAGTCAGCT
 TAGTGCTACAGGTATCACTTACCGTGAGTGGGCTCTGGTGCCAGTCAGCT
 TAGTGCTACAGGTATCACTTACCGTGAGTGGGCTCTGGTGCCAGTCAGCT

Fig. 8
Sheet
3

Fig. 8 SHEET 2

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ACTCCTATCACTTATCAGATCTCTATT 11con.seq
ACTCCTATCACTTATCAGATCTCTATT 19con.seq
ACT~~CC~~CATCACTTATCAGATCTCTATT 10con.seq
ACTCCTATCACT~~C~~AGATCTCTATT psbe2con.seq

GGAGTTCGTTTCCTACTGTTCCATCAG 11con.seq
GGAGTTCGTTTCCTACTGTTCCATCAG 19con.seq
GGAGTTCGTTTCCTACTGTTCCATCAG 10con.seq
GGAGTTCGTTTCCTACTGTTCCATCAG psbe2con.seq

TTGGCTGAAAAGTCTTCTTACAATTCCG 11con.seq
TTGGCTGAAAAGTCTTCTTACAATTCCG 19con.seq
TTGGCTGAAAAGTCTTCTTACAATTCCG 10con.seq
TTGGCTGAAAAGTCTTCTTAC~~C~~ATTCCG psbe2con.seq

TTCACTGAGACATCTCCAGAAAATTCCC 11con.seq
TTCACTGAGACATCTCCAGAAAATTCCC 19con.seq
TTC~~C~~CTGAGACATCTCCAGAAAATTCCC 10con.seq
TTCACTGAGACAGCTCCAGAAAATTCCC psbe2con.seq

GGAAGTGTGAAGAGCTGGATTTGCTT 11con.seq
GGAAGTGTGAAGAGCTGGATTTGCTT 19con.seq
GGAAGTGTGAAGAGCTGGATTTGCTT 10con.seq
GGAAGTGTGAAGAG~~T~~TGGATTTGCTT psbe2con.seq

AGAGAGAGGGGCATCCCTCACCTGGAC 11con.seq
AGAGAGAGGGGCATCCCTCACCTGGAC 19con.seq
AGAGAGAGGGGCATCCCTCACCTGGAC 10con.seq
AGAGAGAGGGGCATCCCTCACCTGGAC psbe2con.seq

GCAATTGACAAGTATGAGGGTGGTTGG 11con.seq
GCAATTGACAAGTATGAGGGTGGTTGG 19con.seq
GCAATTGACAAGTATGAGGGTGGTTGG 10con.seq
GCAATTGACAAGTATGAGGGTGGTTGG psbe2con.seq

GCCCTCATTGGAGATTCAACAATTGGG 11con.seq
GCCCTCATTGGAGATTCAACAATTGGG 19con.seq
GCCCTCATTGG~~C~~GATTCAACAATTGGG 10con.seq
GCTCTCATTGGAGATTCAACAATTGGG psbe2con.seq

Fig. 8

SHEET 3

910 ACGCAAATGCTGACATTATGACTCGGAATGAATTGGTGT
 911 ACGCAAATGCTGACATTATGACTCGGAATGAATTGGTGT
 909 ACGCAAATGCTGACATTATGACTCGGAATGAATTGGTGT
 994 ACGCAAATGCTGACATTATGACTCGGAATGAATTGGTGT

 1030 CTCCATCAGGTGTTAAGGATTCCATTCTGCTTGGATCAAC
 1031 CTCCATCAGGTGTTAAGGATTCCATTCTGCTTGGATCAAC
 1029 CTCCATCAGGTGTTAAGGATTCCATTCTGCTTGGATCAAC
 1114 CT~~T~~CATCAGGTGTTAAGGATTCCATTCTGCTTGGATCAAC

 1150 AACACCCACGCCAAAGAAACCAAGTCGCTGAGAATATAT
 1151 AACACCCACGCCAAAGAAACCAAGTCGCTGAGAATATAT
 1149 AACACCCACGCCAAAGAAACCAAGTC~~G~~CTGAGAATATAT
 1234 AACACCCACGCCAAAGAAACCAAGTCGCTGAGAATATAT

 1270 TAAAAAA-GCTTGGGTACAATGC~~G~~CTG~~C~~AAATTATGGCTAT
 1271 TAAAAAA-GCTTGGGTACAATGC~~G~~CTGCAAATTATGGCTAT
 1269 TAAAAAAAGCTTGGGTACAATGC~~G~~GTGCAAATTATGGCTAT
 1354 TAAAAAAAC-CTTGGGTACAATGC~~G~~GTGCAAATTATGGCTAT

 1389 GACGACCTTAAGTCTT~~G~~GATTGATAAAGCTCATGAGCTAGG
 1390 GACGACCTTAAGTCTT~~G~~GATTGATAAAGCTCATGAGCTAGG
 1389 GACGACCTTAAGTCTT~~G~~GATTGATAAAGCTCATGAGCTAGG
 1473 GACGACCTTAAGTCTT~~G~~GATTGATAAAGCTCATGAGCTAGG

 1509 GATAGTTGTTACTTTCACTCTGGAGCTCGTGGTTATCATTG
 1510 GATAGTTGTTACTTTCACTCTGGAGCTCGTGGTTATCATTG
 1509 GATAGTTGTTACTTTCACTCTGGAGCTCGTGGTTATCATTG
 1593 GATAGTTGTTACTTTCACTCTGGAGCTCGTGGTTATCATTG

 1628 GATGAGTTCAAATTGATGGATTAGATT~~G~~GATGGTGTGAC
 1630 GAT~~G~~GTTCAAATTGATGGATTAGATTGATGGTGTGAC
 1629 GATGAGTTCAAATTGATGGATTAGATTGATGGTGTGAC
 1713 GATGAGT~~G~~CAAATTG~~G~~TGGATTAGATTGATGGTGTGAC

 1748 GTGGATGCTGTTGTATCTGATGCTGGTCAACGATCTTAT
 1750 GTGGATGCTGTTGTATCTGATGCTGGTCAACGATCTTAT
 1749 GTGGATGCTGTTGTATCTGATGCTGGTCAACGATCTTAT
 1833 GTRGATGCTG~~G~~GTATCTGATGCTGG~~G~~CAACGATCTTAT

Fig. 8
Sheet 5Fig. 8
SHEET 4

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TGGGAGATTTCTGCCAAATAATGTGGATGGTCTCCTGCAATT
 TGGGAGATTTCTGCCAAATAATGTGGATGGTCTCCTGCAATT
 TCAAGAGATTTCTGCCAAATAATGTGGATGGTCTCCTGCAATT
 TGGGAGATTTCTGCCAAATAATGTGGATGGTCTCCTGCAATT
 TACTCTTACAGCTCCTGATGAAATTCCATATAATGGAATATATT
 TACTCTTACAGCTCCTGATGAAATTCCATATAATGGAATACATT
 TACTCTTACAGCTCCTGATGAAATTCCATATAATGGAATATATT
 TACTCTTACAGCTCCTGATGAAATTCCATATAATGGAATATATT
 GAATCTCATATTGAATGAGTAGTCCGGAGCCTAAAATTAACCAT
 GAATCTCATATTGAATGAGTAGTCCGGAGCCTAAAATTAACCAT
 GAATCTCATATTGAATGAGTAGTCCGGAGCCTAAAATTAACCAT
 GAATCTCATATTGAATGAGTAGTCCGGAGCCTAAAATTAACCAT
 TCAAGAGCATTCTTATTATGCTAGTTTGGTTATCATGTACAAAT
 TCAAGAGCATTCTTATTACGCTAGTTTGGTTATCATGTACAAAT
 TCAAGAGCATTCTTATTATGCTAGTTTGGTTATCATGTACAAAT
 TCAAGAGCATTCTTATTATGCTAGTTTGGTTATCATGTACAAAT
 AATTGTTGTTCTCATGGACATCGTTACAGCCATGCATCAAATAAT
 AATTGTTGTTCTCATGGACATTGTTACAGCCATGCATCAAATAAT
 AATTGTTGTTCTCATGGACATTGTTACAGCCATGCATCAAATAAT
 AATTGTTGTTCTCATGGACATTGTTACAGCCATGCATCAAATAAT
 GATGTGGGATTCCGCCTCTTAACTATGGAAACTGGGAGGTACTT
 GATGTGGGATTCCGCCTCTTAACTATGGAAACTGGGAGGTACTT
 GATGTGGGATTCCGCCTCTTAACTATGGAAACTGGGAGGTACTT
 GATGTGGGATTCCGCCTCTTAACTATGGAAACTGGGAGGTACTT
 ATCAATGATGTACTCACCAACGGATTATCGGTGGGATTCACTGGG
 ATCAATGATGTATACTCACCAACGGATTATCGGTGGGATTCACTGGG
 ATCAATGATGTACTCACCAACGGATTATCGGTGGGATTCACTGGG
 ATCAATGATGTACTCACCAACGGATTATCGGTGGGATTCACTGGG
 TCATAGGCTTTCCCAGATGCAATTACCATGGTGAAGATGTTAGC
 TCATGGGCTTTCCCAGATGCAATTACCATGGTGAAGATGTTAGC
 TCATGGGCTTTCCCAGATGCAATTACCATGGTGAAGATGTTAGC
 TCATGGGCTTTCCCAGATGCAATTACCATGGTGAAGATGTTAGC

Fig. 8
Sheet 6Fig. 8
SHEET 5

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CTCATGGTCCAGAGTGAAGATACTATGGACA 11con.seq
 CTCATGGTCCAGAGTGAAGATACTATGGACA 19con.seq
 CTCATGGTCCAGAGTGAAGATACTATGGACA 10con.seq
 CTCATGGTCCAGAGTGAAGATACTATGGACA psbe2con.seq

ATGATCCACCGAAGAGGGAGAGGTATATCTTCC 11con.seq
 ATGATCCACCGAAGAGGGAGAGGTATATCTTCC 19con.seq
 ATGATCCACCGAAGAGGGAGAGGTATATCTTCC 10con.seq
 ATGATCCACCGAAGAGGGAGAGGTATATCTTCC psbe2con.seq

ACGTGAATTTAGAGATGAAGTTCTCCTCGCA 11con.seq
 ACGTGAATTTAGAGATGAAGTTCTCCTCGCA 19con.seq
 ACGTGAATTTAGAGATGAAGTTCTCCTCGCA 10con.seq
 ACGTGAATTTAGAGATGAAGTTCTCCTCGCA psbe2con.seq

TTTTTGACCAAGCAGCCGTTTGGAACGCC 11con.seq
 TTTTTGACCAAGCAGCCGTTTGGAACGCC 19con.seq
 TTTTTGACCAAGCAGCCGTTTGGAACGCC 10con.seq
 TTTTTGACCAAGCAGCCGTTTGGAACGCC psbe2con.seq

ACTTAGATGGACTAACATGTTGACGGCACC 11con.seq
 ACTTAGATGGACTAACATGTTGACGGCACC 19con.seq
 ACTTAGATGGACTAACATGTTGACGGCACC 10con.seq
 ACTTAGATGGACTAACATGTTGACGGCACC psbe2con.seq

AGGTATCTCTCTCAAATGCGAGATGGTGGTTG 11con.seq
 AGGTATCTCTCTCAAATGCGAGATGGTGGTTG 19con.seq
 AGGTATCTCTCTCAAATGCGAGATGGTGGTTG 10con.seq
 AGGTATCTCTCTCAAATGCGAGATGGTGGTTG psbe2con.seq

AACTACGAGGAATACTTGGACTCGCAACTGAT 11con.seq
 AACTACGAGGAATACTTGGACTCGCAACTGAT 19con.seq
 AACTACGAGGAATACTTGGACTCGCAACTGAT 10con.seq
 AACTACGAGGAATACTTGGACTCGCAACTGAT psbe2con.seq

GGAATGCCGACATTTGTATTCCGTTCAAGAT 11con.seq
 GGAATGCCGACATTTGTATTCCGTCAGAG 19con.seq
 GGAATGCCGACATTTGTATTCCGTTCAAGAT 10con.seq
 GGAATGCCGACATTTGTATTCCGTTCAAGAT psbe2con.seq

Fig. 8
SHEET 6

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Fig.8
Sheet 8

Fig. 8
SHEET 7

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TGATAAAATGGATTGAGTTGCTCAAGAACGGATGAGGATTGGAGA
 TGATAAACGGATTGAGTTGCTCAAGAACGGATGAGGATTGGAGA
 TGATAAAATGGATTGAGTTGCTCAAGAACGGATGAGGATTGGAGA
 TGATAAAATGGATTGAGTTGCTCAAGAACGGATGAGGATTGGAGA

 TCAAGCTCTAGTCGGTGATAAAACTATAGCATTCTGGCTGATGGAC
 TCAAGCTCTAGTCGGTGATAAAACTATAGCATTCTGGCTGATGGAC
 TCAAGCTCTAGTCGGTGATAAAACTATAGCATTCTGGCTGATGGAC
 TCAAGCTCTAGTCGGTGATAAAACTATAGCATTCTGGCTGATGGAC

 GATGATTAGGCTTGTAACTATGGGATTAGGAGGAAGGGTACCTA
 GATGATTAGGCTTGTAACTATGGGATTAGGAGGAAGGGTACCTA
 GATGATTAGGCTTGTAACTATGGGATTAGGAGGAAGGGTACCTA
 GATGATTAGGCTTGTAACTATGGGATTAGGAGGAAGGGTACCTA

 CTCAGTAATTCCCGAAACCAATTCAGTTTATGATAAATGCAGACGG
 CTCAGTAATCCCCGAAACCAATTCAGTTTATGATAAATGCAGACGG
 CTCAGTAATTCCGAAACCAATTCAGTTTATGATAAATGCAGACGG
 CTCAGTAATTCCCGAAACCAATTCAGTTTATGATAAATGCAGACGG

 TGAAGATAAAATATGAGTTATGACTTCAGAACACCAGTTCATATCA
 TGAAGATAAAATATGAGTTATGACTTCAGAACACCAGTTCATATCA
 TGAAGATAAAATATGAGTTATGACTTCAGAACACCAGTTCATATCA
 TGAAGATAAAATATGAGTTATGACTTCAGAACACCAGTTCATATCA

 AAATAGCTATTCAGACTATCGCATAGGCTGCCTGAAGCCTGGAAAA
 AAAAGCTATTCAGACTATCGCATAGCTGCCTGAAGCCTGGAAAA
 AAAGCTATTCAGACTATCGCATAGGCTGCCTGAAGCCTGGAAAA
 AAAAGCTATTCAGACTATCGCATAGGCTGCTGAAGCCTGGAAAA

 CACCTGTGAAGGATCGTATGATGATCGTCTGTCAATTATGGTG
 CACCTTGAGGATGGTATGATGATCGTCTCGTTCAATTATGGTG
 CACCTTGAGGATGGTATGATGATCGTCTCGTTCAATTATGGTG
 CACCTTGAGGATGGTATGATGATCGTCTCGTTCAATTATGGTG

 -----TAGCAGTAGTAGAAGAACCCATG-----AAGAACG
AGAAATGCACGTAGTAGAAGTAGTAGTAGAAGAACG
-----TAGCAGTAGTAGAAGAAGTAGTAGTAGAAGAACG
-----TAGCAGTAGTAGAAGAAGTAGTAGTAGAAGAACG

Fig. 8
Sheet 9Fig. 8
SHEET 8

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GTGGGTGATATTGTTCATACACTGACAAATAGA 11con.seq
 GTGGGTGATATTGTTCATACACTGACAAATAGA 19con.seq
 GTGGGTGATATTGTTCATACACTGACAAATAGA 10con.seq
 GTGGGTGATATTGTTCATACACTGACAAATAGA psbe2con.seq

AAGGATATGTATGATTTATGGCTCTGGATAGA 11con.seq
 AAGGATATGTATGATTTATGGCTCTGGATAGA 19con.seq
 AAGGATATGTATGATTTATGGCTCTGGATAGA 10con.seq
 AAGGATATGTATGATTTATGGCTCTGGATAGA psbe2con.seq

AATTCATGGAAATGAATTGGCCACCCCTGAG 11con.seq
 AATTCATGGAAATGAATTGGCCACCCCTGAG 19con.seq
 AATTCATGGAAATGAATTGGCCACCCCTGAG 10con.seq
 AATTCATGGAAATGAATTGGCCACCCCTGAG psbe2con.seq

AGATTGACCTGGAGATGCAGAATATTAAGA 11con.seq
 AGATTGACCTGGAGATGCAGAATATTAAGA 19con.seq
 AGATTGACCTGGAGATGCAGAATATTAAGA 10con.seq
 AGATTGACCTGGAGATGCAGAATATTAAGA psbe2con.seq

CGAAAGGATGAAGGAGATAGGATGATTGTATTT 11con.seq
 CGAAAGGATGAAGGAGATAGGATGATTGTATTT 19con.seq
 CGAAAGGATGAAGGAGATAGGATGATTGTATTT 10con.seq
 CGAAAGGATGAAGGAGATAGGATGATTGTATTT psbe2con.seq

TACAAGGTTGCTTGGACTCAGATGATCCACTT 11con.seq
 TACAAGGTTGCCTTGGACTCAGATGATCCACTT 19con.seq
 TACAAGGTTGCCTTGGACTCAGATGATCCACTT 10con.seq
 TACAAGGTTGCCTTGGACTCAGATGATCCACTT psbe2con.seq

TATGCACCTAGTAGAACAGCAGTGGTCTATGCA 11con.seq
 TATGCACCTGTAACAGCAGTGGTCTATGCA 19con.seq
 TATGCACCTAGTAGAACAGCAGTGGTCTATGCA 10con.seq
 TATGCACCTAGTAGAACAGCAGTGGTCTATGCA psbe2con.seq

AACTTGTGATCGCGTTGAAAGATTGAACCTTA 11con.seq
 AACTTGTGATCGCGTTGAAAGATTGAACG--- 19con.seq
 AACTTGTGATCGCGTTGAAAGATTGAACG--- 10con.seq
 AACTTGTGATCGCGTTGAAAGATTGAACG--- psbe2con.seq

Fig. 8
SHEET 9

33/75

2795 CTTGGTCATCCACATAGAGCTTCTTGAC-----
2827 -----CTACATAGAGCTTCTTGACGTATCTGGCAATAT
2814 -----CCACATAGAGCTTCTTGACGTATCTGGCAATAT
2895 -----CTACATAGAGCTTCTTGACGTATCTGGCAATAT

2898 AGAGATGAAGTGCTGAACAAA--CATATGTAATCGATGAA
2937 AGAGATGAAGTGCTGAACAAA--CATATGTAATCGATGAA
2924 AGAGATGAAGTGCTGAACAAAACATATGTAATCGATGAA
3005 AGAGATGAAGTGCTGAACAAA--CATATGTAATCGATGAA

2975
3012
3003
3123 GCCCACTAGAAATCAATTATGTGAGACCTAAAAACAATAAC

{ Fig. 8
Sheet 11 }

Fig. 8 SHEET 10

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---ATCAGTCTTGGCGGAATTCCATGTGACAA-AAGGTTGCAATT
TGCATCAGTCTTGGCGGAATTTCATGTGACAC-AAGGTTGCAATT
TGCATTAAGTCTTGGCGGAATTTCATGTGACAA-CAGGTTGCAATT
TGCATCAGTCTTGGCGGAATTTCATGTGACAA-AAGGTTGCAATT

TTTATGTCGAATGCTGGACGATCGAATT CCTGCAGCC
TTTATGTCGAATGCTGGACGATCGAATT CCTGCAG
TTTATGTCGAATGCTGGACGATCGAATT CCTGCAGCC
TTTATGTCGAATGCTGGACGGGCTTCAGCAGGTTGCTTAGTGA

CATAAAATGGAAATAGTGCTGATCTAATGATGTTAACNNNNNA

Fig. 8
Sheet 12

Fig. 8 SHEET 11

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CTTTCCACTATTAGTAGT**CCACCGATATAACGC** 11con.seq
CTTTCCACTATTAGTAGTGCAACGATATAACGC 19con.seq
CTTTCCACTATTAGTAGTGCAACGATATAACGC 10con.seq
CTTTCCACTATTAGTAGTGCAACGATATAACGC psbe2con.seq

11con.seq
19con.seq
10con.seq
GTTCTGTAAATTGTCATCTCTTANATGTACA psbe2con.seq

11con.seq
19con.seq
10con.seq
AAAAAAAAAAAAAAACTCGAG psbe2con.seq

Fig. 8 SHEET 12

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GGATGCTAATGTTCTGTATTCTTAAAAAGCACTCTCTTCACGG
 CCTACGATTACAAAGACATAAGAACCTTTCTGTGAGAGAAAGTGCC
 A N V S V F L K K H S L S R

TTCTACAGTTGCAGCATCGGGAAAGTCCTTGTGCCCTGGAAYCCAG
 AAGATGTCAACGTCGTAGCCCCCTTCAGGAACACGGACCTTRGGTC
 S T V A A S G K V L V P G ? O

GACATCTCCAGAAAATTCCCCAGCATCAACTGATGTAGATAGTTCA
 CTGTAGAGGTCTTTAAGGGTCGTAGTTGACTACATCTATCAAGT
 T S P E N S P A S T D V D S S

TGAGCCGTCAAGTGATCTTACAGGAAGTGTGAAGAGAGCTGGATTT
 ACTCGGCAGTTCACTAGAATGTCCTTCACAACCTCTGACCTAAAA
 E P S S D L T G S V E E L D F

TAAAACATTAATACTTCTGAAGAGACAATTATTGATGAATCTGAT
 ATTTGTAATTTATGAAGACTTCTCTGTTAATAACTACTTAGACTA
 K T L N T S E E T I I D E S D

Hinc II

GATTATGAAATAGACCCCCCTTTGACAAACTATCGTCAACACCTT
 CTAATACTTTATCTGGGGAAAACGTGTTGATAGCAGTTGTGGAA
 I Y E I D P L L T N Y R Q H L

Fig.9
Sheet
2

Fig. 9 SHEET 1

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Bgl II

AAGATCTTGGCTGAAAAGTCTTCTTACAATTCCGAATCCGACC 90
 TTCTAGAACCGACTTTCAAGAAGAATGTTAAGGCTTAGGGCTGG
 K I L A E K S S Y N S E S R P

 AGTGATAGCTCCTCATCCTCAACAGACCAATTGAGTTCACTGA 180
 TCACTATCGAGGAGTAGGGAGTTGTCTGGTTAACTCAAGTGACT
 S D S S S S S T D Q F E F T E

 ACAATGGAACACGCTAGCCAGATTAAACTGAGAACGATGACGT 270
 TGTTACCTTGTGCGATCGGTCTAATTTGACTCTTGCTACTGCA
 T M E H A S Q I K T E N D D V

 GCTTCATCACTACAACATACAAGAAGGTGGTAAACTGGAGGAGTC 360
 CGAAGTAGTGTGATGTTGATGTTCTTCCACCATTTGACCTCCTCAG
 A S S L Q L Q E G G K L E E S

 AGGATCAGAGAGAGGGGCATCCCTCCACCTGGACTTGGTCAGAA 450
 TCCTAGTCTCTCCCCGTAGGGAGGTGGACCTGAACCAGTCTT
 R I R E R G I P P P G L G Q K

 GATTACAGGTATTCACAGTACAAGAAACTGAGGGAGGCAATTGA 540
 CTAATGTCCATAAGTGTCACTGTTCTTGACTCCCTCCGTTAACT
 D Y R Y S Q Y K K L R E A I D

Fig. 9 SHEET 2

38/75

HinD III

CAAGTATGAGGGTGGTTGGAAAGCTTTCTCGTGGTTATGAAAAA
 GTTCATACTCCCACCAAACCTCGAAAAAGAGCACCAATACTTTT
 K Y E G G L E A F S R G Y E K

Pvu II

GGCTCCTGGTGCCAGTCAGCTGCCCTCATGGAGATTCAACAAT
 CCGAGGACCACGGGTCAAGTCGACGGGAGTAACCTCTAAAGTTGTTA
 A P G A Q S A A L I G D F N N

CTGGGAGATTTCTGCCAAATAATGTGGATGGTCTCCTGCAATT
 GACCCCTCTAAAAAGACGGTTATTACACCTACCAAGAGGACGTTAA
 W E I F L P N N V D G S P A I

TGTTAAGGATTCCATTCTGCTTGGATCAACTACTCTTACAGCTT
 ACAATTCTAAGGTAAAGGACGAACCTAGTTGATGAGAAATGTCGAA
 V K D S I P A W I N Y S L Q L

AGAGGAGAGGTATRTCTTCCAACACCCACGGCCAAAGAAACCAAAG
 TCTCCTCTCCATAYAGAAGGTTGTGGGTGCCGGTTCTTGTTTC
 E E R Y ? F Q H P R P K K P K

Fig.9
Sheet
4

Fig.9 SHEET 3

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ATGGGTTTCACTCGTAGTGCTACAGGTATCACTTACCGTGAGTG
630
TACCCAAAGTGAGCATCACGATGTCCATAGTGAATGGCACTCAC
M G F T R S A T G I T Y R E W

TGGGACGCAAATGCTGACATTATGACTCGGAATGAATTGGTGT
720
ACCCCTGCCTTACGACTGTAATACTGAGCCTTACTTAAACCACA
W D A N A D I M T R N E F G V

CCTCATGGGTCCAGAGTGAAGATACTGYATGGACACTCCATCAGG
810
GGAGTACCCAGGTCTCACTTCTATGCRTACCTGTGAGGTAGTCC
P H G S R V K I R M D T P S G

CCTGATGAAATTCCATATAATGGAATATATTATGATCCACCGA
900
GGACTACTTAAGGTATATTACCTTATATAATAACTAGGTGGGCT
P D E I P Y N G I Y Y D P P E

TCGCTGAGAATATATGAATCTCATATTGGAATGAGTAGTCCGGA
990
AGCGACTCTTATATACTTAGAGTATAACCTTACTCATCAGGCCT
S L R I Y E S H I G M S S P E

Fig. 9 SHEET 4

40/75

Xmn I

GCCTAAAATTAAC	TCACTCATACGTGA	ATTTAGAGATGAAG	TTCTTCCT
P K I N S Y V	N F R D E V L P		
TCAAGAGCATTCTT	ATTATGCTAGTTGGTT	ATCATGTCACA	AAT
AGTTCTCGTAAGA	ATAATACGATCAAACCA	ATAGTACAGTGT	TTA
Q E H S Y Y A S	F G Y H V T N		
GTCTTGATTGATAAA	AGCTCATGAGCTAGGA	ATTGTTGTTCTCATG	
CAGAAACTAACTATTC	GAGTACTCGATCCTAAC	ACAACAAGAGTAC	
S L I D K A H E	L G I V V L M		
GAACATGTTGACGG	CACAGATAAGTTGTT	ACTCTGGAGCT	
CTTGTACAAACTGCC	GTGTCTATCAACAA	ATGAAAGTGAGAC	CTCGA
N M F D G T D S	C Y F H S G A		
AAACTGGGAGGTACTT	AGGTATCTTCTCTCA	AAATGCGAGATGGT	GG
TTTGACCCTCCATGA	ATCCATAGAAGAGAG	TTACGCTCTACCACC	
N W E V L R Y L	L S N A R W W		
ATCAATGATGTATA	ACTCACCA	CACGGATTATCGGT	GGGATTCACTGGG
TAGTTACTACATATG	AGTGCGCTAATAGCC	ACCCCTAAGTGACCC	
S M M Y T H H G	L S V G F T G		

Fig.9
Sheet
6

Fig.9 SHEET 5

SUBSTITUTE SHEET (RULE 26)

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CGCATAAAAAASCTTGGGTACAATGCGGTGCAAATTATGGCTAT
 1080
 GCGTATTTTSGAACCCATGTTACGCCACGTTAATACCGATA
 R I K ? L G Y N A V Q I M A I

 TTTTTGCACCAAGCAGCCGTTTGGAACGCCGACGACCTAA
 1170
 AAAAACGTGGTTCGTCGGCAAAACCTTGCAGGGCTGCTGGAATT
 F F A P S S R F G T P D D L K

 GACATTGTTCACAGCCATGCATCAAATAACTTTAGATGGACT
 1260
 CTGTAACAAGTGTGGTACGTAGTTATTATGAAATCTACCTGA
 D I V H S H A S N N T L D G L

Sac I

CGTGGTTATCATTGGATGTGGATTCCCGCCTCTTAACATGG
 1350
 GCACCAATAGTAACCTACACCCCTAACGGCGGAGAAATTGATACC
 R G Y H W M W D S R L F N Y G

 TTGGATGAGTTCAAATTGATGGATTAGATTGATGGTGTGAC
 1440
 AACCTACTCAAGTTAAACTACCTAAATCTAAACTACCCACACTG
 L D E F K F D G F R F D G V T

 AACTACGAGGAATACTTGGACTCGCAACTGATGTGGATGCTGT
 1530
 TTGATGCTCCTTATGAAACCTGAGCGTTGACTACACCTACGACA
 N Y E E Y F G L A T D V D A V

Fig. 9 SHEET 6

42/75

Hinc II

TGTGTATCTGATGCTGGTCAACGATCTTATTACGGGCTTTCCCA
 ACACATAGACTACGACCAGTTGCTAGAATAAGTGCCCCAAAAGGGT
 V Y L M L V N D L I H G L F P

TTGTATTCCC GTTCAAGATGGGGTGTGGCTTGACTATCGGCTG
 AACATAAGGGCAAGTTCTACCCCCACAACCGAAACTGATAGCCGAC
 C I P V Q D G G V G F D Y R L

GGATGAGGATTGGAGAGTGGGTGATATTGTTCATACACTGACAAAT
 CCTACTCCTAACCTCTCACCCACTATAACAAGTATGTGACTGTTA
 D E D W R V G D I V H T L T N

TCAAGCTCTAGTCGGT GATAAAACTATAGCATYCTGGCTGATGGAC
 AGTTCGAGATCAGCCACTATTTGATATCGTARGACCGACTACCTG
 Q A L V G D K T I A ? W L M D

ATTAATAGATCGTGGGATAGCATTGCACAAGATGATTAGGCTTGT A
 TAATTATCTAGCACCCCTATCGTAACGTGTTCTACTAATCCGAACAT
 L I D R G I A L H K M I R L V

Fig.9
Sheet
8

Fig. 9 SHEET 7

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GATGCAATTACCATGGTGAAGATGTTAGCGGAATGCCGACATT
 1620
 CTACGTTAACGGTAACCACTTCTACAATCGCCTTACGGCTGTAA
 D A I T I G E D V S G M P T F

Nde I

CATATGGCAATTGCTGATAAATGGATTGAGTTGCTCAAGAAACG
 1710
 GTATACCGTTAACGACTATTTACCTAACTCAACGAGTTCTTGC
 H M A I A D K W I E L L K K R

AGAAGATGGTCGGAAAAGTGTGTTCATMCGCTGAAAGTCATGA
 1800
 TCTTCTACCAGCCTTTCACACAAAGTAKGCGACTTCAGTACT
 R R W S E K C V S ? A E S H D

Hinc II

AAGGATATGTATGATTTATGGCTCTGGATAGACCGTCAACATC
 1890
 TTCCCTATACATACTAAAATACCGAGACCTATCTGGCAGTTGTAG
 K D M Y D F M A L D R P S T S

Asp 718
Kpn I

ACTATGGGATTAGGAGGAGAAGGGTACCTAAATTATGGAAA
 1980
 TGATACCCCTAACCTCCCTTCCATGGATTAAAGTACCCCTT
 T M G L G G E G Y L N F M G N

Fig. 9 SHEET 8

44/75

EcoR I

TGAATTGGCCACCCCTGAGTGGATTGATTCCTAGGGCTGARCAA
 ACTTAAGCCGGTGGGACTCACCTAACTAAAGGGATCCCAGCTYGT.
 E F G H P E W I D F P R A E Q.

Ssp I

TGATAAATGCAGACGGAGATTGACCTGGGAGATGCAGAATATTAA
 ACTATTTACGTCTGCCTCTAAACTGGACCCCTACGTCTTATAAAAT
 D K C R R R F D L G D A E Y L

 TGAAGATAAAATATGAGTTATGACTTCAGAACACCAGTTCATATCA
 ACTTCTATTTATACTCAAATACTGAAGTCTTGTGGTCAAGTATAAGT
 E D K Y E F M T S E H Q F I S

 CCTAGTTTTGTCTTAATTTCACTGGACAAATAGCTATTAGAC
 GGATCAAAAACAGAAATTAAAAGTGACCTGTTATCGATAAGTCTG
 L V F V F N F H W T N S Y S D

Fig 9
Sheet
10

GGACTCAGATGATCCACTTTGGTGGCTTCGGGAGAATTGATCAT
 CCTGAGTCTACTAGGTGAAAAACCACCGAAGCCCTCTTAACTAGTA
 D S D D P L F G G F G R I D H

YCGYYCAATTATGGTGTATGCACCTAGTAGAACAGCAGTGGTCTAT
 RGCCRRTTAATACACATACGTGGATCATCTTGTGTCGTACCCAGATA
 R ? I M V Y A P S R T A V V Y

NGAAGAATT
 NCTTCTTAAAAA
 E E F

2531

Fig 9 SHEET 9

45/75

CACCTCTCTGATGGCTCAGTAATTCCCGGAAACCAATTCAAGTTA
 GTGGAGAGACTACCGAGTCATTAAGGGCCTTGGTTAACGTCAAT
 H L S D G S V I P G N Q F S Y

Nco I

AGATACCATTGGTTGCAAGAATTGACCGGGCTATGCAGTATCT
 TCTATGGTACCCAACGTTCTTAAACTGGCCCATACTGTACAGTACAGA
 R Y H G L Q E F D R A M Q Y L

CGAAAGGATGAAGGAGATAGGATGATTGTATTTGAAARAGGAAA
 GCTTCCTACTTCCTCTATCCTACTAACATAAACTTTYTCCTTT
 R K D E G D R M I V F E ? G N

TATCGCATAGGCTGCCTGAAGCCTGGAAAATACAAGGTTGGCTT
 ATAGCGTATCCGACGGACTTCGGACCTTTATGTTCCAACCGAA
 Y R I G C L K P G K Y K V G L

Ssp I

AATGCCGAATATTCACCTCTGAAGGATCGTATGATGATCGYCC
 TTACGGCTTATAAAGTGGAGACTTCCTACTACACTAGCRGG
 N A E Y F T S E G S Y D D R P

GCACTAGTAGACAAANTAGAAGNAGAAGAAGAAGAAGAAGAANCCGN
 CGTGATCATCTGTTNATCTCNTCTTCTTCTTNGGCN
 A L V D K ? E ? E E E E E ? ?

Fig. 9 SHEET 10

SUBSTITUTE SHEET (RULE 26)

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	10	20	30
1	-GATGGGGCCTTGAAC	TCAAGCAATTGACACTCA	GTCAGT
1	TTGATGGG-CCTTGAAC	TCAAGCAATTGACACTCA	GTCAGT
1	TTGATGGGGCCTTGAAC	TCAAGCAATTGACACTCA	GTCAGT
1	T-----	-----	-----
1	-----	-----	-----
	80	90	100
69	TTTTCTCTTAATTCCAACC	AAGG-AATGAATAAAA	A
70	TTTTCTCTTAATTCCAACC	AGGG-AATGAATAAAAG	A
71	TTTTCTCTTAATTCCAACC	AAGG-AATGAATAAAAG	A
7	-----	-----	AAGAG
1	-----	-----	-----
	150	160	170
138	GAAAGATGGGTATA	ACTCTCTGGAGTTCGTTTCC	C
140	GAAAGATGGGTATA	TA	T
140	GAAAGATGGGTATA	ACTCTCTGGAGTTCGTTTCC	C
33	-----	-----	TCT
1	-----	-----	-----
	220	230	240
208	CAGCAGTAATGGTGATCGGAGGAATGCTAAT	ATTTCT	*
210	CAGCAGTAATGGTGATCGGAGGAATGCTAATGTTCT	-----	*
210	CAGCAGTAATGGTGATCGGAGGAATGCTAATGTTCT	-----	*
48	CA-----	-----	*
1	-----	GGATGCTAATGTTCT	*
	290	300	310
278	ATCTTGGCTGAAAAGTCTTCTTACAATTCCGAAT	CCC	*
280	ATCTTGGCTGAAAAGTCTTCTTACAATTCCGAATTCC	-----	*
280	ATCTTGGCTGAAAAGTCTTCTTACAATTCCGAATTCC	-----	*
57	ATCTTGGCTGAAAAGTCTTCTTACAATTCCGAATTCC	-----	*
50	ATCTTGGCTGAAAAGTCTTCTTACAATTCCGAAT	CCC	*

Fig.10
Sheet 2

Fig. 10 SHEET 1

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40	50	60	70	
TAGTTACACT	CCC ATCACTTATCAGATCTCTAT			10con. seq
TAGTTACACT	CCT ATCACTTATCAGATCTCTAT			11con. seq
TAGTTACACT	CCT ATCACTTATCAGATCTCTAT			19con. seq
	-CATTA-			86CON. SEQ
				pcrsbe2con. seq
110	120	130	140	
GATAGATTG	AAAAACCC	TAAGGAGAGAAGAA		10con. seq
GATAGATTG	AAAAACCC	TAAGGAGAGAAGAA		11con. seq
GATAGATTG	AAAAACCC	TAAGGAGAGAAGAA		19con. seq
GAGAAATT	-	AACTATGAGAGGA		86CON. SEQ
				pcrsbe2con. seq
180	190	200	210	
TACTGTTCC	CATCAGTGTACAAATCTAATGGATT			10con. seq
TACTGTTCC	CATCAGTGTACAAATCTAATGGATT			11con. seq
TACTGTTCC	CATCAGTGTACAAATCTAATGGATT			19con. seq
CACCAT	-CACCA-			86CON. SEQ
				pcrsbe2con. seq
250	260	270	280	
GTATTCTTG	AAAAAACACTCTCTTCACGGAAAG			10con. seq
GTATTCTTG	AAAAAGCACTCTCTTCACGGAAAG			11con. seq
GTATTCTTG	AAAAAGCACTCTCTTCACGGAAAG			19con. seq
	-CCATGG-	G		86CON. SEQ
GTATTCTTG	AAAAAGCACTCTCTTCACGGAAAG			pcrsbe2con. seq
320	330	340	350	
GACCTTCTACA	A TTGCAGCATCGGGGAAAGTCC			10con. seq
GACCTTCTACAG	TTGCAGCATCGGGGAAAGTCC			11con. seq
GACCTTCTACAG	TTGCAGCATCGGGGAAAGTCC			19con. seq
GACCTTCTACAG	TTGCAGCATCGGGGAAAGTCC			86CON. SEQ
GACCTTCTACAG	TTGCAGCATCGGGGAAAGTCC			pcrsbe2con. seq

Fig. 10 SHEET 2

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	360 *	370	380
348	TTGTGCCTGGAA <u>T</u> CCAGAGTGTAGCTCCTCATCCTC		
350	TTGTGCCTGGAA <u>ACCC</u> CAGAGTGTAGCTCCTCATCCTC		
350	TTGTGCCTGGAA <u>ACCC</u> CAGAGTGTAGCTCCTCATCCTC		
127	TTGTGCCTGGAA <u>ACCC</u> CAGAGTGTAGCTCCTCATCCTC		
120	TTGTGCCTGGAA <u>A</u> CCAGAGTGTAGCTCCTCATCCTC		
	430	440	450
418	AGAAAATTCCCCAGCATCAACTGATGTAGATAGTTCA		
420	AGAAAATTCCCCAGCATCAACTGATGTAGATAGTTCA		
420	AGAAAATTCCCCAGCATCAACTGATGTAGATAGTTCA		
197	AGAAAATTCCCCAGCATCAACTGATGTAGATAGTTCA		
190	AGAAAATTCCCCAGCATCAACTGATGTAGATAGTTCA		
	500	510	520
488	AACGATGACGTTGAGCCGTCAAGTGATCTTACAGGAA		
490	AACGATGACGTTGAGCCGTCAAGTGATCTTACAGGAA		
490	AACGATGACGTTGAGCCGTCAAGTGATCTTACAGGAA		
267	AACGATGACGTTGAGCCGTCAAGTGATCTTACAGGAA		
260	AACGATGACGTTGAGCCGTCAAGTGATCTTACAGGAA		
	570	580	590
558	AACTACAAGAAGGTGGTAAACTGGAGGGAGTCTAAAAC		
560	AACTACAAGAAGGTGGTAAACTGGAGGGAGTCTAAAAC		
560	AACTACAAGAAGGTGGTAAACTGGAGGGAGTCTAAAAC		
337	AACTACAAGAAGGTGGTAAACTGGAGGGAGTCTAAAAC		
330	AACTACAAGAAGGTGGTAAACTGGAGGGAGTCTAAAAC		
	640	650	660
628	ATCTGATAGGATCAGAGAGAGGGGCATCCCTCCACCT		
630	ATCTGATAGGATCAGAGAGAGGGGCATCCCTCCACCT		
630	ATCTGATAGGATCAGAGAGAGGGGCATCCCTCCACCT		
407	ATCTGATAGGATCAGAGAGAGGGGCATCCCTCCACCT		
400	ATCTGATAGGATCAGAGAGAGGGGCATCCCTCCACCT		

Fig. 10
Sheet 4

Fig. 10 SHEET 3

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390

400

410

420

AACAGATCAATTGAGTTCGGACTGAGACATCTCC 10con. seq
 AACAGACCAATTGAGTTCACTGAGACATCTCC 11con. seq
 AACAGACCAATTGAGTTCACTGAGACATCTCC 19con. seq
 AACAAACCAATTGAGTTCACTGAGACATCTCC 86CON. SEQ
 AACAGACCAATTGAGTTCACTGAGACATCTCC pcrsbe2con. seq

460

470

480

490

ACAATGGAACACGCTAGCCAGATTAAAACTGAG 10con. seq
 ACAATGGAACACGCTAGCCAGATTAAAACTGAG 11con. seq
 ACAATGGAACACGCTAGCCAGATTAAAACTGAG 19con. seq
 ACAATGGAACACGCTAGCCAGATTAAAACTGAG 86CON. SEQ
 ACAATGGAACACGCTAGCCAGATTAAAACTGAG pcrsbe2con. seq

530

540

550

560

GTGTTGAAGAGCTGGATTTGCTTCATCACTAC 10con. seq
 GTGTTGAAGAGCTGGATTTGCTTCATCACTAC 11con. seq
 GTGTTGAAGAGCTGGATTTGCTTCATCACTAC 19con. seq
 GTGTTGAAGAGCTGGATTTGCTTCATCACTAC 86CON. SEQ
 GTGTTGAAGAGCTGGATTTGCTTCATCACTAC pcrsbe2con. seq

600

610

620

630

ATTAATAACTTCTGAAGAGACAATTATTGATGA 10con. seq
 ATTAATAACTTCTGAAGAGACAATTATTGATGA 11con. seq
 ATTAATAACTTCTGAAGAGACAATTATTGATGA 19con. seq
 ATTAATAACTTCTGAAGAGACAATTATTGATGA 86CON. SEQ
 ATTAATAACTTCTGAAGAGACAATTATTGATGA pcrsbe2con. seq

670

680

690

700

GGACTTGGTCAGAACGATTATGAAATAGACCCC 10con. seq
 GGACTTGGTCAGAACGATTATGAAATAGACCCC 11con. seq
 GGACTTGGTCAGAACGATTATGAAATAGACCCC 19con. seq
 GGACTTGGTCAGAACGATTATGAAATAGACCCC 86CON. SEQ
 GGACTTGGTCAGAACGATTATGAAATAGACCCC pcrsbe2con. seq

Fig.10 SHEET 4

50 / 75

	710	720	730
698	CTTTGACAAACTATCGTCAACACACCTTGATTACAGGT		
700	CTTTGACAAACTATCGTCAACACACCTTGATTACAGGT		
700	CTTTGACAAACTATCGTCAACACACCTTGATTACAGGT		
477	CTTTGACAAACTATCGTCAACACACCTTGATTACAGGT		
470	CTTTGACAAACTATCGTCAACACACCTTGATTACAGGT		
	780	790	800
768	ACAAGTATGAGGGTGGTTGGAAGCTTTTCCTCGTGG		
770	ACAAGTATGAGGGTGGTTGGAAGC T TTTTCCTCGTGG		
770	ACAAGTATGAGGGTGGTTGGAAGC T TTTTCCTCGTGG		
547	ACAAGTATGAGGGTGGTTGGAAGCTTTTCCTCGTGG		
540	ACAAGTATGAGGGTGGTTGGAAGCTTTTCCTCGTGG		
	850	860	870
838	AGGTATCACTTACCGTGAGTGGGCTCCTGGTGCCCCAG		
839	AGGTATCACTTACCGTGAGTGGGCTCCTGGTGCCCCAG		
840	AGGTATCACTTACCGTGAGTGGGCTC T GGTGCCCCAG		
617	AGGTATCACTTACCGTGAGTGGGCTCCTGGTGCCCCAG		
610	AGGTATCACTTACCGTGAGTGGGCTCCTGGTGCCCCAG		
	920	930	940
908	GACGCAAATGCTGAC T TATGACTCGGAATGAATTTG		
909	GACGCAAATGCTGACATTATGACTCGGAATGAATTTG		
910	GACGCAAATGCTGACATTATGACTCGGAATGAATTTG		
687	GACGCAAATGCTGACATTATGACTCGGAATGAATTTG		
680	GACGCAAATGCTGACATTATGACTCGGAATGAATTTG		
	990	1000	1010
978	ATGGTTCTCCTGCAATT CCTCATGGGTCCAGAGTGAA		
979	ATGGTTCTCCTGCAATT CCTCATGGGTCCAGAGTGAA		
980	ATGGTTCTCCTGCAATT CCTCATGGGTCCAGAGTGAA		
757	ATGGTTCTCCTGCAATT CCTCATGGGTCCAGAGTGAA		
750	ATGGTTCTCCTGCAATT CCTCATGGGTCCAGAGTGAA		

Fig.10
Sheet 6

Fig.10 SHEET 5

51/75

740	750	760	770	
ATTCACAGTACAAGAAA ATTCACAGTACAAGAAA ATTCACAGTACAAGAAA ATTCACAGTACAAGAAA ATTCACAGTACAAGAAA	ACTGAGGGAGGCATTG ACTGAGGGAGGCATTG ACTGAGGGAGGCATTG ACTGAGGGAGGCATTG ACTGAGGGAGGCATTG	10con. seq 11con. seq 19con. seq 86CON. SEQ pcrsbe2con. seq		
810	820	830	840	
TTATGAAA TTATGAAAAA TTATGAAAAA TTATGAAAAA TTATGAAAAA	CAATGGGTTTCACTCGTAGTGCTAC ATGGGTTTCACTCGTAGTGCTAC ATGGGTTTCACTCGTAGTGCTAC ATGGGTTTCACTCGTAGTGCTAC ATGGGTTTCACTCGTAGTGCTAC	10con. seq 11con. seq 19con. seq 86CON. SEQ pcrsbe2con. seq		
880	890	900	910	
TCAGCTGCCCTCATTGGG TCAGCTGCCCTCATTGGAG TCAGCTGCCCTCATTGGAG TCAGCTGCCCTCATTGGAG TCAGCTGCCCTCATTGGAG	GATTTCAACAAATTGG GATTTCAACAAATTGG GATTTCAACAAATTGG GATTTCAACAAATTGG GATTTCAACAAATTGG	10con. seq 11con. seq 19con. seq 86CON. SEQ pcrsbe2con. seq		
950	960	970	980	
GTGTCTGA GTGTCTGGGAG GTGTCTGGGAG GTGTCTGGGAG GTGTCTGGGAG	GAGATTTCTGCCAAATAATGTGG GAGATTTCTGCCAAATAATGTGG GAGATTTCTGCCAAATAATGTGG GAGATTTCTGCCAAATAATGTGG GAGATTTCTGCCAAATAATGTGG	10con. seq 11con. seq 19con. seq 86CON. SEQ pcrsbe2con. seq		
1020	1030	1040	1050	
GATACGTATGGACACTCC GATACGTATGGACACTCC GATACGTATGGACACTCC GATACGTATGGACACTCC GATACGYATGGACACTCC	CATCAGGTGTTAAGGA CATCAGGTGTTAAGGA CATCAGGTGTTAAGGA CATCAGGTGTTAAGGA CATCAGGTGTTAAGGA	10con. seq 11con. seq 19con. seq 86CON. SEQ pcrsbe2con. seq		

Fig. 10 SHEET 6

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	1060	1070	1080	
1048	TTCCATT CCTGCTTGGATCAACTACTCTTACAGCTT			
1049	TTCCATT CCTGCTTGGATCAACTACTCTTACAGCTT			
1050	TTCCATT CCTGCTTGGATCAACTACTCTTACAGCTT			
827	TTCCATT CCTGCTTGGATCAACTACTC[REDACTED]TACAGCTT			
820	TTCCATT CCTGCTTGGATCAACTACTCTTACAGCTT			
<hr/>				
	1130	1140	1150	
1118	GATCCACCCGAAGAGGGAGAGGTATATCTTCCAACACC			
1119	GATCCACCCGAAGAGGGAGAGGTATATCTTCCAACACC			
1120	GATCCACCCGAAGAGGGAGAGGTATATCTTCCAACACC			
895	GATCCACCCGAAGAGGGAGAGGTATATCTTCCAACACC			
890	GATCCACCCGAAGAGGGAGAGGTAT[REDACTED]TCTTCCAACACC			
<hr/>				
	1200	1210	1220	
1188	ATGAATCTCATATTGGAATGAGTAGTAGTCCGGAGCCTAA			
1189	ATGAATCTCATATTGGAATGAGTAGTAGTCCGGAGCCTAA			
1190	ATGAATCTCATATTGGAATGAGTAGTAGTCCGGAGCCTAA			
965	ATGAATCTCATATTGGAATGAGTAGTAGTCCGGAGCCTAA			
960	ATGAATCTCATATTGGAATGAGTAGTAGTCCGGAGCCTAA			
<hr/>				
	1270	1280	1290	*
1258	TCTTCCTCGCATAAAAAAAGCTTGGGTACAATGCGGT			*
1259	TCTTCCTCGCATAAAAAA-GCTTGGGTACAATGCGCT			
1260	TCTTCCTCGCATAAAAAA-GCTTGGGTACAATGCGCT			
1035	TCTTCCTCGCATAAAAAA-GCTTGGGTACAATGCGCT			
1030	TCTTCCTCGCATAAAAAA-[REDACTED]CTTGGGTACAATGCGGT			*
<hr/>				
	1340	1350	1360	
1328	TGCTAGTTTGGTTATCATGTCACAAATTGGCA			
1328	TGCTAGTTTGGTTATCATGTCACAAATTGGCA			
1329	[REDACTED]GCTAGTTTGGTTATCATGTCACAAATTGGCA			
1104	TGCTAGTTTGGTTATCATGTCACAAATTGGCA			
1099	TGCTAGTTTGGTTATCATGTCACAAATTGGCA			

Fig.10
Sheet 8

Fig. 10 SHEET 7

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1090	1100	1110	1120
CCTGATGAAATTCCATATAATGGAATATATTAT CCTGATGAAATTCCATATAATGGAATATATTAT CCTGATGAAATTCCATATAATGGAATA C ATTAT CCTGATGAAATTCCATATAATGGAATATATTAT CCTGATGAAATTCCATATAATGGAATATATTAT			
10con. seq 11con. seq 19con. seq 86CON. SEQ pcrsbe2con. seq			

1160	1170	1180	1190
CACGGCCAAGAACCAAGTC G TGAGAATAT CACGGCCAAGAACCAAGTCGCTGAGAATAT CACGGCCAAGAACCAAGTCGCTGAGAATAT CACGGCCAAGAACCAAGTCGCTGAGAATAT CACGGCCAAGAACCAAGTCGCTGAGAATAT			
10con. seq 11con. seq 19con. seq 86CON. SEQ pcrsbe2con. seq			

1230	1240	1250	1260
AATTAACTCATACGTGAATTAGAGATGAAGT AATTAACTCATACGTGAATTAGAGATGAAGT AATTAACTCATACGTGAATTAGAGATGAAGT AATTAACTCATACGTGAATTAGAGATGAAGT AATTAACTCATACGTGAATTAGAGATGAAGT			
10con. seq 11con. seq 19con. seq 86CON. SEQ pcrsbe2con. seq			

1300	1310	1320	1330
GCAAATTATGGCTATTCAAGAGCATTCTTATTA GC CAAATTATGGCTATTCAAGAGCATTCTTATTA GCAAATTATGGCTATTCAAGAGCATTCTTATTA GCAAATTATGGCTATTCAAGAGCATTCTTATTA GCAAATTATGGCTATTCAAGAGCATTCTTATTA			
10con. seq 11con. seq 19con. seq 86CON. SEQ pcrsbe2con. seq			

1370	1380	1390	1400
CCAAGCAGCCGTTTGGAACGCCCGACGACCTT CCAAGCAGCCGTTTGGAACGCCCGACGACCTT CCAAGCAGCCGTTTGGAACGCCCGACGACCTT CCAAGCAGCCGTTTGGAACGCCCGACGACCTT CCAAGCAGCCGTTTGGAACGCCCGACGACCTT			
10con. seq 11con. seq 19con. seq 86CON. SEQ pcrsbe2con. seq			

Fig. 10 SHEET 8

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	1410	1420	1430
1398	AAGTCTTGATTGATAAAAGCTCATGAGCTAGGAATTG		
1398	AAGTCTTCGATTGATAAAAGCTCATGAGCTAGGAATTG		
1399	AAGTCTTGATTGATAAAAGCTCATGAGCTAGGAATTG		
1174	AAGTCTTGATTGATAAAAGCTCATGAGCTAGGAATTG		
1169	AAGTCTTGATTGATAAAAGCTCATGAGCTAGGAATTG		
	1480	1490	1500
1468	CAAATAATACCTTAGATGGACTGAACATGTTGACGG		
1468	CAAATAATACCTTAGATGGACTGAACATGTTGACGG		
1469	CAAATAATACCTTAGATGGACTGAACATGTTGAC TG		
1244	CAAATAATACCTTAGATGGACTGAACATGTTGACGG		
1239	CAAATAATACCTTAGATGGACTGAACATGTTGACGG		
	1550	1560	1570
1538	TGGTTATCATTGGATGTGGGATT T CCGCCTCTTAAC		
1538	TGGTTATCATTGGATGTGGGATT T CCGCCTCTTAAC		
1539	TGGTTATCATTGGATGTGGGATT T CCGCCTCTTAAC		
1314	TGGTTATCATTGGATGTGGGATT T CCGCCT T TTAAC		
1309	TGGTTATCATTGGATGTGGGATT T CCGCCTCTTAAC		
	1620	1630	1640
1608	TCAAATGCGAGATGGTGGTTGGATGAGTTCAAATTG		
1607	TCAAATGCGAGATGGTGGTTGGATGAGTTCAAATTG		
1609	TCAAATGCGAGATGGTGGTTGGAT G GTTCAAATTG		
1384	TCAAATGCGAGATGGTGGTTGGATGAGTTCAAATTG		
1379	TCAAATGCGAGATGGTGGTTGGATGAGTTCAAATTG		
	1690	1700	1710
1678	TGT C TACTCACCAACGGATTATCGGTGGGATTCACTGG		
1677	TGTATACTCACCAACGGATTATCGGTGGGATTCACTGG		
1679	TGTATA T CACCAACGGATTATCGGTGGGATTCACTGG		
1454	TGTATACTCACCAACGGATTATCGGTGGGATTCACTGG		
1449	TGTATACTCACCAACGGATTATCGGTGGGATTCACTGG		

Fig. 10
Sheet 10

Fig. 10 SHEET 9

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1440	1450	1460	1470
<hr/>			
TTGTTCTCATGGACATTGTTACAGCCATGCAT			10con. seq
TTGTTCTCATGGACATCGTTACAGCCATGCAT			11con. seq
TTGTTCTCATGGACATTGTTACAGCCATGCAT			19con. seq
TTGTTCTCATGGACATTGTTACAGCCATGCAT			86CON. SEQ
TTGTTCTCATGGACATTGTTACAGCCATGCAT			pcrsbe2con. seq
<hr/>			
1510	1520	1530	1540
<hr/>			
CACAGATAGTTTACTTCACTCTGGAGCTCG			10con. seq
CACCGATAGTTTACTTCACTCTGGAGCTCG			11con. seq
CACCGATAGTTTACTTCACTCTGGAGCTCG			19con. seq
CACCGATAGTTTACTTCACTCTGGAGCTCG			86CON. SEQ
CACAGATAGTTTACTTCACTCTGGAGCTCG			pcrsbe2con. seq
<hr/>			
1580	1590	1600	1610
<hr/>			
TATGGAAACTGGGAGGTACTTAGGTATCTTCTC			10con. seq
TATGGAAACTGGGAGGTACTTAGGTATCTTCTC			11con. seq
TATGGAAACTGGGAGGTACTTAGGTATCTTCTC			19con. seq
TATGGAAACTGGGAGGTACTTAGGTATCTTCTC			86CON. SEQ
TATGGAAACTGGGAGGTACTTAGGTATCTTCTC			pcrsbe2con. seq
<hr/>			
1650	1660	1670	1680
<hr/>			
ATGGATTAGATTGATGGTGTGACATCAATGA			10con. seq
ATGGATTAGATTGATGGTGTGACATCAATGA			11con. seq
ATGGATTAGATTGATGGTGTGACATCAATGA			19con. seq
ATGGATTAGATTGATGGTGTGACATCAATGA			86CON. SEQ
ATGGATTAGATTGATGGTGTGACATCAATGA			pcrsbe2con. seq
<hr/>			
1720	1730	1740	1750
<hr/>			
GAAC TAC GAG GA A AT CTT GG ACT CGC A ACT GA			10con. seq
GAAC TAC GAG GA A AT CTT GG ACT CGC A ACT GA			11con. seq
GAAC TAC GAG GA A AT CTT GG ACT CGC A ACT GA			19con. seq
GAAC TAC GAG GA A AT CTT GG ACT CGC A ACT GA			86CON. SEQ
GAAC TAC GAG GA A AT CTT GG ACT CGC A ACT GA			pcrsbe2con. seq

Fig. 10 SHEET 10

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	1760	1770	1780
1748	TGTGGATGCTGTTGTATCTGATGCTGGTCAACGAT		
1747	TGTGGATGCTGTTGTATCTGATGCTGGTCAACGAT		
1749	TGTGGATGCTGTTGTATCTGATGCTGGTCAACGAT		
1524	TGTGGATGCTGTTGTATCTGATGCTGGTCAACGAT		
1519	TGTGGATGCTGTTGTATCTGATGCTGGTCAACGAT		
	1830	1840	1850
1818	ATTGGTAAAGATGTTAGCGGAATGCCGACATTTGTG		
1817	ATTGGTAAAGATGTTAGCGGAATGCCGACATTTGTA		
1819	ATTGGTAAAGATGTTAGCGGAATGCCGACATTTGTA		
1594	ATTGGTAAAGATGTTAGCGGAATGCCGACATTTGTA		
1589	ATTGGTAAAGATGTTAGCGGAATGCCGACATTTGTA		
	1900	1910	1920
1888	ATCGGCTGCATATGGCAATTGCTGATAAAATGGATTGA		
1887	ATCGGCTGCATATGGCAATTGCTGATAAAATGGATTGA		
1889	ATCGGCTGCATATGGCAATTGCTGATAAAACGGATTGA		
1664	ATCGGCTGCATATGGCAATTGCTGATAAAATGGATTGA		
1659	ATCGGCTGCATATGGCAATTGCTGATAAAATGGATTGA		
	1970	1980	1990
1958	GGGTGATATTGTTCATACACTGACAAATAGAAGATGG		
1957	GGGTGATATTGTTCATACACTGACAAATAGAAGATGG		
1959	GGGTGATATTGTTCATACACTGACAAATAGAAGATGG		
1734	GGGTGATATTGTTCATACACTGACAAATAGAAGATGG		
1729	GGGTGATATTGTTCATACACTGACAAATAGAAGATGG		
	2040	2050	2060
2028	GATCAAGCTCTAGTCGGTGATAAAACTATAGCATTCT		
2027	GATCAAGCTCTAGTCGGTGATAAAACTATAGCATTCT		
2029	GATCAAGCTCTAGTCGGTGATAAAACTATAGCATTCT		
1804	GATCAAGCTCTAGTCGGTGATAAAACTATAGCATTCT		
1799	GATCAAGCTCTAGTCGGTGATAAAACTATAGCATYCT		

Fig.10
Sheet 12

Fig. 10 SHEET 11

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1790	1800	1810	1820
CTTATTCA <u>GGG</u> CTTTCCCAGATGCAATTACC		10con. seq	
CTTATTCA <u>GGG</u> CTTTCCCAGATGCAATTACC		11con. seq	
CTTATTCA <u>GGG</u> CTTTCCCAGATGCAATTACC		19con. seq	
CTTATTCA <u>GGG</u> CTTTCCCAGATGCAATTACC		86CON. SEQ	
CTTATTCA <u>GGG</u> CTTTCCCAGATGCAATTACC		pcrsbe2con. seq	
1860	1870	1880	1890
TTCCCGTTCAAGATGGGGGTGTTGGCTTGACT		10con. seq	
TTCCCGTTCAAGATGGGGGTGTTGGCTTGACT		11con. seq	
TTCCCGTTCAAGATGGGGGTGTTGGCTTGACT		19con. seq	
TTCCCGTTCAAGATGGGGGTGTTGGCTTGACT		86CON. SEQ	
TTCCCGTTCAAGATGGGGGTGTTGGCTTGACT		pcrsbe2con. seq	
1930	1940	1950	1960
GTTGCTCAAGAACGGGATGAGGATTGGAGAGT		10con. seq	
GTTGCTCAAGAACGGGATGAGGATTGGAGAGT		11con. seq	
GTTGCTCAAGAACGGGATGAGGATTGGAGAGT		19con. seq	
GTTGCTCAAGAACGGGATGAGGATTGGAGAGT		86CON. SEQ	
GTTGCTCAAGAACGGGATGAGGATTGGAGAGT		pcrsbe2con. seq	
2000	2010	2020	2030
TCGGAAAAGTGTGTTCATACGCTGAAAGTCAT		10con. seq	
TCGGAAAAGTGTGTTCATACGCTGAAAGTCAT		11con. seq	
TCGGAAAAGTGTGTTCATACGCTGAAAGTCAT		19con. seq	
TCGGAAAAGTGTGTTCATACGCTGAAAGTCAT		86CON. SEQ	
TCGGAAAAGTGTGTTCATACGCTGAAAGTCAT		pcrsbe2con. seq	
2070	2080	2090	2100
GGCTGATGGACAAGGATATGTATGATTTATGG		10con. seq	
GGCTGATGGACAAGGATATGTATGATTTATGG		11con. seq	
GGCTGATGGACAAGGATATGTATGATTTATGG		19con. seq	
GGCTGATGGACAAGGATATGTATGATTTATGG		86CON. SEQ	
GGCTGATGGACAAGGATATGTATGATTTATGG		pcrsbe2con. seq	

Fig. 10 SHEET 12

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	2110	*	2120	2130
2098	CTCTGGATAGACCGTCAACATCATTAAATAGATCGTGG			
2097	CTCTGGATAGACCGC G CAACATCATTAAATAGATCGTGG			
2099	CTCTGGATAGACCGTCAACATCATTAAATAGATCGTGG			
1874	CTCTGGATAGACCGC G CAACATCATTAAATAGATCGTGG			
1869	CTCTGGATAGACCGY C AACAYCATTAATAGATCGTGG			
	*			
	2180	2190	2200	
2168	TATGGGATTAGGAGGAGAAGGGTACCTAAATTCATG			
2167	TATGGGATTAGGAGGAGAAGGGTACCTAAATTCATG			
2169	TATGGGATTAGGAGGAGAAGGGTACCTAAATTCATG			
1944	TATGGGATTAGGAGGAGAAGGGTACCTAAATTCATG			
1939	TATGGGATTAGGAGGAGAAGGGTACCTAAATTCATG			
	2250	*	2260	2270
2238	TTCCCTAGGGCTGAACAAACACCTCTCTGATGGCTCAG			
2237	TTCCCTAGGGCTGA G C ACACCTTCTGATGGCTCAG			
2239	TTCCCTAGGGCTGAACAAACACCTCTCTGATGGCTCAG			
2014	TTCCCTAGGGCTGAACAAACACCTCTGATG A CTCAG			
2009	TTCCCTAGGGCTGA R ACACACCTCTGATGGCTCAG			
	*			
	2320	2330	2340	
2308	GCAGACGGAGATTGACCTGGGAGATGCAGAAATATT			
2307	GCAGACGGAGATTGACCTGGGAGATGCAGAAATATT			
2309	GCAGACGGAGATTGACCTGGGAGATGCAGAAATATT			
2084	GCAGACGGAGATTGACCTGGGAGATGCAGAAATATT			
2079	GCAGACGGAGATTGACCTGGGAGATGCAGAAATATT			
	2390	2400	2410	
2378	TATGCAGTATCTTGAAGATAAATATGAGTTATGACT			
2377	TATGCAGTATCTTGAAGATAAATATGAGTTATGACT			
2379	TATGCAGTATCTTGAAGATAAATATGAGTTATGACT			
2154	TATGCAGTATCTTGAAGATAAATATGAGTTATGACT			
2149	TATGCAGTATCTTGAAGATAAATATGAGTTATGACT			

Fig.10
Sheet 14

Fig. 10 SHEET 13

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2140	2150	2160	2170
GATAGCATTACACAAGATGATTAGGCTTGTAAAC			10con. seq
GATAGCATTGCACAAGATGATTAGGCTTGTAAAS			11con. seq
GATAGCATTGCACAAGATGATTAGGCTTGTAAAC			19con. seq
GATAGCATTGCACAAGATGATTAGGCTTGTAAAC			86CON. SEQ
GATAGCATTGCACAAGATGATTAGGCTTGTAAAC			pcrsbe2con. seq

2210	2220	2230	2240
GGAAATGAATTGGCCACCCCTGAGTGGATTGAT			10con. seq
GGAAATGAATTGGCCACCCCTGAGTGGATTGAT			11con. seq
GGAAATGAATTGGCCACCCCTGAGTGGATTGAT			19con. seq
GGAAATGAATTGGCCACCCCTGAGTGGATTGAT			86CON. SEQ
GGAAATGAATTGGCCACCCCTGAGTGGATTGAT			pcrsbe2con. seq

2280	2290	2300	2310
TAATTCCCAGAAACCAATTCAAGTTATGATAAAT			10con. seq
TAATTCCCGGAAACCAATTCAAGTTATGATAAAT			11con. seq
TAATCCCCGGAAACCAATTCAAGTTATGATAAAT			19con. seq
TAATTCCCGGAAACCAATTCAAGTTATGATAAAT			86CON. SEQ
TAATTCCCGGAAACCAATTCAAGTTATGATAAAT			pcrsbe2con. seq

2350	2360	2370	2380
AAGATACCGTGGGTTGCAAGAATTGACCGGGC			10con. seq
AAGATACCATGGGTTACAAGAATTGACGGGC			11con. seq
AAGATACCGTGGGTTGCAAGAATTGACCGGGC			19con. seq
AAGATACCGTGGGTTGCAAGAATTGACCGGGC			86CON. SEQ
AAGATACCATGGGTTGCAAGAATTGACCGGGC			pcrsbe2con. seq

2420	2430	2440	2450
TCAGAACACCAAGTTCATATCACGAAAGGATGAA			10con. seq.
TCAGAACACCAAGTTCATATCACGAAAGGATGAA			11con. seq
TCAGAACACCAAGTTCATATCACGAAAGGATGAA			19con. seq
TCAGAACACCAAGTTCATATCACGAAAGGATGAA			86CON. SEQ
TCAGAACACCAAGTTCATATCACGAAAGGATGAA			pcrsbe2con. seq

Fig. 10 SHEET 14

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	2460	2470	*	2480
2448	GGAGATAGGATGATTGTATTGAAAAAGGAAACCTAG			
2447	GGAGATAGGATGATTGTATTGAAA G AGGAAACCTAG			
2449	GGAGATAGGATGATTGTATTGAAAAAGGAAACCTAG			
2224	GGAGATAGGATGATTGTATTGAAAAAGGAAACCTAG			
2219	GGAGATAGGATGATTGTATTGAAA R AGGAAACCTAG			
	*			
	2530	2540		2550
2518	ATTCAGACTATCGCATAGGCTGCCCTGAAGCCTGGAAA			
2517	ATTCAGACTATCGCATAGGCTGCCCTGAAGCCTGGAAA			
2519	ATTCAGACTATCGCATAG C CTGCCCTGAAGCCTGGAAA			
2294	ATTCAGACTATCGCATAGGCTGCCCTGAAGCCTGGAAA			
2289	ATTCAGACTATCGCATAGGCTGCCCTGAAGCCTGGAAA			
	2600	2610		2620
2588	TTTTGGTGGCTTCGGGAGAATTGATCATAATGCCGAA			
2587	TTTTGGTGGCTTCGGGAGAATTGATCATAATGCCGAA			
2589	TTTTGGTGGCTTCGGGAGAATTGATCATAATGCCGAA			
2364	TTTTGGTGGCTTCGGGAGAATTGATCATAATGCCGAA			
2359	TTTTGGTGGCTTCGGGAGAATTGATCATAATGCCGAA			
	2670	2680	*	2690
2658	CCTCGTTCAATTATGGTGTATGCACCTAGTAGAACAG			
2657	CCT I GTTCAATTATGGTGTATGCACCTAGTAGAACAG			
2659	CCTCGTTCAATTATGGTGTATGCACCT T GTA A ACAG			
2434	CCTCGTTCAATTATGGTGTATGCACCT T GTAACAG			
2429	CCTCGTTCAATTATGGTGTATGCACCTAGTAGAACAG			
	*			
	2740	2750		2760
2722	- AAGAAGAAGAAGAAGAAGAAGTAGCAGTAGT			
2722	- AGAAGTAGCAGTAGT			
2729	AAGAAGAAGAAGAAGAAGAAGAAGTAGCAG C AGT			
2501	AAGAAGAAGAAGAAGAAGAAGAAGTAGCAGTAGT			
2499	N AGAAGAAGAAGAAGAAN -			

Fig. 10
Sheet 16

Fig. 10 SHEET 15

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2490	2500	2510	*	2520	
TTTTGTCTTAATTTCACTGGACAAAAAGCT					10con. seq
TTTTCGTCTTAATTTCACTGGACAAA[TAGCT					11con. seq
TTTTGTCTTAATTTCACTGGACAAAAAGCT					19con. seq
TTTTGTCTTAATTTCACTGGACAAAAAGCT					86CON. SEQ
TTTTGTCTTAATTTCACTGGACAAA[TAGCT					pcrsbe2con. seq
*					
2560	2570	2580		2590	
ATACAAGGTTGCCTTGGACTCAGATGATCCACT					10con. seq
ATACAAGGTTG[CTTGGACTCAGATGATCCACT					11con. seq
ATACAAGGTTGCCTTGGACTCAGATGATCCACT					19con. seq
ATACAAGGTTGCCTTGGACTCAGATGATCCACT					86CON. SEQ
ATACAAGGTTGCCTTGGACTCAGATGATCCACT					pcrsbe2con. seq
2630	*	2640	*	2650	2660
TATTCACCTTGAAAGGATGGTATGATGATCGT					10con. seq
TATTCACCT[GTAAAGGAT[GTATGATGATCGT					11con. seq
TATTCACCTTGAAAGGATGGTATGATGATCGT					19con. seq
TATTCACCTTGAAAGGATGGTATGATGATCGT					86CON. SEQ
TATTCACCT[GTAAAGGAT[GTATGATGATCGT					pcrsbe2con. seq
*		*			
2700	2710	2720		2730	
CAGTGGTCTATGCACTAGTAGACAAAG-----					10con. seq
CAGTGGTCTATGCACTAGTAGACAAACT-----					11con. seq
CAGTGGTCTATGCACTAGTAGACAAAGAAGAAG					19con. seq
CAGTGGTCTATGCACTAGTAGACAAAG-----AAG					86CON. SEQ
CAGTGGTCTATGCACTAGTAGACAAANTAGAAG					pcrsbe2con. seq
2770	2780	2790		2800	
AGAAGAAGTAGTAGTAGAAGAAGAACGAA					10con. seq
AGAAGAACCCAT[TG-----AAGAACGAA					11con. seq
AGAAGAAGTAGTAGTAGAAGAAGAACGAA					19con. seq
AGAAGAAGTAGTAGTAGAAGAAGAACGAA					86CON. SEQ
-----CCGNNGAAGAACGAA-----					pcrsbe2con. seq

Fig. 10 SHEET 16

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	2810	2820	2830
2786	CTTGTGATCGCGTTGAAAGATTGAACGCCACATAGA		
2764	CTTGTGATCGCGTTGAAAGATTGAACGTTACTTGG-		
2799	CTTGTGATCGCGTTGAAAGATTGAACGCTACATAGA		
2571	CTTGTG		
2529	[REDACTED]		
	2880	2890	2900
2856	CTTGGCGGAATTTCATGTGACAACA-GGTTTGCAATT		
2829	CTTGGCGGAATTGATGTGACAACAAGGTTTGCAAGTT		
2869	CTTGGCGGAATTTCATGTGACACAA-GGTTTGCAATT		
2576	[REDACTED]		
2529	[REDACTED]		
	2950	2960	2970
2925	GAGATGAAGTGCTGAACAAAAACATATGAAAATCGA		
2899	GAGATGAAGTGCTGAACAAA--CATATGAAAATCGA		
2938	GAGATGAAGTGCTGAACAAA--CATATGAAAATCGA		
2576	[REDACTED]		
2529	[REDACTED]		
	3020	3030	
2995	CCTGCAG-----CC		
2967	CCTGCAG-----CC		
3006	CCTGCAGGCCGGGGACCCCTTAGTTCT		
2576	[REDACTED]		
2529	[REDACTED]T		

Fig. 10
Sheet 18

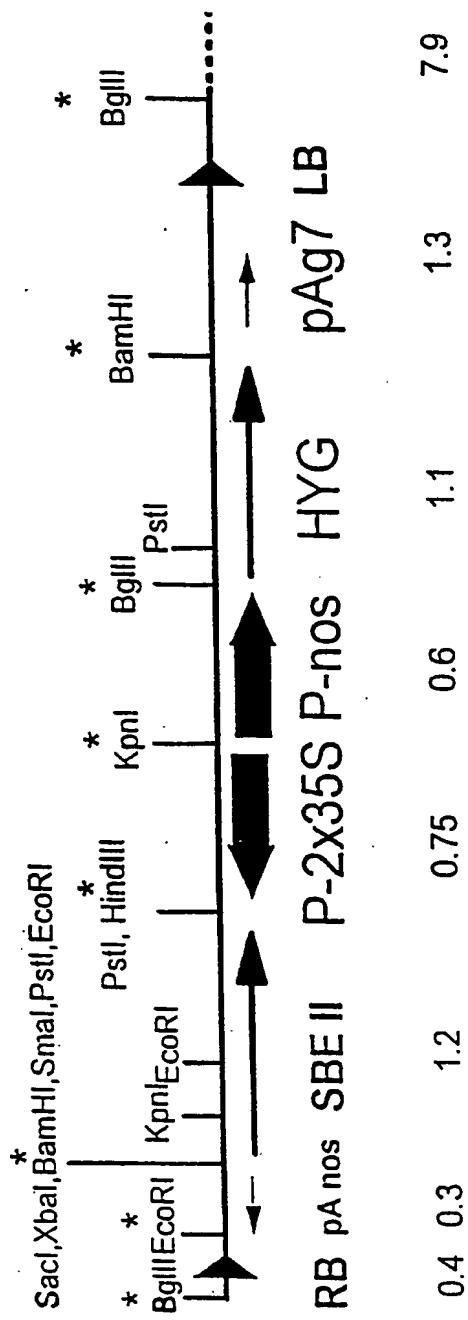
Fig. 10 SHEET 17

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2840	2850	2860	2870
GCTTCTTGACGTATCTGGCAATATTGCAT[AGT]			10con. seq
[TCATCCACATA--GAGCTTCTTGACATCAGT]			11con. seq
GCTTCTTGACGTATCTGGCAATATTGCATCAGT			19con. seq
			86CON. SEQ
			pcrsbe2con. seq
2910	2920	2930	2940
CTTTCCACTATTAGTAGTGCAACGATATACGCA			10con. seq
CTTTCCACTATTAGTAGT[CCACCGATATACGCA]			11con. seq
CTTTCCACTATTAGTAGTGCAACGATATACGCA			19con. seq
			86CON. SEQ
			pcrsbe2con. seq
2980	2990	3000	3010
TGAATTATGTCGAATGCTGGGACGATCGAATT			10con. seq
TGAATTATGTCGAATGCTGGGACGATCGAATT			11con. seq
TGAATTATGTCGAATGCTGGGACGATCGAATT			19con. seq
			86CON. SEQ
			pcrsbe2con. seq
			10con. seq
			11con. seq
			19con. seq
			86CON. SEQ
			pcrsbe2con. seq

Fig. 10 SHEET 18

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0.4	0.3	1.2	0.75	0.6	1.1	1.3	7.9
-----	-----	-----	------	-----	-----	-----	-----

Fig. 11

65/75

TCATTAAGGAGAAATTAAACTATGAGAGGGATCTCACCATCACCATGGATCT
 |
 BstX I
 AGTAAATTCTCCTTTAATTGATACTCTTAGAGTGGTAGTGGTACCCCTAGA
 M R G S H H H H H G I

 Nco I
 |
 TGGCTGAAAAGCTCTTACAAATTCCGAATTCCGACCTCTACAGTTGCAGGCCATCGGGGA
 ACCGACTTTTCAGAAGGAATGTAAAGGCTTAAGGCTAAGGCTGGAAAGATGTCAACCGTGTAGCCCCCT
 L A E K S S Y N S E F R P S T V A A S G

 EcoRI
 |
 AAGTCCCTTGCCCTGGAACCCAGAGTGTAGCTCATCCTCAACAAACCAATTGAGT
 TTCAAGGAACACGGACCTGGGTCTCACTATCGAGGAGTAGGAGTTGTTGGTTAAACTCA
 K V L V P G T Q S D S S S S T N Q F E

 TCACTGAGACATCTCAGAAAATTCCCCAGGATCAACTGATGTAGATAGTTCAACAAATGG
 AGTGAECTCTGAGGGTCTTTAAGGGGTCTAGTTGACTACATCTCAAGTTGTTAC
 F T E T S P E N S P A S T D V D S S T M

SUBSTITUTE SHEET (RULE 26)

Fig. 12
SHEET 1

66/75

AACACGCTAGCCAGATTAACACTGAGAACGATGACGTGAGCGTCAAGTGATCTTACAG
 TTGGCGATGGTCTAATTGACTCTTGCTACTGCACACTCGGCAGTTACTAGAATGTC
 E H A S Q I K T E N D D V E P S S D L T

GAAGTGTGAAGAGCTGGATTGGCTTCATCACTACAACACTACAAGAAGGTGGTAAACTTGG
 CTTCACACCTTCTGACCTAAACGAAGTAGTGTGATGTTGATGTTCCACCATTGACC
 G S V E E L D F A S S L Q L Q E G G K L

AGGAGTCTAACATTAAATACTTCTGAAGAGACAATTATTGATGAATCTGATAGGATCA
 TCCTCAGATTGTAATTGAAAGACTTCTCTGTTAATAACTACTTAGACTATCCTAGT
 E E S K T L N T S E E T I I D E S D R I

GAGAGGGGCATCCCTCACCTGGACTTGGTCAAGAGATTATGAAATAGACCCCCCTT
 CTCTCTCCCGTAGGGAGGCTGGACCTGAACCAAGTCTAAATACTTTATCTGGGGAAA
 R E R G I P P G L G Q K I Y E I D P L

Hinc II

TGACAAACTATCGTCAACACCTTGATTACAGGTATTACAGTACAAGAAACTGAGGGAGG
 ACTGTTGATAGCAGTTGTGGAACATAATGTCCATAAGTGTCACTGTTGACTCCCTCC
 L T N Y R Q H L D Y R Y S Q Y K K L R E

Fig.12
SHEET 2

67/75

Hind III

CAATTGACAAAGTATGAGGGTGGTTGGAAAGCCTTTCTCGTGTTATGAAAAAATGGGT
 GTTAACTGTTCATACTCCCACCAACCTTCGAAAAAGAGGACCCAATACTTTTACCCAA
 A I D K Y E G G L E A F S R G Y E K M G

Pvu II

TCACTCGTAGTGCTACAGGTATCACCTTACCGTGAGTGCGCTCCTGGTGCCCAGTCAGCTG
 AGTGAGCATCACGATGTCATAGTGAAATGGCACTCACCCGAGGACCCACGGTCAGTCGAC
 F T R S A T G I T Y R E W A P G A Q S A

CCCTCATGGAGATTCAACAATTGGGACGCAAATGCTGACATTATGACTCGGAATGAAI
 GGGAGTAACCTCTAAAGTTGTTAACCCCTGCCTTACGACTGTAATACTGAGCCTTACTTA
 A L I G D F N N W D A N A D I M T R N E

TTGGTGTCTGGAGATTTCCTGCCAAATAATGTTGGATGGTTCTCCTGCAATTCTCATG
 AACACAGACCCCTCTAAAGACGGTTTACACCTACCAAGAGGACGTTAAGGGAGTAC
 F G V W E I F L P N N V D G S P A I P H

Fig 12
SHEET 3

68/75

SnaBI

GGTCCAGAGTGAAGATACTGTATGGACACTCCATCAGGTTAAGGATTCCATTCCCTGCTT
840
CCAGGTCTCACCTCTATGCCATACCTGTGAGGTAGTCACAAATTCCAAGGTAAAGGACGAA

G S R V K I R M D T P S G V K D S I P A

GGATCAACTACTCTCACAGCTTCCTGTGAAATTCCATATAATGGAATAATTATGATC
900
CCTAGTTGATGAGAAGTGTCAAGGACTACTTAAGGTATTTAACCTTATAATACTAG

W I N Y S S Q L P D E I P Y N G I Y Y D

CACCGAAGGAGAGGTATATCTTCCAAACACCCACGGCCAAGAAACCAAGTGGCTGA
960
GTGGGCCTCTCTCCATAGAAGGTGTGGTGCCTTCTTTGGTTTCAGGGACT

P P E E R Y I F Q H P R P K K P K S L

GAATAATGAATCTCATATTGAAATGAGTAGTCCGGAGCCTAAATTAACCTCATACGTGA
1020
CTTATATACTTAGAGTATAACCTTACTCATCAGGCCCTGGATTAAATTGAGTATGCACT

R I Y E S H I G M S S P E P K I N S Y V

SUBSTITUTE SHEET (RULE 26)

Fig. 12
SHEET 4

69/75

Xmn I HinD III

```

ATTTAGAGATGAAGTTCTTCGCATAAAAAGCTTGGGTACAAATGGGGTGCAAATT 1080
TAAATCTCAACTCAAGAAGGAGCGTATTTTCAACCCATGTTACGCCACGTTAAC
N F R D E V L P R I K K L G Y N A V Q I

```

```

TGGCTATTCAAGAGCATTCTTATTATGCTAGTTGGTTATCATGTCACAAATTTTTTG 1140
ACCGATAAGTTCTCGTAAGAATAATACGATCAAAACCAATAGTACAGTGTTAAAAAAC
M A I Q E H S Y A S F G Y H V T N F F

```

```

CACCAAGGCCGTTTGGAACGCCGACGCCCTTAAGTCCTTGATTGATAAAGCTCATG 1200
GTGGTTCGCAAAACCTTGGGGCTGCTGGATTCAAACTAACATTTCGAGTAC
A P S S R F G T P D D L K S L I D K A H

```

Nsi I

```

AGCTTAGGAATTGTTCTCATGGACATTGTTCACAGGCCATGCATCAAATAACTTTAG 1260
TCGATCCTTAACACAAGAGTACCTGTAACAAAGTGTCGGTAGTTTATTGAAATC
E L G I V V L M D I V H S H A S N N T L

```

Fig. 12
SHEET 5

70/75

Sac I

ATGGACTGAACATGTTGACGGCACCGATAAGTTTACTTTCACTCTGGAGCTCGTGGTT 1320
 TACCTGACTTGTACAACAACTGCCGTGGCTATCAACAAATGAAAGTGAGACCTCGAGCACCAA
 D G L N M F D G T D S C Y F H S G A R G

ATCATGGATGGATTCCGCCTTTTAACATGGAAACTGGGGAGGTACTTAGGTATC 1380
 TAGTAACCTACCCCTAACGGGGGGAAAAATTGATACCTTGCACCTCCATGAATCCATAG
 Y H W M W D S R L F N Y G N W E V L R Y

TTCTCTCAAATGCGAGATGGGGTTGGATGAGTTCAAATTGATGGATTAGATTGATG 1440
 AAGAGAGTTAACGCTTACACCAACCTACTCAAGTTAACTACCTAAATCTAAACTAC
 L L S N A R W W L D E F K F D G F R F D

G V T S M M Y T H H G L S V G F T G N Y
 GTGTGACATCAATGATGATGATACCTACCCACGGATTATCGGTGGGATTCACTGGGAACCTACG 1500
 CACACTGTAGTTACTACATATGAGTGGTGCCTATAGCCACCCCTAAGTGACCCCTTGATGC

Fig. 12
SHEET 6

SUBSTITUTE SHEET (RULE 26)

71/75

Hinc II

AGGAATACCTGGACTCGCAACTGATGGATGCTGGTGTATCTGATGCTGGTCACG
 TCCTTATGAAACCTGAGCGTTGACTACACCTACGACAACACATAGACTACGACCAGTTGC
 E E Y F G L A T D V D A V V Y L M L V N

ATCTTATTCAATTGGCTTTCCAGATGCAATTACCATGGTGAAGATGTTAGCGGAAATGC
 TAGAATAAGTACCGAAAAGGGTCTACGTTAACGGTAAACTCTACATCGCCCTTACG
 D L I H G L F P D A I T I G E D V S G M

CGACATTTGTATTCCCGTTCAAGATGGGGGTGTTGGCTTTGACTATGGCTGCATATGG
 GCTGTAAACATAAGGGCAAGTTCTACCCCCACAACCGAAACTGATAAGCCGACGTATACC
 P T F C I P V Q D G G V G F D Y R L H M

CAATTGCTGATAATGGATTGAGTTGCTCAAGAACGGGATGAGGATTGGAGAGTGGGTG
 GTTAACCGACTATTACCTAACGAGTTCTTGCCTACTCCTAACCTCTCACCCAC
 A I A D K W I E L L K K R D E D W R V G

ATATTGTTCAACTGACAAATAGAAGATGGTCGGAAAAGTGTTTACGGCTGAAA
 TATAACAAGTATGTTGACTGTTTACCAACAAAGTATGGACTTT
 D I V H T L T N R R W S E K C V S Y A E

Fig 12
SHEET 7

72/75

GTCATGATCAAGCTCTAGTCGGTATAAAACTATAGCATCTGGCTGATGGACAAGGATA 1860
 CAGTACTAGTTCGAGATCAGCCACTATTGTATCGTAAGACCCGACTACCTGTTCTAT
 S H D Q A L V G D K T I A F W L M D K D

TGTATGATTATGGCTCTGGATAAGACGCCAACATCATTAATAGATCGTGGATAGCAT 1920
 ACATACTAAATAACCGAGACCTATCTGGCGGTTGTAGTATTATCTAGCACCCCTATCGTA
 M Y D F M A L D R P P T S L I D R G I A

Asp 718
Kpn I

TGCACAAAGATGATTAGGCTTGTAACTATGGGATTAGGAGGAAGGGTACCTAAATTCA 1980
 ACGTGTTCTACTAATCGAACATTGATAACCTATCCTCTCCATGGATTAAAGT
 L H K M I R L V T M G L G G E G Y L N F

EcoR I

TGGAAATGAATTGGCCACCCCTGAGTGGATTGATTCCCTAGGGCTGAACAACACCTCT 2040
 ACCCTTTACTTAAGCCGGGGACTCACCTAACTAAAGGGATCCGACTTGTGGAGA
 M G N E F G H P E W I D F P R A E Q H L

Fig 12
SHEET 8

SUBSTITUTE SHEET (RULE 26)

73/75

CTGATCAGTAATTCCGGAAACCAATTCAAGTTATGATAAATGCAGACGGAGATTTG 2100
 GACTACTGAGTCATTAAGGGCCTTTGGTTAACGTTAACATTTACGTCTGCCCTCTAAAC
 S D S V I P G N Q F S Y D K C R R R F

Ssp I

ACCTGGGAGATTCAGAAATTAAAGATAACCGTGGGTTGCAAGAAATTGACCGGGCTATGC 2160
 TGGACCCCTCTACGTCTTAAATTCATGGCACCCAAACGTTCTAAACTGGCCGATACG
 D L G D A E Y L R Y R G L Q E F D R A M
 AGTATCTTGAAGATAAAATATGAGTTTATGACTTCAGAACCCAGTCAATATCACGAAAGG 2220
 TCATAGAACTCTTAACTCAAAATACTGAAGTCTGGTCAAGTATAGTGCTTCC
 Q Y L E D K Y E F M T S E H Q F I S R K
 ATGAAGGAGATAGGTATTGAAAAAGGAAACCTAGTTTGTCTTTAATTTC 2280
 TACTTCCTCATCCTACTAACATAAACCTTTCTGGATCAAACAGAAAATTAAAG
 D E G D R M I V F E K G N L V F V F N F
 ACTGGACAAAGCTTCAAGACTATGGCATAGGCTGCCCTGAAGGCTGGAAAATACAAGG 2340
 TGACCTGTTTCGATAAGCTGTAGCGTATCCGACGGACTTGGACCTTTATGTTCC
 H W T K S Y S D Y R I G C L K P G K Y K

SUBSTITUTE SHEET (RULE 26)

Fig. 12
SHEET 9

74/75

TTGCCTGGACTCAGATGATCCACTTTGGCTCGGGAGAATTGATCATATAATGCCG
 AACGGAACCTGAGTCTACTAGGTGAAACCAACCGAAGCCCTCTTAACTAGTATTACGGC
 V A L D S D P L F G G F G R I D H N A

SspI

AATATTCACCTTGAGGTGGTATGATCGTCCTCGTTCAATTATGGTGTATGCAC 2460
 TTATAAGTGGAAACTTCCCTACCATACTACTAGCAGGAGCAAGTTAACACATACGTG
 E Y F T F E G W Y D D R P R S I M V Y A

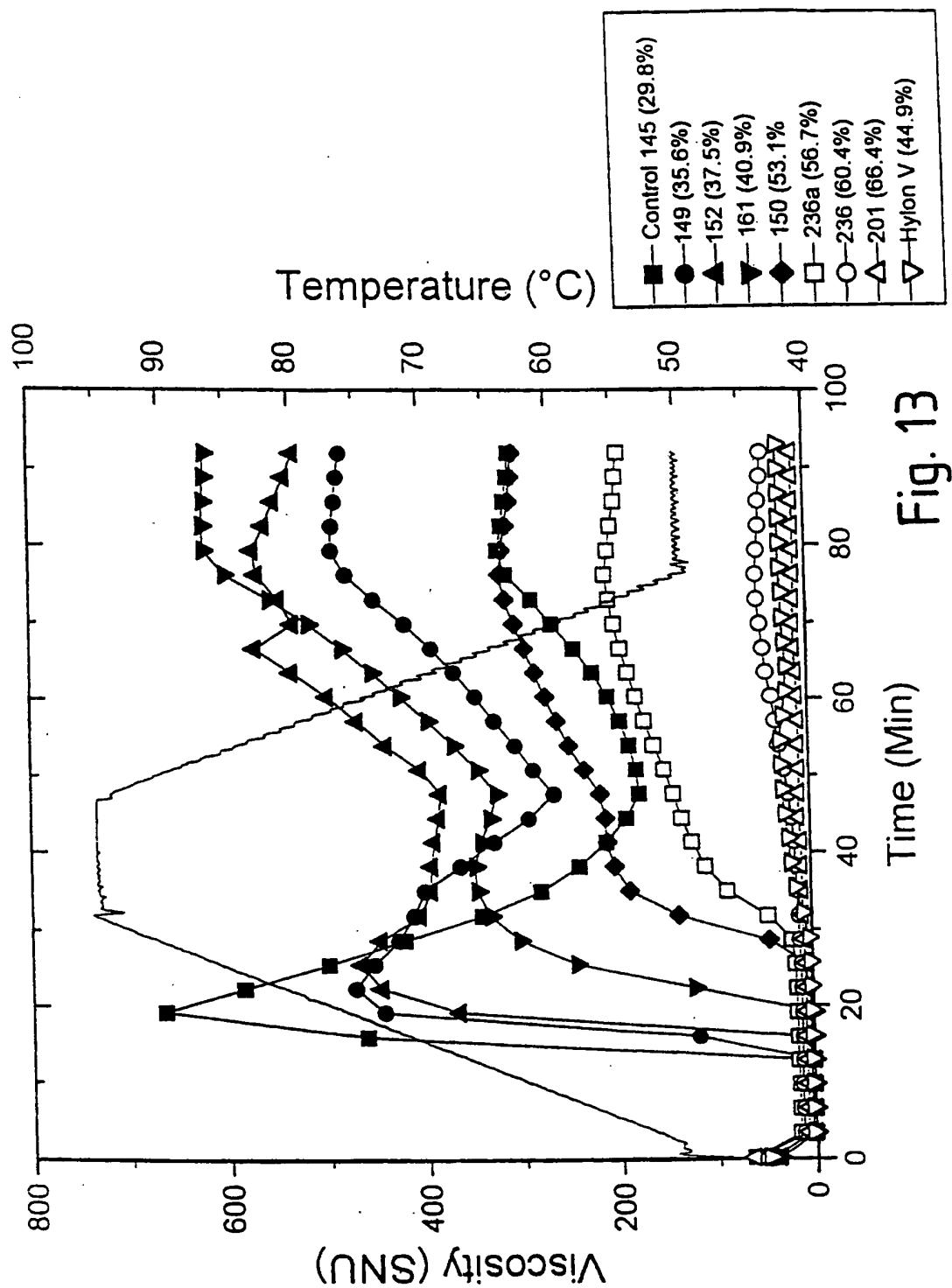
SUBSTITUTE SHEET (RULE 26)

CTTGTAGAACAGCAGTGGTCTATGCCACTAGTAGACAAGAAGAAGAAGAAGAAG 2520
 GAACATCTTGTGTCACAGATAACGTGATCATCTGTTCTTCTTCTTCTTCTTC
 P C R T A V V Y A L V D K E E E E E E E

AAGAAGTAGCAGTAGTAGAAGTAGTAGTAGAAGAAGAATGAACGAACCTTG
 TTCTTCTTCATCGTCATCATCTTCTTCATCATCTTACTTTGCTTGAACAC 2578
 E E E V A V V E E V V V E E E

Fig 12
SHEET 10

75/75

Fig. 13
Time (Min)